

**APPLICATION
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FOR

**METHOD FOR IDENTIFYING ALZHEIMER'S DISEASE
THERAPEUTICS USING TRANSGENIC ANIMAL MODELS**

METHOD FOR IDENTIFYING ALZHEIMER'S DISEASE THERAPEUTICS USING TRANSGENIC ANIMAL MODELS

This application is a continuation-in-part of U.S. Application Serial No. 08/480,653, ^{filed June 17, 1995, now abandoned.}

Background of the Invention

Transgenic animal models of Alzheimer's disease are described along with a method of using the transgenic animal models to screen for therapeutics useful for the treatment of Alzheimer's disease.

Alzheimer's disease (AD) is a degenerative disorder of the brain first described by Alois Alzheimer in 1907 after examining one of his patients who suffered drastic reduction in cognitive abilities and had generalized dementia (*The early story of Alzheimer's Disease*, edited by Bick *et al.* (Raven Press, New York 1987)). It is the leading cause of dementia in elderly persons. AD patients have increased problems with memory loss and intellectual functions which progress to the point where they cannot function as normal individuals. With the loss of intellectual skills the patients exhibit personality changes, socially inappropriate actions and schizophrenia (*A Guide to the Understanding of Alzheimer's Disease and Related Disorders*, edited by Jorm (New York University Press, New York 1987). AD is devastating for both victims and their families, for there is no effective palliative or preventive treatment for the inevitable neurodegeneration.

The impact of AD on society and on the national economy is enormous. It is expected that the demented elderly population in the United States will increase by 41% by the year 2000. It is expensive for the health care systems that must provide institutional and ancillary care for the AD patients at an estimated annual cost of \$40 billion (Jorm (1987); Fisher, "Alzheimer's Disease", New York Times, August 23, 1989, page D1, edited

by Reisberg (The Free Press, New York & London 1983)). These factors imply action must be taken to generate effective treatments for AD.

At a macroscopic level, the brains of AD patients are usually smaller, sometimes weighing less than 1,000 grams. At a microscopic level, the

5 histopathological hallmarks of AD include neurofibrillary tangles (NFT), neuritic plaques, and degeneration of neurons. AD patients exhibit degeneration of nerve cells in the frontal and temporal cortex of the cerebral cortex, pyramidal neurons of hippocampus, neurons in the medial, medial central, and cortical nuclei of the amygdala, noradrenergic neurons in the

10 locus coeruleus, and the neurons in the basal forebrain cholinergic system. Loss of neurons in the cholinergic system leads to a consistent deficit in cholinergic presynaptic markers in AD (Fisher (1983); *Alzheimer's Disease and Related Disorders, Research and Development* edited by Kelly (Charles C. Thomas, Springfield, IL. 1984)). In fact, AD is defined by the

15 neuropathology of the brain.

AD is associated with neuritic plaques measuring up to 200 μm in diameter in the cortex, hippocampus, subiculum, hippocampal gyrus, and amygdala. One of the principal constituents of neuritic plaques is amyloid, which is stained by Congo Red (Fisher (1983); Kelly (1984)). Amyloid

20 plaques stained by Congo Red are extracellular, pink or rust-colored in bright field, and birefringent in polarized light. The plaques are composed of polypeptide fibrils and are often present around blood vessels, reducing blood supply to various neurons in the brain.

Various factors such as genetic predisposition, infectious agents,

25 toxins, metals, and head trauma have all been suggested as possible mechanisms of AD neuropathy. However, available evidence strongly indicates that there are distinct types of genetic predispositions for AD. First, molecular analysis has provided evidence for mutations in the amyloid precursor protein (APP) gene in certain AD-stricken families (Goate *et al.*

Nature 349:704-706 (1991); Murrell *et al.* *Science* 254:97-99 (1991); Chartier-Harlin *et al.* *Nature* 353:844-846 (1991); Mullan *et al.*, *Nature Genet.* 1:345-347 (1992)). Additional genes for dominant forms of early onset AD reside on chromosome 14 and chromosome 1 (Rogaev *et al.*,
 5 *Nature* 376:775-778 (1995); Levy-Lahad *et al.*, *Science* 269:973-977 (1995); Sherrington *et al.*, *Nature* 375:754-760 (1995)). Another loci associated with AD resides on chromosome 19 and encodes a variant form of apolipoprotein E (Corder, *Science* 261:921-923 (1993)).

Amyloid plaques are abundantly present in AD patients and in Down's
 10 Syndrome individuals surviving to the age of 40. The overexpression of APP in Down's Syndrome is recognized as a possible cause of the development of AD in Down's patients over thirty years of age (Rumble *et al.*, *New England J. Med.* 320:1446-1452 (1989); Mann *et al.*, *Neurobiol. Aging* 10:397-399 (1989)). The plaques are also present in the normal aging
 15 brain, although at a lower number. These plaques are made up primarily of the amyloid β peptide ($A\beta$; sometimes also referred to in the literature as β -amyloid peptide or β peptide) (Glenner and Wong, *Biochem. Biophys. Res. Comm.* 120:885-890 (1984)), which is also the primary protein constituent in cerebrovascular amyloid deposits. The amyloid is a filamentous material that
 20 is arranged in beta-pleated sheets. $A\beta$ is a hydrophobic peptide comprising up to 43 amino acids. The determination of its amino acid sequence led to the cloning of the APP cDNA (Kang *et al.*, *Nature* 325:733-735 (1987); Goldgaber *et al.*, *Science* 235:877-880 (1987); Robakis *et al.*, *Proc. Natl. Acad. Sci.* 84:4190-4194 (1987); Tanzi *et al.*, *Nature* 331:528-530 (1988))
 25 and genomic APP DNA (Lemaire *et al.*, *Nucl. Acids Res.* 17:517-522 (1989); Yoshikai *et al.*, *Gene* 87, 257-263 (1990)). A number of forms of APP cDNA have been identified, including the three most abundant forms, APP695, APP751, and APP770. These forms arise from a single precursor

RNA by alternate splicing. The gene spans more than 175 kb with 18 exons (Yoshikai *et al.* (1990)). APP contains an extracellular domain, a transmembrane region and a cytoplasmic domain. A β consists of up to 28 amino acids just outside the hydrophobic transmembrane domain and up to 15 residues of this transmembrane domain. Thus, A β is a cleavage product derived from APP which is normally found in brain and other tissues such as heart, kidney and spleen. However, A β deposits are usually found in abundance only in the brain.

The larger alternate forms of APP (APP751, APP770) consist of APP695 plus one or two additional domains. APP751 consists of all 695 amino acids of APP695 plus an additional 56 amino acids which has homology to the Kunitz family of serine protease inhibitors (KPI) (Tanzi *et al.* (1988); Weidemann *et al.*, *Cell* 57:115-126 (1989); Kitaguchi *et al.*, *Nature* 331:530-532 (1988); Tanzi *et al.*, *Nature* 329:156 (1987)). APP770 contains all 751 amino acids of APP751 and an additional 19 amino acid domain homologous to the neuron cell surface antigen OX-2 (Weidemann *et al.* (1989); Kitaguchi *et al.* (1988)). Unless otherwise noted, the amino acid positions referred to herein are the positions as they appear in APP770. The amino acid number of equivalent positions in APP695 and APP751 differ in some cases due to the absence of the OX-2 and KPI domains. By convention, the amino acid positions of all forms of APP are referenced by the equivalent positions in the APP770 form. Unless otherwise noted, this convention is followed herein. Unless otherwise noted, all forms of APP and fragments of APP, including all forms of A β , referred to herein are based on the human APP amino acid sequence. APP is post-translationally modified by the removal of the leader sequence and by the addition of sulfate and sugar groups.

Van Broeckhaven *et al.*, *Science* 248:1120-1122 (1990), have demonstrated that the APP gene is tightly linked to hereditary cerebral hemorrhage with amyloidosis (HCHWA-D) in two Dutch families. This was confirmed by the finding of a point mutation in the APP coding region in two Dutch patients (Levy *et al.*, *Science* 248:1124-1128 (1990)). The mutation substituted a glutamine for glutamic acid at position 22 of the A β (position 618 of APP695, or position 693 of APP770). In addition, certain families are genetically predisposed to Alzheimer's disease, a condition referred to as familial Alzheimer's disease (FAD), through mutations resulting in an amino acid replacement at position 717 of the full length protein (Goate *et al.* (1991); Murrell *et al.* (1991); Chartier-Harlin *et al.* (1991)). These mutations co-segregate with the disease within the families and are absent in families with late-onset AD. This mutation at amino acid 717 increases the production of the A β ₁₋₄₂ form of A β from APP (Suzuki *et al.*, *Science* 264:1336-1340 (1994)). Another mutant form contains a change in amino acids at positions 670 and 671 of the full length protein (Mullan *et al.* (1992)). This mutation to amino acids 670 and 671 increases the production of total A β from APP (Citron *et al.*, *Nature* 360:622-674 (1992)).

There are no robust animal models to study AD, although aging nonhuman primates seem to develop amyloid plaques of A β in brain parenchyma and in the walls of some meningeal and cortical vessels. Although aged primates and canines can serve as animal models, they are expensive to maintain, need lengthy study periods, and are quite variable in the extent of pathology that develops.

There are no spontaneous animal mutations with sufficient similarities to AD to be useful as experimental models. Various models have been proposed in which some AD-like symptoms may be induced by electrolysis, transplantation of AD brain samples, aluminum chloride, kainic acid or

choline analogs (Kisner *et al.*, *Neurobiol. Aging* 7:287-292 (1986); Mistry *et al.*, *J Med Chem* 29:337-343 (1986)). Flood *et al.*, *Proc. Natl. Acad. Sci.* 88:3363-3366 (1986), reported amnesic effects in mice of four synthetic peptides homologous to the A β . Because none of these share with AD either
 5 common symptoms, biochemistry or pathogenesis, they are not likely to yield much useful information on etiology or treatment.

Several transgenic rodent lines have been produced that express either the human APP gene or human APP complementary DNA regulated by a variety of promoters. Transgenic mice with the human APP promoter linked
 10 to *E. coli* β -galactosidase (Wirak *et al.*, *The EMBO J* 10:289-296 (1991)) as well as transgenic mice expressing the human APP751 cDNA (Quon *et al.*, *Nature* 352:239-241 (1991)) or subfragments of the cDNA including the A β (Wirak *et al.*, *Science* 253:323-325 (1991); Sandhu *et al.*, *J. Biol. Chem.* 266:21331-21334 (1991); Kawabata *et al.*, *Nature* 354:476-478 (1991)) have
 15 been produced. Results obtained in the different studies appear to depend upon the source of promoter and the protein coding sequence used. For example, Wirak *et al.*, *Science* 253:323-325 (1991), found that in transgenic mice expressing a form of the A β , intracellular accumulation of "amyloid-like" material, reactive with antibodies prepared against A β were observed
 20 but did not find other histopathological disease symptoms. The intracellular nature of the antibody-reactive material and the lack of other symptoms suggest that this particular transgenic animal is not a faithful model system for Alzheimer's disease. Later studies have shown that similar staining is seen in non-transgenic control mice and Wirak *et al.*, *Science* 253:323-325
 25 (1991) was partially retracted in a comment in *Science* 255:143-145 (1992). Thus, the staining seen by Wirak *et al.* appears to be artifactual.

Kawabata *et al.* (1991) report the production of amyloid plaques, neurofibrillary tangles, and neuronal cell death in their transgenic animals.

In each of these studies, A β or a fragment containing A β was expressed. Wirak *et al.* (1991), used the human APP promoter while Kawabata *et al.* (1991) used the human thy-1 promoter. However, Kawabata *et al.* (1991) was later retracted by Kawabata *et al.*, *Nature* 356:23 (1992) and Kawabata *et al.*, *Nature* 356:265 (1992). In transgenic mice expressing the APP751 cDNA from the neuron-specific enolase promoter of Quon *et al.* (1991), rare, small extracellular deposits of material reactive with antibody prepared against synthetic A β were observed. A review of the papers describing these early transgenic mice indicate that do not produce characteristic Alzheimer pathologies (see Marx, *Science* 255:1200-1202 (1992)).

Transgenic mice expressing APP751 from a neuron-specific enolase (NSE) promoter were recently described by McConlogue *et al.*, *Neurobiol. Aging* 15:S12 (1994), Higgins *et al.*, *Ann Neurol.* 35:598-607 (1995), Mucke *et al.*, *Brain Res.* 666:151-167 (1994), Higgins *et al.*, *Proc. Natl. Acad. Sci. USA* 92:4402-4406 (1995), and U.S. Patent 5,387,742 to Cordell. Higgins *et al.*, *Ann Neurol.* 35:598-607 (1995) describe results with the same mice as described by Quon *et al.* (1991). Such mice have only sparse A β deposits which are more typical of very early AD and young Down's syndrome cases. The deposits seen in this transgenic mouse were also seen, although at a lower abundance, in non-transgenic control animals. Mature lesions such as frequent compacted plaques, neuritic dystrophy and extensive gliosis are not seen in these mice (Higgins *et al.*, *Ann Neurol.* 35:598-607 (1995)). McConlogue *et al.* (1994) reported finding no A β deposits in these mice.

Transgenic mice in which APP is expressed from the neuronal specific synaptophysin promoter express APP at low levels equivalent to that in brain tissue from the NSE APP mice described above. These mice were also reported not to display any brain lesions (Higgins *et al.*).

Transgenic mice containing yeast artificial chromosome (YAC) APP constructs have also been made (Pearson and Choi, *Proc. Natl. Acad. Sci. USA* 90:10578-10582 (1993); Lamb *et al.*, *Nature Genetics* 5:22-30 (1993); Buxbaum *et al.*, *Biochem. Biophys. Res. Comm.* 197:639-645 (1993)).

- 5 These mice contain the entire human APP genomic gene and express human APP protein at levels similar to endogenous APP; higher levels of expression than that obtained in mice using the NSE promoter. None of these mice, however, show evidence of pathology similar to AD.

- 10 Alzheimer's disease animal models, including transgenic models, have been recently reviewed by Lannfelt *et al.*, *Behavioural Brain Res.* 57:207-213 (1993), and Fukuchi *et al.*, *Ann. N.Y. Acad. Sci.* 695:217-223 (1993). Lannfelt *et al.* points out that none of the prior transgenic animals that show apparent plaques demonstrate neuropathological changes characteristic of AD. Lannfelt *et al.* also discusses possible reasons for the "failure" of previous
15 transgenic animal models. Similarly, Fukuchi *et al.* discusses the failure of prior transgenic animal models to display most of the characteristics known to be associated with AD. For example, the transgenic mouse reported by Quon *et al.* is reported to produce A β immunoreactive deposits that stain only infrequently with thioflavin S and not at all with Congo Red, in contrast
20 to the staining pattern of AD A β deposits.

- 25 Alzheimer's disease is characterized by numerous changes in the expression levels of various proteins, the biochemical activity and histopathology of brain tissue, as well as cognitive changes in affected individuals. Such characteristic changes associated with AD have been well documented. The most prominent change, as noted above, is the deposition of A β into amyloid plaques (Haass and Selkoe, *Cell* 75:1039-1042 (1993)). A variety of other molecules are also present in plaques, such as apolipoprotein E, laminin, amyloid P component, and collagen type IV

(Kalaria and Perry, *Brain Research* 631:151-155 (1993); Ueda *et al.*, *Proc. Natl. Aca. Sci. USA* 90:11282-11286 (1993)). Changes in cytoskeletal markers have also been associated with AD, such as the changes in microtubule-associated protein tau, MAP-2 or neurofilaments (Kosik *et al.*, *Science* 256: 780-783 (1992); Lovestone and Anderton, *Current Opinion in Neurology & Neurosurgery* 5:883-888 (1992); Brandan and Inestrosa, *General Pharmacology* 24:1063-1068 (1993); Trojanowski *et al.*, *Brain Pathology* 3:45-54 (1993); Masliah *et al.*, *American Journal of Pathology* 142:871-882 (1993)). Alzheimer's disease is also known to stimulate an immunoinflammatory response, increasing such inflammatory markers as glial fibrillary acidic protein (GFAP), α 2-macroglobulin, and interleukins 1 and 6 (IL-1 and IL-6) (Frederickson and Brunden, *Alzheimer Disease and Associated Disorders* 8:159-165 (1994); McGeer *et al.*, *Canadian Journal of Neurological Sciences* 18:376-379 (1991); Wood *et al.*, *Brain Research* 629:245-252 (1993)). Finally, neuronal and neurotransmitter changes have been associated with AD, such as the cholinergic, muscarinic, serotonergic, adrenergic, and adenosine receptor systems (Rylett *et al.*, *Brain Res* 289:169-175 (1983); Sims *et al.*, *Lancet* 1:333-336 (1980); Nitsch *et al.*, *Science* 258:304-307 (1992); Masliah and Terry, *Clinical Neuroscience* 1:192-198 (1993); Greenamyre and Maragos, *Cerebrovascular and Brain Metabolism Reviews* 5:61-94 (1993); McDonald and Nemeroff, *Psychiatric Clinics of North America* 14:421-422 (1991); Mohr *et al.*, *Journal of Psychiatry & Neuroscience* 19:17-23 (1994)).

It is therefore an object of the present invention to provide an animal model for Alzheimer's disease that is constructed using transgenic technology.

It is a further object of the present invention to provide transgenic animals characterized by certain genetic abnormalities in the expression of the amyloid precursor protein.

5 It is a further object of the present invention to provide transgenic animals exhibiting one or more histopathologies similar to those of Alzheimer's disease.

It is a further object of the present invention to provide transgenic animals expressing one or more $A\beta$ -containing proteins at high levels in brain tissue.

10 It is a further object of the present invention to provide a method of screening potential drugs for the treatment of Alzheimer's disease using transgenic animal models.

Summary of the Invention

15 The construction of transgenic animal models for testing potential treatments for Alzheimer's disease is described. The models are characterized by a greater similarity to the conditions existing in naturally occurring Alzheimer's disease, based on the ability to control expression of one or more of the three major forms of the β -amyloid precursor protein (APP), APP695, APP751, and APP770, or subfragments thereof, as well as various point mutations based on naturally occurring mutations, such as the FAD mutations at amino acid 717, and predicted mutations in the APP gene. The APP gene constructs are prepared using the naturally occurring APP
20 promoter of human, mouse, or rat origin, efficient promoters such as human platelet derived growth factor β chain (PDGF-B) gene promoter, as well as inducible promoters such as the mouse metallothionine promoter, which can be regulated by addition of heavy metals such as zinc to the animal's water
25

or diet. Neuron-specific expression of constructs can be achieved by using the rat neuron specific enolase promoter.

The constructs are introduced into animal embryos using standard techniques such as microinjection or embryonic stem cells. Cell culture
 5 based models can also be prepared by two methods. Cells can be isolated from the transgenic animals or prepared from established cell cultures using the same constructs with standard cell transfection techniques.

The constructs disclosed herein generally encode all or a contiguous portion of one of the three forms of APP: APP695, APP751, or APP770,
 10 preferably an A β -containing protein, as described herein. Examples of A β -containing proteins are proteins that include all or a contiguous portion of APP770, APP770 bearing a mutation in amino acid 669, 670, 671, 690, 692, and/or 717, APP751, APP751 bearing a mutation in amino acid 669, 670, 671, 690, 692, and/or 717, APP695, and APP695 bearing a mutation in
 15 amino acid 669, 670, 671, 690, 692, and/or 717, where each of these A β -containing proteins includes amino acids 672 to 714 of human APP. Some specific constructs that are described employ the following protein coding sequences: the APP770 cDNA; the APP770 cDNA bearing a mutation at amino acid 669, 670, 671, 690, 692, 717, or a combination of these
 20 mutations; the APP751 cDNA containing the KPI protease inhibitor domain without the OX-2 domain in the construct; the APP751 cDNA bearing a mutation at amino acid 669, 670, 671, 690, 692, 717, or a combination of these mutations; the APP695 cDNA; the APP695 cDNA bearing a mutation at amino acid 669, 670, 671, 690, 692, 717, or a combination of these
 25 mutations; APP695, APP751, or APP770 cDNA truncated at amino acid 671 or 685, the sites of β -secretase or α -secretase cleavage, respectfully; APP cDNA truncated to encode amino acids 646 to 770 of APP; APP cDNA truncated to encode amino acids 646 to 770 of APP and including at least one intron; the APP leader sequence followed by the A β region (amino acids 672

to 714 of APP) plus the remaining carboxy terminal 56 amino acids of APP;
the APP leader sequence followed by the A β region plus the remaining
carboxy terminal 56 amino acids with the addition of a mutation at amino
acid 717; the APP leader sequence followed by the A β region; the A β region
5 plus the remaining carboxy terminal 56 amino acids of APP; the A β region
plus the remaining carboxy terminal 56 amino acids of APP with the addition
of a mutation at amino acid 717; a combination cDNA/genomic APP gene
construct; a combination cDNA/genomic APP gene construct with the
addition of a mutation at amino acid 669, 670, 671, 690, 692, 717, or a
10 combination of these mutations; a combination cDNA/genomic APP gene
construct truncated at amino acid 671 or 685; and an APP cDNA construct
containing at least amino acids 672 to 722 of APP.

These protein coding sequences are operably linked to leader
sequences specifying the transport and secretion of the encoded A β related
15 protein. A preferred leader sequence is the APP leader sequence. These
combined protein coding sequences are in turn operably linked to a promoter
that causes high expression of A β in transgenic animal brain tissue. A
preferred promoter is the human platelet derived growth factor β chain
(PDGF-B) gene promoter. Additional constructs include a human yeast
20 artificial chromosome construct controlled by the PDGF-B promoter; a
human yeast artificial chromosome construct controlled by the PDGF-B
promoter with the addition of a mutation at amino acid 669, 670, 671, 690,
692, 717, or a combination of these mutations; the endogenous mouse or rat
APP gene modified through the process of homologous recombination
25 between the APP gene in a mouse or rat embryonic stem (ES) cell and a
vector carrying the human APP cDNA bearing a mutation at amino acid
position 669, 670, 671, 690, 692, 717, or a combination of these mutations,
such that sequences in the resident rodent chromosomal APP gene beyond the
recombination point (the preferred site for recombination is within APP exon

9) are replaced by the analogous human sequences bearing a mutation at amino acid 669, 670, 671, 690, 692, 717, or a combination of these mutations. These constructs can be introduced into the transgenic animals and then combined by mating of animals expressing the different constructs.

5 The transgenic animals, or animal cells, are used to screen for compounds altering the pathological course of Alzheimer's disease as measured by their effect on the amount and/or histopathology of Alzheimer's disease markers in the animals, as well as by behavioral alterations. These markers include APP and APP cleavage products; A β ; other plaque related
10 molecules such as apolipoprotein E, laminin, and collagen type IV; cytoskeletal markers, such as spectrin, tau, neurofilaments, and MAP-2; inflammatory markers, such as GFAP, α 2-macroglobulin, IL-1, and IL-6; and neuronal and synaptic neurotransmitter related markers, such as GAP43 and synaptophysin, and those associated with the cholinergic, muscarinic,
15 serotonergic, adrenergic, and adenosine receptor systems.

Brief Description of the Drawings

20 The boxed portions of the drawings indicate the amino acid coding portions of the constructs. Filled portions indicate the various domains of the protein as indicated in the Figure Legend. Lines indicate sequences in the clones that are 5' or 3' untranslated sequences, flanking genomic sequences, or introns. The break in the line to the left of the constructs in Figures ^{7a and 7b} 7 and ^{8a and 8b} 8 indicates the presence of a long DNA sequence.

25 Figure 1a is a schematic of the APP770 cDNA coding sequence.

Figure 1b is a schematic of the APP770 cDNA coding sequence bearing a mutation at position 717.

Figure 2a is a schematic of the APP751 cDNA coding sequence.

5 bearing a mutation at position 717.

Figure 3b is a schematic of the APP695 cDNA coding sequence

Figure 4b is a schematic of a coding sequence for the carboxy terminal portion of APP bearing a mutation at position 717.

Figure 6a is a schematic of a combination cDNA/genomic coding sequence allowing alternative splicing of the KPI and OX-2 exons.

Figure 7a is a schematic of a human APP YAC coding sequence.

Figure 7b is a schematic of a human APP YAC coding sequence bearing a mutation at position 717.

Figures 8a and 8b are schematics of genetic alteration of the mouse APP gene by homologous recombination between the mouse APP gene in a mouse ES cell and a vector carrying the human APP cDNA (either of the wild-type (Figure 8a) or FAD mutant form (Figure 8b)) directed to the exon 9 portion of the gene. As a result of this recombination event, sequences in the resident mouse chromosomal APP gene beyond the recombination point in exon 9 are replaced by the analogous human sequences.

Figure 9 is a schematic map of the PDAPP vector, a combination cDNA/genomic APP construct.

Figure 10 is a diagram of the genomic region of APP present in the PDAPP construct. The sizes of original introns 6, 7 and 8, as well as the sizes of the final introns are indicated on the diagram. The locations of the deletions in introns 6 and 8 present in the PDAPP construct are also indicated.

Figure 11 is a diagram of the intermediate constructs used to construct the APP splicing cassette and the PDAPP vector.

Figure 12 is a diagram of the PDAPP-wt vector and the plasmids used to make the PDAPP-wt vector.

Figure 13 is a diagram of the PDAPP-Sw/Ha vector and the plasmids and intermediate constructs used to make the PDAPP-Sw/Ha vector.

Figure 14 is a diagram of the PDAPP695_{V.F} vector and the plasmids and intermediate constructs used to make the PDAPP695_{V.F} vector.

Figures 15a and 15b are
~~Figure 15 is~~ a diagram of the PDAPP751_{V.F} vector and the plasmids and intermediate constructs used to make the PDAPP751_{V.F} vector.

Detailed Description of the Invention

The constructs and transgenic animals and animal cells are prepared using the methods and materials described below.

Sources of materials.

Restriction endonucleases are obtained from conventional commercial sources such as New England Biolabs (Beverly, MA.), Promega Biological Research Products (Madison, WI.), and Stratagene (La Jolla CA.). Radioactive materials are obtained from conventional commercial sources such as Dupont/NEN or Amersham. Custom-designed oligonucleotides for site-directed mutagenesis are available from any of several commercial providers of such materials such as Bio-Synthesis Inc., Lewisville, TX. Kits

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 868060-8724760

for carrying out site-directed mutagenesis are available from commercial suppliers such as Promega Biological Research Products and Stratagene. Clones of cDNA including the APP695, APP751, and APP770 forms of APP mRNA were obtained directly from Dr. Dmitry Goldgaber, NIH. Libraries
 5 of DNA are available from commercial providers such as Stratagene, La Jolla, CA., or Clontech, Palo Alto, CA. PC12 and 3T3 cells were obtained from ATCC (#CRL1721 and #CCL92, respectively). An additional PC12 cell line was obtained from Dr. Charles Marotta of Harvard Medical School, Massachusetts General Hospital, and McLean Hospital. Standard cell culture
 10 media appropriate to the cell line are obtained from conventional commercial sources such as Gibco/BRL. Murine stem cells, strain D3, were obtained from Dr. Rolf Kemler (Doetschman *et al.*, *J. Embryol. Exp. Morphol.* 87:27 (1985)). Lipofectin for DNA transfection and the drug G418 for selection of stable transformants are available from Gibco/BRL.

15 Definition of APP cDNA clones.

The cDNA clone APP695 is of the form of cDNA described by Kang *et al.*, *Nature* 325:733-735 (1987), and represents the most predominant form of APP in the brain. The cDNA clone APP751 is of the form described by
 20 Ponte *et al.*, *Nature* 331:525-527 (1988). This form contains an insert of 168 nucleotides relative to the APP695 cDNA. The 168 nucleotide insert encodes the KPI domain. The cDNA clone APP770 is of the form described by Kitaguchi *et al.* *Nature* 331:530-532 (1988). This form contains an insert of 225 nucleotides relative to the APP695 cDNA. This insert includes the
 25 168 nucleotides present in the insert of the APP751 cDNA, as well as an addition 57 nucleotide region that does not appear in APP751 cDNA. The 225 nucleotide insert encodes for the KPI domain as well as the OX-2 domain. All three forms arise from the same precursor RNA transcript by

alternative splicing. The 168 nucleotide insert is present in both APP751 cDNA and APP770 cDNA.

The sequence encoding APP751 is shown in SEQ ID NO:3. This sequence begins with the first base of the initiation codon AUG and encodes a 751 amino acid protein. Nucleotides 866 to 1033 of SEQ ID NO:3 do not appear in APP695 cDNA. The region from nucleotide 1957 to 2085 of SEQ ID NO:3 encodes the A β . The amino acid sequence of APP751 is shown in SEQ ID NO:4. Amino acids 289 to 345 of SEQ ID NO:4 do not appear in APP695. This 57 amino acid region includes the KPI domain. Amino acids 653 to 695 of SEQ ID NO:4 form the A β . These sequences are derived from Ponte *et al.* (1988).

region includes the OX-2 domain. Amino acids 672 to 714 form the A β . A probable membrane-spanning region of the APP occurs from amino acid 700 to 723. Unless otherwise stated, all references herein to nucleotide positions refer to the numbering of SEQ ID NO:5. This is the numbering derived from the APP770 cDNA. Unless otherwise stated, all references herein to amino acid positions refer to the numbering of SEQ ID NO:6. This is the numbering derived from APP770. According to this numbering convention, for example, amino acid position 717 refers to amino acid 717 of APP770, amino acid 698 of APP751, and amino acid 642 of APP695. The above sequences are derived from Kang *et al.* (1988) and Kitaguchi *et al.* (1988).

Unless otherwise noted, all forms of APP and fragments of APP, including all forms of A β , referred to herein are based on the human APP amino acid sequence. For example, A β refers to the human A β , APP refers to human APP, and APP770 refers to human APP770. As used herein, the term cDNA refers not only to DNA molecules actually prepared by reverse transcription of mRNA, but also any DNA molecule encoding a protein where the coding region is not interrupted, that is, a DNA molecule having a continuous open reading frame encoding a protein. As such, the term cDNA as used herein provides a convenient means of referring to a protein encoding DNA molecule where the protein encoding region is not interrupted by intron sequences (or any other sequences not encoding protein).

Definition of the APP genomic locus.

Characterization of phage and cosmid clones of human genomic DNA clones listed in Table 1 below originally established a minimum size of at least 100 kb for the Alzheimer's gene. There are a total of 18 exons in the APP gene (Lemaire *et al.*, *Nucl. Acid Res* 17:517-522 (1989); Yoshikai *et al.* (1990); Yoshikai *et al.*, *Nucleic Acids Res* 102:291-292 (1991)). Yoshikai *et*

al. (1990) describes the sequences of the exon-intron boundaries of the APP gene. These results taken together indicate that the minimum size of the Alzheimer's gene is 175 kb.

Table 1. Alzheimer's Cosmid and Lambda Clones.

Library	Name of Clone	Insert Size (kb)	Assigned APP Region
Cosmid	1 GPAPP47A	35	25 kb promoter & 9 kb intron 1
	2 GPAAP36A	35	12 kb promoter & 22 kb intron 1
	3 GAPP30A	30-35	5' coding region
	4 GAPP43A	30-35	exons 9, 10 and 11
Lambda	1 GAPP6A	12	exon 6
	2 GAPP6B	18	exons 4 and 5
	3 GAPP20A	20	exon 6
	4 GAPP20B	17	exons 4 and 5
	5 GAPP28A	18	exons 4 and 5
	6 GAPP3A	14	exon 6
	7 GAPP4A	19	exon 6
	8 GAPP10A	16	exons 9, 10 and 11
	9 GAPP16A	21	exon 6

Table 2 indicates where the 17 introns interrupt the APP coding sequence. The numbering refers to the nucleotide positions of APP770 cDNA as shown in SEQ ID NO:5. The starting nucleotide of exon 1 represents the first transcribed nucleotide. It is negative because the +1 nucleotide is the first nucleotide of the AUG initiator codon by convention (Kang *et al.* (1988)). The ending nucleotide of exon 18 represents the last nucleotide present in the mRNA prior to the poly(A) tail (Yoshikai *et al.* (1990)). It has been discovered that Yoshikai *et al.* (1990) and Yoshikai *et al.* (1991) contain an error in the location of exon 8. Figure 1 of Yoshikai *et al.* (1991) includes an EcoRI fragment between EcoRI fragments containing

exon 7 and exon 8. In fact, this intervening EcoRI fragment is actually located immediately after exon 8, so that the EcoRI fragment containing exon 7 and the EcoRI fragment containing exon 8 are adjacent to each other.

Table 2. Location of Introns in APP Gene Sequence.

	Starting nucleotide	Ending nucleotide	Following Intron
Exon 1	-146	57	Intron 1
Exon 2	58	225	Intron 2
Exon 3	226	355	Intron 3
Exon 4	356	468	Intron 4
Exon 5	469	662	Intron 5
Exon 6	663	865	Intron 6
Exon 7	866	1033	Intron 7
Exon 8	1034	1090	Intron 8
Exon 9	1091	1224	Intron 9
Exon 10	1225	1299	Intron 10
Exon 11	1300	1458	Intron 11
Exon 12	1459	1587	Intron 12
Exon 13	1588	1687	Intron 13
Exon 14	1688	1909	Intron 14
Exon 15	1910	1963	Intron 15
Exon 16	1964	2064	Intron 16
Exon 17	2065	2211	Intron 17
Exon 18	2212	3432	

APP Gene Mutations.

Certain families are genetically predisposed to Alzheimer's disease, a condition referred to as familial Alzheimer's disease (FAD), through mutations resulting in an amino acid replacement at position 717 of the full length protein (Goate *et al.* (1991); Murrell *et al.* (1991); Chartier-Harlin *et al.* (1991)). These mutations co-segregate with the disease within the families. For example, Murrell *et al.* (1991) described a specific mutation found in exon 17 (which Murrell *et al.* refers to as exon 15) where the valine of position 717 is replaced by phenylalanine.

Another FAD mutant form contains a change in amino acids at positions 670 and 671 of the full length protein (Mullan *et al.* (1992)). In one form of this mutation, the lysine at position 670 is replaced by asparagine and the methionine at position 671 is replaced by leucine. The effect of this mutation is to increase the production of A β in cultured cells approximately 7-fold (Citron *et al.*, *Nature* 360: 672-674 (1992); Lai *et al.*, *Science* 259:514-516 (1993)). Replacement of the methionine at position 671 with leucine by itself has also been shown to increase production of A β . Additional mutations in APP at amino acids 669, 670, and 671 have been shown to reduce the amount of A β processed from APP (Citron *et al.*, *Neuron* 14:661-670 (1995)). The APP construct with Val at amino acid 690 produces an increased amount of a truncated form of A β .

APP expression clones can be constructed that bear a mutation at amino acid 669, 670, 671, 690, 692, or 717 of the full length protein. The mutations from Lys to Asn and from Met to Leu at amino acids 670 and 671, respectively, are sometimes referred to as the Swedish mutation. Additional mutations can also be introduced at amino acids 669, 670, or 671 which either increase or reduce the amount of A β processed from APP. Mutations at these amino acids in any APP clone or transgene can be created by site-directed mutagenesis (Vincent *et al.*, *Genes & Devel.* 3:334-347 (1989)), or, once made, can be incorporated into other constructs using standard genetic engineering techniques. Some mutations at amino acid 717 are sometimes referred to as the Hardy mutation. Such mutations can include conversion of the wild-type Val717 codon to a codon for Ile, Phe, Gly, Tyr, Leu, Ala, Pro, Trp, Met, Ser, Thr, Asn, or Gln. A preferred substitution for Val717 is Phe. These mutations predispose individuals expressing the mutant proteins to develop Alzheimer's disease. It is believed that the mutations affect the expression and/or processing of APP, shifting

the balance toward Alzheimer's pathology. Mutations at amino acid 669 can include conversion of the wild-type Val669 codon to a codon for Trp, or deletion of the codon. Mutations at amino acid 670 can include conversion of the wild-type Lys670 codon to a codon for Asn or Glu, or deletion of the codon. Mutations at amino acid 671 can include conversion of the wild-type Met671 codon to a codon for Leu, Val, Lys, Tyr, Glu, or Ile, or deletion of the codon. A preferred substitution for Lys670 is Asn, and a preferred substitution for Met671 is Leu. These mutations predispose individuals expressing the mutant proteins to develop Alzheimer's disease. The other listed mutations to amino acids 669, 670, and 671 are known to reduce the amount of A β processed from APP (Citron *et al.* (1995)). It is believed that these mutations affect processing of APP leading to a change in A β production.

Truncated forms of APP can also be expressed from transgene constructs. For example, APP cDNA truncated to encode amino acids 646 to 770 of APP. The APP cDNA construct truncated to encode amino acids 646 to 770 of APP, and operatively linked to the PDGF-B promoter, is referred to as PDAPPc125.

20 **Nucleic Acid Constructs Encoding A β -containing Proteins.**

Constructs for use in transgenic animals include a promoter for expression of the construct in a mammalian cell and a region encoding a protein that includes all or a contiguous portion of one of the three forms of APP: APP695, APP751, or APP770, with or without specific amino acid mutations as described herein. It is preferred that protein encoded is an A β -containing protein. As used herein, an A β -containing protein is a protein that includes all or a contiguous portion of one of the three forms of APP: APP695, APP751, or APP770, with or without specific amino acid mutations as described herein, where the protein includes all or a portion of amino

acids 672 to 714 of human APP. Preferred A β -containing proteins include amino acids 672 to 714 of human APP. Preferred forms of such A β -containing proteins include all or a contiguous portion of APP770, APP770 bearing a mutation in amino acid 669, 670, 671, 690, 692, and/or 717, APP751, APP751 bearing a mutation in amino acid 669, 670, 671, 690, 692, and/or 717, APP695, and APP695 bearing a mutation in amino acid 669, 670, 671, 690, 692, and/or 717, where each of these A β -containing proteins includes amino acids 672 to 714 of human APP.

Preferred forms of the above A β -containing proteins are APP770; APP770 bearing a mutation in the codon encoding one or more amino acids selected from the group consisting of amino acid 669, 670, 671, 690, 692, 717; APP751; APP751 bearing a mutation in the codon encoding one or more amino acids selected from the group consisting of amino acid 669, 670, 671, 690, 692, 717; APP695; APP695 bearing a mutation in the codon encoding one or more amino acids selected from the group consisting of amino acid 669, 670, 671, 690, 692, 717; a protein consisting of amino acids 646 to 770 of APP; a protein consisting of amino acids 670 to 770 of APP; a protein consisting of amino acids 672 to 770 of APP; and a protein consisting of amino acids 672 to 714 of APP.

In the constructs disclosed herein, the DNA encoding the A β -containing protein can be cDNA or a cDNA/genomic DNA hybrid, wherein the cDNA/genomic DNA hybrid includes at least one APP intron sequence wherein the intron sequence is sufficient for splicing.

Preferred constructs contain DNA encoding APP770; DNA encoding APP770 bearing a mutation in the codon encoding amino acid 669, 670, 671, 690, 692, 717, or a combination of these mutations; a fragment of DNA encoding APP770 which encodes an amino acid sequence comprising amino acids 672 to 714 of APP770; DNA encoding APP751; DNA encoding APP751 bearing a mutation in the codon encoding amino acid 669, 670, 671,

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670, 671, 690, 692, 717, and a combination of these mutations; and a combination cDNA/genomic DNA hybrid APP gene construct truncated at amino acid 671 or 685.

5 Construction of Transgenes.

Construction of various APP transgenes can be accomplished using any suitable genetic engineering technique, such as those described in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (Cold Spring Harbor Laboratory, N.Y., 1989). Regions of APP clones that have been engineered or mutated can be interchanged by using convenient restriction enzyme sites present in APP cDNA clones. A *NruI* site starts at position -5 (relative to the first nucleotide of the AUG initiator codon). A *KpnI* and an *Asp718* site both start at position 57 (these are isoschizomers leaving different sticky ends). A *XcmI* site starts at position 836 and cuts at position 843. A *ScaI* site starts at position 1004. A *XhoI* site starts at position 1135. A *BamHI* site starts at position 1554. A *BglII* site starts at position 1994. An *EcoRI* site starts at position 2020. A *SpeI* site starts at position 2583. Another *EcoRI* site starts at position 3076.

The clones bearing various portions of the human APP gene sequence shown in Figures 1^a to 5 can be constructed in a common manner using standard genetic engineering techniques. For example, these clones can be constructed by first cloning the polyA addition signal from SV40 virus, as a 253 base pair *BclII* to *BamHI* fragment (Reddy *et al.*, *Science* 200:494-502 (1978), into a modified vector from the pUC series. Next, the cDNA coding sequences (APP770, APP751, or APP695) can be inserted. Correct orientation and content of the fragments inserted can be determined through restriction endonuclease mapping and limited sequencing. The clones bearing various carboxy terminal portions of the human APP gene sequence shown in

Figures 4a, 4b, and 5 can be constructed through several steps in addition to those indicated above. For example, an APP770 cDNA clone (SEQ ID NO:5) can be digested with *Asp718* which cleaves after nucleotide position 57. The resulting 5' extension is filled in using the Klenow enzyme (Sambrook *et al.* (1989)) and ligated to a hexanucleotide of the following sequence: AGATCT, the recognition site for *Bgl*II. After cleavage with *Bgl*II, which also cuts after position 1994, and re-ligation, the translational reading frame of the protein is preserved. The truncated protein thus encoded contains the leader sequence, followed by approximately 6 amino acids that precede the A β , followed by the A β , and the 56 terminal amino acids of APP. The clone in Figure 5 is created by converting the nucleotide at position 2138 to a T by site directed mutagenesis in the clone of Figure 4a, thus creating a termination codon directly following the last amino acid codon of the A β . APP cDNA clones naturally contain an *Nru*I site that cuts 2 nucleotides upstream from the initiator methionine codon. This site can be used for attachment of the different promoters used to complete each construct.

APP transgenes can also be constructed using PCR cloning techniques. Such techniques allow precise coupling of DNA fragments in the transgenes.

Combination cDNA/Genomic DNA Clones.

Endogenous APP expression results from transcription of precursor mRNA followed by alternative splicing to produce three main forms of APP. It is believed that this alternative splicing may be important in producing the pattern of APP expression involved in Alzheimer's disease. It is also believed that the presence of introns in expression constructs can influence the level and nature of expression by, for example, targeting precursor mRNA to mRNA processing and transport pathways (Huang *et al.*, *Nucleic Acids Res.* 18:937-947 (1990)). Accordingly, transgenes combining cDNA

and genomic DNA, which include intron sequences, are a preferred type of construct.

The RNA splicing mechanism requires only a few specific and well known consensus sequences. Such sequences have been identified in APP
 5 genomic DNA by Yoshikai *et al.* (1990). The disclosed transgenes can be constructed using one or more complete and intact intron sequences. However, it is preferred that the transgenes are constructed using truncated intron sequences that contain an effective amount of intron sequence to allow splicing. In general, truncated intron sequences that retain the splicing donor
 10 site, the splicing acceptor site, and the splicing branchpoint sequence will constitute an effective amount of an intron. The sufficiency of any truncated intron sequence can be determined by testing for the presence of correctly spliced mRNA in transgenic cells using methods described below.

Other intron sequences and splicing signals which are not derived
 15 from APP gene sequences may also be used in the transgene constructs. Such intron sequences will enhance expression of the transgene construct. A preferred heterologous intron is a hybrid between the adenovirus major late region first exon and intron junction and an IgG variable region splice acceptor. This hybrid intron can be constructed, for example, by joining the
 20 162 bp *PvuII* to *HindIII* fragment of the adenovirus major late region, containing 8 bp of the first exon and 145 bp of the first intron, and the 99 bp *HindIII* to *PstI* fragment of the IgG variable region splice acceptor clone-6, as described by Bothwell *et al.*, *Cell* 24:625-637 (1981). A similar splice signal has been shown to enhance expression of a construct to which it was
 25 attached, as described by Manley *et al.*, *Nucleic Acids Res.* 18:937-947 (1990). It is preferred that the heterologous intron be placed between the promoter and the region encoding the APP.

¹⁰³ A preferred APP combination cDNA/genomic expression clone includes an effective amount of introns 6, 7 and 8, as shown in Figure 6. ^{6a and 6b,}

Such a transgene can be constructed as follows. A preferred method of construction is described in Example 5. A plasmid containing the cDNA
 5 portion of the clone can be constructed by first converting the *TaqI* site at position 860 in an APP770 cDNA clone to an *XhoI* site by site-directed mutagenesis. Cleavage of the resulting plasmid with *XhoI* cuts at the new *XhoI* site and a pre-existing *XhoI* site at position 1135, and releases the KPI and OX-2 coding sequence. The plasmid thus generated serves as the
 10 acceptor for the KPI and OX-2 alternative splicing cassette.

The alternative splicing cassette can be created through a series of cloning steps involving genomic DNA. First, the *TaqI* site at position 860 in a genomic clone containing exon 6 and the adjacent downstream intron can be converted to an *XhoI* site by site-directed mutagenesis. Cleavage of the
 15 resulting plasmid with *XhoI* cuts at the new *XhoI* site and an *XhoI* site within either intron 6 or 7. This fragment, containing a part of exon 6 and at least a part of adjacent intron 6, can then be cloned into the *XhoI* site in a plasmid vector. Second, a genomic clone containing exon 9 and the adjacent upstream genomic sequences is cleaved with *XhoI*, cleaving the clone at the
 20 *XhoI* site at position 1135 (position 910 using the numbering system of Kang *et al.* (1987)) and an *XhoI* site in either intron 7 or 8. This fragment, containing a part of exon 9 and at least a part of adjacent intron 8, can then be cloned into the *XhoI* site of another plasmid vector. These two
 25 exon/intron junction fragments can then be released from their respective plasmid vectors by cleavage with *XhoI* and either *BamHI* or *BglII*, and cloned together into the *XhoI* site of another plasmid vector. It is preferred that the exon/intron junction fragments be excised with *BamHI*. It is most preferable that *BamHI* sites are engineered in the intron portion of the

exon/intron junction fragments prior to their excision. This allows the elimination of lengthy extraneous intron sequences from the cDNA/genomic clone.

The *XhoI* fragment resulting from cloning the two exon/intron
 5 junction fragments together can be cleaved with either *Bam*HI or *Bgl*II, depending on which enzyme was used for excision step above, and the genomic 6.8 kb *Bam*HI segment, containing the KPI and OX-2 coding region along with their flanking intron sequences, can be inserted. This fragment was identified by Kitaguchi *et al.* (1988) using Southern blot analysis of
 10 *Bam*HI-digested lymphocyte DNA from one normal individual and eight Alzheimer's disease patients using a 212 bp *TaqI-AvaI* fragment, nucleotides 862 to 1,073, of APP770 cDNA as the hybridization probe. Genomic DNA clones containing the region of the 225 bp insert can be isolated, for example, from a human leukocyte DNA library using the 212 bp *TaqI-AvaI*
 15 fragment as a probe. In the genomic DNA, the 225 bp sequence is located in a 168 bp exon (exon 7) and a 57 bp exon (exon 8), separated by an intron of approximately 2.6 kb (intron 7), with both exons flanked by intron-exon consensus sequences. The exon 7 corresponds to nucleotides 866 to 1,033 of APP770, and the exon 8 to nucleotides 1,034 to 1,090. Exon 7 encodes the
 20 highly conserved region of the Kunitz-type protease inhibitor family domain.

After cleavage with *XhoI*, this alternative splicing cassette, containing both exon and intron sequences, can then be excised by cleavage with *XhoI* and inserted into the *XhoI* site of the modified APP770 cDNA plasmid (the acceptor plasmid) constructed above. These cloning steps generate a
 25 combination cDNA/genomic expression clone that allows cells in a transgenic animal to regulate the inclusion of the KPI and OX-2 domains by a natural alternative splicing mechanism. An analogous gene bearing a mutation at amino acid 669, 670, 671, 690, 692, 717, or a combination of these

mutations, can be constructed either directly by *in vitro* mutagenesis. A mutation to amino acid 717 can also be made by using the mutated form of APP770 cDNA described above to construct an acceptor plasmid.

5 Promoters.

Different promoter sequences can be used to control expression of nucleotide sequences encoding A β -containing proteins. The ability to regulate expression of the gene encoding an A β -containing protein in transgenic animals is believed to be useful in evaluating the roles of the different APP gene products in AD. The ability to regulate expression of the gene encoding an A β -containing protein in cultured cells is believed to be useful in evaluating expression and processing of the different A β -containing gene products and may provide the basis for cell cultured drug screens. A preferred promoter is the human platelet derived growth factor β (PDGF-B) chain gene promoter (Sasahara *et al.*, *Cell* 64:217-227 (1991)).

Preferred promoters for the disclosed APP constructs are those that, when operatively linked to the protein coding sequences, mediate expression of one or more of the following expression products to at least a specific level in brain tissue of a two to four month old animal transgenic for one of the disclosed APP constructs. The products and their expression levels are A β_{tot} to a level of at least 30 ng/g (6.8 pmoles/g) brain tissue and preferably at least 40 ng/g (9.12 pmoles/g) brain tissue, A β_{1-42} to a level of at least 8.5 ng/g (1.82 pmoles/g) brain tissue and preferably at least 11.5 ng/g (2.5 pmoles/g) brain tissue, full length APP (FLAPP) and APP α combined (FLAPP+APP α) to a level of at least 150 pmoles/g brain tissue, APP β to a level of at least 42 pmoles/g brain tissue, and mRNA encoding human A β -containing protein to a level at least twice that of mRNA encoding the endogenous APP of the transgenic animal. A β_{tot} is the total of all forms of A β . A β_{1-42} is a form of A β having amino acids 1 to 42 of A β (corresponding

to amino acids 672 to 714 of APP). FLAPP+APP α refers to APP forms containing the first 12 amino acids of the A β region (corresponding to amino acids 672 to 684 of APP). Thus, FLAPP+APP α represents a mixture of full length forms of APP and APP cleaved at the α -secretase site (Esch *et al.*,
 5 *Science* 248:1122-1124 (1990)). APP β is APP cleaved at the β -secretase site (Seubert *et al.*, *Nature* 361:260-263 (1993)).

It is intended that the levels of expression described above refer to amounts of expression product present and are not limited to the specific units of measure used above. Thus, an expression level can be measured, for
 10 example, in moles per gram of tissue, grams per grams of tissue, moles per volume of tissue, and in grams per volume of tissue. The equivalence of these units of measure to the measures listed above can be determined using known conversion methods..

The levels of expression described above need not occur in all brain
 15 tissues. Thus, a promoter is considered preferred if at least one of the levels of expression described above occurs in at least one type of brain tissue. Where expression is tissue-specific, it is understood that if the expression level is sufficient in the specific brain tissue, the promoter is considered preferred even though the expression level in brain tissue as a whole may
 20 not, and need not, reach a threshold level. It is preferred that this level of expression is observed in hippocampal and/or cortical brain tissue. The promoter can mediate expression of the above expression products to the levels described above either constitutively or by induction. Induction can be accomplished by, for example, administration of an activator molecule, by
 25 heat, or by expression of a protein activator of transcription for the promoter operatively linked to the gene encoding an A β -containing protein. Many inducible expression systems which would be suitable for this purpose are known to those of skill in the art.

It is preferred that, in making the above measurements, the brain tissue is prepared by the following method. A brain from a transgenic test animal is dissected and the tissue is kept on ice throughout the homogenization procedure except as noted. The brain tissue is homogenized in 10 volumes (w/v) of 5 M guanidine-HCl, 50 mM Tris-HCl, pH 8.5. The sample is then gently mixed for 2 to 4 hours at room temperature. Homogenates are then diluted 1:10 in cold casein buffer #1 (0.25% casein/phosphate buffered saline (PBS) 0.05% sodium azide, pH 7.4, 1X protease inhibitor cocktail) for a final 0.5 M guanidine concentration and kept on ice. 100X protease inhibitor cocktail is composed of 2 mg/ml aprotinin, 0.5 M EDTA, pH 8.0, 1 mg/ml leupeptin. Diluted homogenates are then spun in an Eppendorf microfuge at 14,000 rpm for 20 minutes at 4°C. If further dilutions are required, they can be made with cold guanidine buffer #2 (1 part guanidine buffer #1 to 9 parts casein buffer #1).

It is preferred that the following assay be used to identify preferred promoters for their ability to mediate expression of A β to the levels described above. Antibody 266 (Seubert *et al.*, *Nature* 359:325-327 (1992)) is dissolved at 10 μ g/ml in buffer (0.23 g/L NaH₂PO₄-H₂O, 26.2 g/L NaHPO₄-7H₂O, 1 g/L sodium azide adjusted to pH 7.4) and 100 μ l/well is coated onto 96-well immunoassay plates (Costar) and allowed to bind overnight. The plate is then aspirated and blocked for at least 1 hour with a 0.25% human serum albumin solution in 25 g/L sucrose, 10.8 g/L Na₂HPO₄-7H₂O, 1.0 g/L NaH₂PO₄-H₂O, 0.5 g/L sodium azide adjusted to pH 7.4. The 266 coated plate is then washed 1X with wash buffer (PBS/0.05% Tween 20) using a Skatron plate washer. 100 μ l/well of A β 1-40 standards and brain tissue samples are added to the plate in triplicate and incubated overnight at 4°C. A β 1-40 standards are made from 0.0156, 0.0312, 0.0625, 0.125, 0.250, 0.500, and 1.000 μ g/ml stocks in DMSO stored at -40° C as well as a DMSO only control for background determination. A β standards

consist of 1:100 dilution of each standard into guanidine buffer #3 (1 part BSA buffer to 9 parts guanidine buffer #1) followed by a 1:10 dilution into casein buffer #1 (Note: the final A β concentration range is 15.6 to 1000 pg/ml and the final guanidine concentration is 0.5 M). BSA buffer consists
 5 of 1% bovine serum albumin (BSA, immunoglobulin-free)/PBS/0.05% sodium azide. The plates and casein buffer #2 (0.25% casein/PBS/0.05% Tween 20/pH 7.4) are then brought to room temperature (RT). The plates are then washed 3X with wash buffer. Next, 100 μ l/well of 3D6-biotin at 0.5 μ g/ml in casein buffer #2 is added to each well and incubated at 1 hour
 10 at RT.

Monoclonal antibody 3D6 was raised against the synthetic peptide DAEFRGGC (SEQ ID NO:10) which was conjugated through the cysteine to sheep anti-mouse immunoglobulin. The antibody does not recognize secreted APP but does recognize species that begin at A β position 1 (Asp). For
 15 biotinylating 3D6, follow Pierce's NHS-Biotin protocol for labeling IgG (cat. #20217X) except use 100 mM sodium bicarbonate, pH 8.5 and 24 mg NHS-biotin per ml of DMSO.

The plates are then again washed 3X with wash buffer. Then, 100 μ l/well of horseradish peroxidase (HRP)-avidin (Vector Labs, cat. # A-2004) diluted 1:4000 in casein buffer #2 is added to each well and incubated for 1
 20 hour at RT. The plates are washed 4X with wash buffer and then 100 μ l/well of TMB substrate (Slow TMB-ELISA (Pierce cat. # 34024)) at RT is added to each well and incubated for 15 minutes at RT. Finally, 25 μ l/well of 2 N H₂SO₄ is added to each well to stop the enzymatic reaction, and the
 25 plate is read at 450 nm to 650 nm using the Molecular Devices Vmax reader.

It is preferred that the relative levels of mRNA encoding human A β -containing protein mRNA encoding the endogenous APP of the transgenic animal be measured in the manner described by Bordonaro *et al.*, *Biotechniques* 16:428-430 (1994), and Rockenstein *et al.*, *J. Biol. Chem.*

270:28257-28267 (1995). Preferred methods for measuring the expression level of $A\beta_{1-42}$, FLAPP+APP α , and APP β are described in Example 8.

Yeast Artificial Chromosomes

5 The constructs shown in ^{Figures 7a and 7b} Figure 7 can be constructed as follows. Large segments of human genomic DNA, when cloned into certain vectors, can be propagated as autonomously-replicating units in the yeast cell. Such vector-borne segments are referred to as yeast artificial chromosomes (YAC; Burke *et al. Science* 236:806 (1987)). A human YAC library is

10 commercially available (Clontech, Palo Alto, CA) with an average insert size of 250,000 base pairs (range of 180,000 to 500,000 base pairs). A YAC clone of the Alzheimer's gene can be directly isolated by screening the library with the human APP770 cDNA. The inclusion of all of the essential gene regions in the clone can be confirmed by PCR analysis.

15 The YAC-APP clone, shown in Figure 7a, can be established in embryonic stem (ES) cells by selecting for neomycin resistance encoded by the YAC vector. ES cells bearing the YAC-APP clone can be used to produce transgenic mice by established methods described below under "Transgenic Mice" and "Embryonic Stem Cell Methods". The YAC-APP

20 gene bearing a mutation at amino acid 717 (Figure 7b) can be produced through the generation of a YAC library using genomic DNA from a person affected by a mutation at amino acid 717. Such a clone can be identified and established in ES cells as described above.

25 Genetic Alteration of the Mouse APP Gene.

The nucleotide sequence homology between the human and murine Alzheimer's protein genes is approximately 85%. Within the $A\beta$ -coding region, there are three amino acid differences between the two sequences. Amino acids Lys 670, Met671, and Val717, which can be mutated to alter

APP processing, are conserved between mouse, rat, and man. Wild-type rodents do not develop Alzheimer's disease nor do they develop deposits or plaques in their central nervous system (CNS) analogous to those present in human Alzheimer's patients. Therefore, it is possible that the human but not the rodent form of A β is capable of causing disease. Homologous recombination (Capecchi, *Science* 244:1288-1292 (1989)) can be used to convert the mouse Alzheimer's gene *in situ* to a gene encoding the human A β by gene replacement. This recombination is directed to a site downstream from the KPI and OX-2 domains, for example, within exon 9, so that the natural alternative splicing mechanisms appropriate to all cells within the transgenic animal can be employed in expressing the final gene product.

Both wild-type (Figure 8a) and mutant (Figure 8b) forms of human cDNA can be used to produce transgenic models expressing either the wild-type or mutant forms of APP. The recombination vector can be constructed from a human APP cDNA (APP695, APP751, or APP770 form), either wild-type, mutant at amino acid 669, 670, 671, 690, 692, 717, or a combination of these mutations. Cleavage of the recombination vector, for example, at the *Xho*I site within exon 9, promotes homologous recombination within the directly adjacent sequences (Capecchi (1989)). The endogenous APP gene resulting from this event would be normal up to the point of recombination, within exon 9 in this example, and would consist of the human cDNA sequence thereafter.

Preparation of Constructs for Transfections and Microinjections.

DNA clones for microinjection are cleaved with enzymes appropriate for removing the bacterial plasmid sequences, such as *Sal*I and *Not*I, and the DNA fragments electrophoresed on 1% agarose gels in TBE buffer (Sambrook *et al.* (1989)). The DNA bands are visualized by staining with

ethidium bromide, and the band containing the APP expression sequences is excised. The excised band is then placed in dialysis bags containing 0.3 M sodium acetate, pH 7.0. DNA is electroeluted into the dialysis bags, extracted with phenol-chloroform (1:1), and precipitated by two volumes of ethanol. The DNA is redissolved in 1 ml of low salt buffer (0.2 M NaCl, 20 mM Tris, pH 7.4, and 1 mM EDTA) and purified on an Elutip-D™ column. The column is first primed with 3 ml of high salt buffer (1 M NaCl, 20 mM Tris, pH 7.4, and 1 mM EDTA) followed by washing with 5 ml of low salt buffer. The DNA solutions are passed through the column for three times to bind DNA to the column matrix. After one wash with 3 ml of low salt buffer, the DNA is eluted with 0.4 ml of high salt buffer and precipitated by two volumes of ethanol. DNA concentrations are measured by absorption at 260 nm in a UV spectrophotometer. For microinjection, DNA concentrations are adjusted to 3 µg/ml in 5 mM Tris, pH 7.4 and 0.1 mM EDTA. Other methods for purification of DNA for microinjection are also described in Hogan *et al.*, *Manipulating the Mouse Embryo* (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1986); in Palmiter *et al.*, *Nature* 300:611 (1982); in *The Qiagenologist, Application Protocols*, 3rd edition, published by Qiagen, Inc., Chatsworth, CA.; and in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989).

Construction of Transgenic Animals.

A. Animal Sources.

Animals suitable for transgenic experiments can be obtained from standard commercial sources such as Charles River (Wilmington, MA), Taconic (Germantown, NY), and Harlan Sprague Dawley (Indianapolis, IN). Many strains are suitable, but Swiss Webster (Taconic) female mice are

preferred for embryo retrieval and transfer. B6D2F₁ (Taconic) males can be used for mating and vasectomized Swiss Webster studs can be used to stimulate pseudopregnancy. Vasectomized mice and rats can be obtained from the supplier.

5 **B. Microinjection Procedures.**

The procedures for manipulation of the rodent embryo and for microinjection of DNA are described in detail in Hogan *et al.*, *Manipulating the Mouse Embryo* (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1986), the teachings of which are incorporated herein.

10 **C. Transgenic Mice.**

Female mice six weeks of age are induced to superovulate with a 5 IU injection (0.1 cc, ip) of pregnant mare serum gonadotropin (PMSG; Sigma) followed 48 hours later by a 5 IU injection (0.1 cc, ip) of human chorionic gonadotropin (hCG; Sigma). Females are placed with males immediately after hCG injection. Twenty-one hours after hCG injection, the mated females are sacrificed by CO₂ asphyxiation or cervical dislocation and embryos are recovered from excised oviducts and placed in Dulbecco's phosphate buffered saline with 0.5% bovine serum albumin (BSA; Sigma). Surrounding cumulus cells are removed with hyaluronidase (1 mg/ml).
15 Pronuclear embryos are then washed and placed in Earle's balanced salt solution containing 0.5% BSA (EBSS) in a 37.5°C incubator with a humidified atmosphere at 5% CO₂, 95% air until the time of injection. Embryos can be implanted at the two cell stage.

Randomly cycling adult female mice are paired with vasectomized males. Swiss Webster or other comparable strains can be used for this purpose. Recipient females are mated at the same time as donor females. At the time of embryo transfer, the recipient females are anesthetized with an intraperitoneal injection of 0.015 ml of 2.5% avertin per gram of body weight. The oviducts are exposed by a single midline dorsal incision. An
25

incision is then made through the body wall directly over the oviduct. The ovarian bursa is then torn with watchmakers forceps. Embryos to be transferred are placed in DPBS (Dulbecco's phosphate buffered saline) and in the tip of a transfer pipet (about 10 to 12 embryos). The pipet tip is inserted into the infundibulum and the embryos transferred. After the transfer, the incision is closed by two sutures.

D. Transgenic Rats.

The procedure for generating transgenic rats is similar to that of mice (Hammer *et al.*, *Cell* 63:1099-112 (1990)). Thirty day-old female rats are given a subcutaneous injection of 20 IU of PMSG (0.1 cc) and 48 hours later each female placed with a proven male. At the same time, 40-80 day old females are placed in cages with vasectomized males. These will provide the foster mothers for embryo transfer. The next morning females are checked for vaginal plugs. Females who have mated with vasectomized males are held aside until the time of transfer. Donor females that have mated are sacrificed (CO₂ asphyxiation) and their oviducts removed, placed in DPBS (Dulbecco's phosphate buffered saline) with 0.5% BSA and the embryos collected. Cumulus cells surrounding the embryos are removed with hyaluronidase (1 mg/ml). The embryos are then washed and placed in EBSS (Earle's balanced salt solution) containing 0.5% BSA in a 37.5°C incubator until the time of microinjection.

Once the embryos are injected, the live embryos are moved to DPBS for transfer into foster mothers. The foster mothers are anesthetized with ketamine (40 mg/kg, ip) and xylazine (5 mg/kg, ip). A dorsal midline incision is made through the skin and the ovary and oviduct are exposed by an incision through the muscle layer directly over the ovary. The ovarian bursa is torn, the embryos are picked up into the transfer pipet, and the tip of the transfer pipet is inserted into the infundibulum. Approximately 10 to 12 embryos are transferred into each rat oviduct through the infundibulum. The

incision is then closed with sutures, and the foster mothers are housed singly.

E. Embryonic Stem (ES) Cell Methods.

1. Introduction of cDNA into ES cells.

Methods for the culturing of ES cells and the subsequent production
 5 of transgenic animals, the introduction of DNA into ES cells by a variety of
 methods such as electroporation, calcium phosphate/DNA precipitation, and
 direct injection are described in detail in *Teratocarcinomas and Embryonic
 Stem Cells, A Practical Approach*, ed. E.J. Robertson, (IRL Press 1987), the
 teachings of which are incorporated herein. Selection of the desired clone of
 10 transgene-containing ES cells can be accomplished through one of several
 means. For random gene integration, an APP clone is co-precipitated with a
 gene encoding neomycin resistance. Transfection is carried out by one of
 several methods described in detail in Lovell-Badge, in *Teratocarcinomas and
 Embryonic Stem Cells, A Practical Approach*, ed. E.J. Robertson, (IRL Press
 15 1987), or in Potter *et al.*, *Proc. Natl. Acad. Sci. USA* 81:7161 (1984).
 Lipofection can be performed using reagents such as provided in
 commercially available kits, for example DOTAP (Boehringer-Mannheim) or
 lipofectin (BRL). Calcium phosphate/DNA precipitation, lipofection, direct
 injection, and electroporation are the preferred methods. In these
 20 procedures, 0.5×10^6 ES cells are plated into tissue culture dishes and
 transfected with a mixture of the linearized APP clone and 1 mg of pSV2neo
 DNA (Southern and Berg, *J. Mol. Appl. Gen.* 1:327-341 (1982)) precipitated
 in the presence of 50 mg lipofectin (BRL) in a final volume of 100 μ l. The
 cells are fed with selection medium containing 10% fetal bovine serum in
 25 DMEM supplemented with G418 (between 200 and 500 μ g/ml). Colonies of
 cells resistant to G418 are isolated using cloning rings and expanded. DNA
 is extracted from drug resistant clones and Southern blots using an APP770
 cDNA probe can be used to identify those clones carrying the APP

sequences. PCR detection methods may also used to identify the clones of interest.

DNA molecules introduced into ES cells can also be integrated into the chromosome through the process of homologous recombination, described by Capecchi (1989). Direct injection results in a high efficiency of integration. Desired clones can be identified through PCR of DNA prepared from pools of injected ES cells. Positive cells within the pools can be identified by PCR subsequent to cell cloning (Zimmer and Gruss, *Nature* 338:150-153 (1989). DNA introduction by electroporation is less efficient and requires a selection step. Methods for positive selection of the recombination event (for example, neo resistance) and dual positive-negative selection (for example, neo resistance and gancyclovir resistance) and the subsequent identification of the desired clones by PCR have been described by Joyner *et al.*, *Nature* 338:153-156 (1989), and Capecchi (1989), the teachings of which are incorporated herein.

2. Embryo Recovery and ES cell Injection.

Naturally cycling or superovulated female mice mated with males can be used to harvest embryos for the implantation of ES cells. It is desirable to use the C57BL/6 strain for this purpose when using mice. Embryos of the appropriate age are recovered approximately 3.5 days after successful mating. Mated females are sacrificed by CO₂ asphyxiation or cervical dislocation and embryos are flushed from excised uterine horns and placed in Dulbecco's modified essential medium plus 10% calf serum for injection with ES cells. Approximately 10 to 20 ES cells are injected into blastocysts using a glass microneedle with an internal diameter of approximately 20 μ m.

3. Transfer of Embryos to Pseudopregnant Females.

Randomly cycling adult female mice are paired with vasectomized males. Mouse strains such as Swiss Webster, ICR or others can be used for this purpose. Recipient females are mated such that they will be at 2.5

3.5 days post-mating when required for implantation with blastocysts containing ES cells. At the time of embryo transfer, the recipient females are anesthetized with an intraperitoneal injection of 0.015 ml of 2.5% avertin per gram of body weight. The ovaries are exposed by making an incision in the body wall directly over the oviduct and the ovary and uterus are externalized. A hole is made in the uterine horn with a 25 gauge needle through which the blastocysts are transferred. After the transfer, the ovary and uterus are pushed back into the body and the incision is closed by two sutures. This procedure is repeated on the opposite side if additional transfers are to be made.

Identification, Characterization, and Utilization of Transgenic Mice and Rats.

Transgenic rodents can be identified by analyzing their DNA. For this purpose, tail samples (1 to 2 cm) can be removed from three week old animals. DNA from these or other samples can then be prepared and analyzed by Southern blot, PCR, or slot blot to detect transgenic founder (F_0) animals and their progeny (F_1 and F_2).

A. Pathological Studies.

The various F_0 , F_1 , and F_2 animals that carry a transgene can be analyzed by immunohistology for evidence of $A\beta$ deposition, expression of APP or APP cleavage products, neuronal or neuritic abnormalities, and inflammatory responses in the brain. Brains of mice and rats from each transgenic line are fixed and then sectioned. Sections are stained with antibodies reactive with the APP and/or the $A\beta$. Secondary antibodies conjugated with fluorescein, rhodamine, horse radish peroxidase, or alkaline phosphatase are used to detect the primary antibody. These methods permit identification of amyloid plaques and other pathological lesions in specific areas of the brain. Plaques ranging in size from 9 to $>50 \mu\text{m}$

characteristically occur in the brains of AD patients in the cerebral cortex, but also may be observed in deeper grey matter including the amygdaloid nucleus, corpus striatum and diencephalon. Sections can also be stained with other antibodies diagnostic of Alzheimer's plaques, recognizing antigens such as APP, Alz-50, tau, A2B5, neurofilaments, synaptophysin, MAP-2, ubiquitin, complement, neuron-specific enolase, and others that are characteristic of Alzheimer's pathology (Wolozin *et al.*, *Science* 232:648 (1986); Hardy and Allsop, *Trends in Pharm. Sci.* 12:383-388 (1991); Selkoe, *Ann. Rev. Neurosci.* 12:463-490 (1989); Arai *et al.*, *Proc. Natl. Acad. Sci. USA* 87:2249-2253 (1990); Majocha *et al.*, *Amer. Assoc. Neuropathology Abs* 99:22 (1988); Masters *et al.*, *Proc. Natl. Acad. Sci.* 82:4245-4249 (1985); Majocha *et al.*, *Can J Biochem Cell Biol* 63:577-584 (1985)). Staining with thioflavin S and Congo Red can also be carried out to analyze the presence of amyloid and co-localization of A β deposits within neuritic plaques and NFTs.

B. Analysis of APP and A β Expression.

1. mRNA.

Messenger RNA can be isolated by the acid guanidinium thiocyanate-phenol:chloroform extraction method (Chomaczynski and Sacchi, *Anal Biochem* 162:156-159 (1987)) from cell lines and tissues of transgenic animals to determine expression levels by Northern blots, RNase and nuclease protection assays.

2. Protein.

APP, A β , and other fragments of APP can and have been detected by using polyclonal and monoclonal antibodies that are specific to the APP extra-cytoplasmic domain, A β region, A β ₁₋₄₂, A β ₁₋₄₀, APP β , FLAPP+APP α , and C-terminus of APP. A variety of antibodies that are human sequence

specific, such as 10D5 and 6C6, are very useful for this purpose (Games *et al.* (1995)).

3. Western Blot Analysis.

Protein fractions can be isolated from tissue homogenates and cell
 5 lysates and subjected to Western blot analysis as described by, for example, Harlow *et al.*, *Antibodies: A Laboratory Manual*, (Cold Spring Harbor, NY, 1988); Brown *et al.*, *J. Neurochem.* 40:299-308 (1983); and Tate-Ostroff *et al.*, *Proc Natl Acad Sci* 86:745-749 (1989).

Briefly, the protein fractions are denatured in Laemmli sample buffer
 10 and electrophoresed on SDS-Polyacrylamide gels. The proteins are then transferred to nitrocellulose filters by electroblotting. The filters are blocked, incubated with primary antibodies, and finally reacted with enzyme conjugated secondary antibodies. Subsequent incubation with the appropriate chromogenic substrate reveals the position of APP derived proteins.

15 C. Pathological and Behavioral Studies.

1. Pathological Studies.

Immunohistology and thioflavin S staining are conducted as described elsewhere herein.

***In situ* Hybridizations:** Radioactive or enzymatically labeled nucleic
 20 acid probes can be used to detect mRNA *in situ*. The probes are degraded or prepared to be approximately 100 nucleotides in length for better penetration of cells. The hybridization procedure of Chou *et al.*, *J. Psych. Res.* 24:27-50 (1990), for fixed and paraffin embedded samples is briefly described below although similar procedures can be employed with samples sectioned
 25 as frozen material. Paraffin slides for *in situ* hybridization are dewaxed in xylene and rehydrated in a graded series of ethanols and finally rinsed in phosphate buffered saline (PBS). The sections are post-fixed in fresh 4% paraformaldehyde. The slides are washed with PBS twice for 5 minutes to

remove paraformaldehyde. Then the sections are permeabilized by treatment with a 20 $\mu\text{g/ml}$ proteinase K solution. The sections are re-fixed in 4% paraformaldehyde, and basic molecules that could give rise to background probe binding are acetylated in a 0.1 M triethanolamine, 0.3 M acetic anhydride solution for 10 minutes. The slides are washed in PBS, then dehydrated in a graded series of ethanols and air dried. Sections are hybridized with antisense probe, using sense probe as a control. After appropriate washing, bound radioactive probes are detected by autoradiography or enzymatically labeled probes are detected through reaction with the appropriate chromogenic substrates.

2. Behavioral Studies.

Behavioral tests designed to assess learning and memory deficits are employed. An example of such as test is the Morris water maze (Morris, *Learn Motiv.* 12:239-260 (1981)). In this procedure, the animal is placed in a circular pool filled with water, with an escape platform submerged just below the surface of the water. A visible marker is placed on the platform so that the animal can find it by navigating toward a proximal visual cue. Alternatively, a more complex form of the test in which there are no local cues to mark the platform's location will be given to the animals. In this form, the animal must learn the platform's location relative to distal visual cues, and can be used to assess both reference and working memory. A learning deficit in the water maze has been demonstrated with PDAPP transgenic mice. An example of behavioral analysis for assessing the effect of transgenic expression of $A\beta$ -containing proteins is described in Example 9.

Operant behavior studies of memory function: Memory function of the disclosed transgenic animals can be assessed by testing memory-related feeding behavior (Dunnett, "Operant delayed matching and non-matching position in rats" in *Behavioral Neuroscience, Volume I: A Practical Approach* (Sagal, ed., IRL Press, N.Y., 1993) pages 123-136; Zornetzen,

Behav. Neur. Biol. 36:49-60 (1982)). Transgenic and non-transgenic mice, are trained to earn food rewards in a two component operant procedure. One component features a delayed spatial alternation schedule. Under this schedule, the mouse must remember over a variable time delay which lever it has pressed in the previous trial so that it can earn a reward by pressing the alternate lever on the current trial. This provides a measure of the animal's recent or "working" memory. The second component features a discrimination spatial alternation schedule. Under this schedule, the mouse earns a reward by pressing whatever lever is illuminated. This discrimination behavior is an example of reference memory. These two groups of mice, transgenic and non-transgenic, can be chronically studied over time, for example, from 3 months of age until the end of their useful life span, in order to assess the development of sensitivity to cholinergic antagonists and behavioral impairment on these memory tasks. It is expected that the disclosed transgenic mice will model the cognitive deficits of Alzheimer's disease with enhanced sensitivity to the memory-disrupting effects of cholinergic antagonists and impairment on "working" and reference memory tasks. Dose-response challenges with the cholinergic antagonist can be conducted at various ages. These memory behavioral tests can also be used to compare the effect of compounds on the behavioral impairment of the disclosed transgenic animals. In this case, the two groups of mice are transgenic mice to which a test compound is administered and transgenic mice to which the compound is not administered.

Emotional reactivity and object recognition: Various functions of the disclosed transgenic animals can be assessed by testing locomotor activity, emotional reactivity to a novel environment or to novel objects, and object recognition. A first set of assessments are performed in the same animals at different ages (each animal is its own control) in order to test their performance in terms of locomotor activity, emotional reactivity to a novel

environment or to novel objects, and object recognition, a form of memory which is severely impaired in AD patients. On the first day, transgenic and non-transgenic control mice are individually placed in a square open field with a central platform. For 30 minutes, horizontal and vertical activity, and crossings of the platform, are recorded by blocks of 5 minutes for each animal. On the second day, each animal is submitted to two trials with an intertrial of 1 hour. On the first trial, two identical objects are placed in the open field and the animal is allowed 3 minutes of exploration. On the second trial, one of the objects is replaced by a new object and the time spent by the animal in exploring the familiar and novel object is recorded during the next 3 minutes (Ennaceur and Delacour, *Behav. Brain Res.* 31:47-59 (1988)). Animals are then tested for neophobic behavior, which is considered as an index of anxiety, in a free exploration situation, in which animals are given the opportunity to move freely between a familiar and a novel environment.

Thereafter, the same animals are submitted to various learning tasks to investigate their learning and memory capacities. They are first tested for spatial recognition memory in a T-maze delayed alternation task at 6 hour and 24 hour delays. This form of memory has been shown to be very sensitive to hippocampal damage. One half of the animals of each group is then trained in a positively reinforced lever-press task as described above. This can be used to measure post training improvement in performance of the animals, which has been shown to involve hippocampal activation. The other half of the animals is trained in spatial discrimination in an 8 arm radial maze (Olton and Samuelson, *J. Exp. Psychol. [Animal Behav.]* 2:97-116 (1976)) in order to evaluate working and reference memory and to analyze their strategies (angle preference), which give a better index of memory capacities. The animals trained and tested in the bar-lever press task at 2 to 3 months old can be trained and tested in radial maze at 9 to 10 months old,

and vice-versa. A working memory deficit has been demonstrated in PDAPP transgenic mice in the radial arm maze.

Two additional groups can also be submitted to the same behavioral tests as above at 9 to 10 months old in order to determine whether behavioral screening performed at 2 to 3 months old influenced further learning and memory capacities.

These memory behavioral tests can also be used to compare the effect of compounds on the behavioral impairment of the disclosed transgenic animals. In this case, the two groups of mice are transgenic mice to which a test compound is administered, and transgenic mice to which the compound is not administered.

The procedures applied to test transgenic mice are similar for transgenic rats.

D. Preferred Characteristics.

The above phenotypic characteristics of the disclosed transgenic animals can be used to identify those forms of the disclosed transgenic animals that are preferred as animal models. Additional phenotypic characteristics, and assays for measuring these characteristics, that can also be used to identify those forms of the disclosed transgenic animals that are preferred as animal models, are described in Example 6. These characteristics are preferably those that are similar to phenotypic characteristics observed in Alzheimer's disease. APP and A β markers which are also useful for identifying those forms of the disclosed transgenic animals that are preferred as animal models are described below. Any or all of the these markers or phenotypic characteristics can be used either alone or in combination to identify preferred forms of the disclosed transgenic animals. For example, the presence of plaques in brain tissue that can be stained with Congo red is a phenotypic characteristic which can identify a disclosed transgenic animal as preferred. It is intended that the levels of expression of

certain APP-related proteins present in preferred transgenic animals (discussed above) is an independent characteristic for identifying preferred transgenic animals. Thus, the most preferred transgenic animals will exhibit both a disclosed expression level for one or more of the APP-related proteins and one or more of the phenotypic characteristics discussed above.

Especially preferred phenotypic characteristics (the presence of which identifies the animal as a preferred transgenic animal) are the presence of amyloid plaques that can be stained with Congo Red (Kelly (1984)), the presence of extracellular amyloid fibrils as identified by electron microscopy by 12 months of age, and the presence of type I dystrophic neurites as identified by electron microscopy by 12 months of age (composed of spherical neurites that contain synaptic proteins and APP; Dickson *et al.*, *Am J Pathol* 132:86-101 (1988); Dickson *et al.*, *Acta Neuropath.* 79:486-493 (1990); Masliah *et al.*, *J Neuropathol Exp Neurol* 52:135-142 (1993); Masliah *et al.*, *Acta Neuropathol* 87:135-142 (1994); Wang and Munoz, *J Neuropathol Exp Neurol* 54:548-556 (1995)). Examples of the detection of these characteristics is provided in Example 6. It is most preferred that the transgenic animals have amyloid plaques that can be stained with Congo Red as of 14 months of age.

Screening of Compounds for Treatment of Alzheimer's Disease.

The transgenic animals, or animal cells derived from transgenic animals, can be used to screen compounds for a potential effect in the treatment of Alzheimer's disease using standard methodology. In such AD screening assays, the compound is administered to the animals, or introduced into the culture media of cells derived from these animals, over a period of time and in various dosages, then the animals or animal cells are examined for alterations in APP expression or processing, expression levels or

localization of other AD markers, histopathology, and/or, in the case of animals, behavior using the procedures described above and in the examples below. In general, any improvement in behavioral tests, alteration in AD-associated markers, reduction in the severity of AD-related histopathology, reduction in the expression of A β or APP cleavage products, and/or changes in the presence, absence or levels of other compounds that are correlated with AD which are observed in treated animals, relative to untreated animals, is indicative of a compound useful for treating Alzheimer's disease. The specific proteins, and the encoding transcripts, the enzymatic or biochemical activity, and/or histopathology of those proteins, that are associated with and characteristic of AD are referred to herein as markers. Expression or localization of these markers characteristic of AD has either been detected, or is expected to be present, in the disclosed transgenic animals. These markers can be measured or detected, and those measurements compared between treated and untreated transgenic animals to determine the effect of a tested compound.

Markers useful for AD screening assays are selected based on detectable changes in these markers that are associated with AD. Many such markers have been identified in AD and have either been detected in the disclosed transgenic animals or are expected to be present in these animals. These markers fall into several categories based on their nature, location, or function. Preferred examples of markers useful in AD screening assays are described below, group as A β -related markers, plaque-related markers, cytoskeletal and neuritic markers, inflammatory markers, and neuronal and neurotransmitter-related markers.

A. A β -related Markers.

Expression of the various forms of APP and A β can be directly measured and compared in treated and untreated transgenic animals both by immunohistochemistry and by quantitative ELISA measurements as described

above and in the examples. Currently, it is known that two forms of APP products are found, APP and A β (Haass and Selkoe, *Cell* 75:1039-1042 (1993)). They have been shown to be intrinsically associated with the pathology of AD in a time dependent manner. Therefore, preferred assays

5 compare age-related changes in APP and A β expression in the transgenic mice. As described in Example 6, increases in A β have been demonstrated during aging of the PDAPP mouse.

Preferred targets for assay measurement are A β markers known to increase in individuals with Alzheimer's disease are total A β (A β_{tot}), A β 1-42

10 (A β_{1-42} ; A β with amino acids 1-42), A β_{1-40} (A β with amino acids 1-40), A β N3(pE) (A $\beta_{\text{N3(pE)}}$); A β X-42 (A $\beta_{\text{X-42}}$; A β forms ending at amino acid 42); A β X-40 (A $\beta_{\text{X-40}}$; A β forms ending at amino acid 40); insoluble A β (A $\beta_{\text{Insoluble}}$); and soluble A β (A β_{Soluble} ; Kuo *et al.*, *J. Biol. Chem.* 271(8):4077-4081 (1996)). A $\beta_{\text{N3(pE)}}$ has pyroglutamic acid at position 3

15 (Saïdo, *Neuron* 14:457-466 (1995)). A $\beta_{\text{X-42}}$ refers to any of the C-terminal forms of A β such as A β_{13-42} . A $\beta_{\text{Insoluble}}$ refers to forms of A β that are recovered as described in Gravina, *J. Biol. Chem.* 270:7013-7016 (1995). APP β can also be specifically measured to assess the amount of β -secretase activity (Seubert *et al.*, *Nature* 361:260-263 (1993)). Several of these A β

20 forms and their association with Alzheimer's disease are described by Haass and Selkoe (1993). Detection and measurement of A β_{tot} , A β_{1-42} , and A $\beta_{\text{X-42}}$ are described in Example 6. Generally, specific forms of A β can be assayed, either quantitatively or qualitatively using specific antibodies, as described below. When referring to amino acid positions in forms of A β , the positions

25 correspond to the A β region of APP. Amino acid 1 of A β corresponds to amino acid 672 of APP, and amino acid 42 of A β corresponds to amino acid 714 of APP.

Also preferred as targets for assay measurement are APP markers. For example, different forms of secreted APP (termed APP α and APP β) can also be measured (Seubert *et al.*, *Nature* 361:260-263 (1993)). Other APP forms can also serve as targets for assays to assess the potential for compounds to affect Alzheimer's disease. These include FLAPP+APP α , full length APP, C-terminal fragments of APP, especially C100 (the last 100 amino acids of APP) and C57 to C60 (the last 57 to 60 amino acids of APP), and any forms of APP that include the region corresponding to A β ₁₋₄₀.

APP forms are also preferred targets for assays to assess the potential for compounds to affect Alzheimer's disease. The absolute level of APP and APP transcripts, the relative levels of the different APP forms and their cleavage products, and localization of APP expression or processing are all markers associated with Alzheimer's disease that can be used to measure the effect of treatment with potential therapeutic compounds. The localization of APP to plaques and neuritic tissue is an especially preferred target for these assays.

Quantitative measurement can be accomplished using many standard assays. For example, transcript levels can be measured using RT-PCR and hybridization methods including RNase protection, Northern analysis, and R-dot analysis. APP and A β levels can be assayed by ELISA, Western analysis, and by comparison of immunohistochemically stained tissue sections. Immunohistochemical staining can also be used to assay localization of APP and A β to particular tissues and cell types. Such assays were described above and specific examples are provided below.

B. Plaque-related Markers.

A variety of other molecules are also present in plaques of individuals with AD and in the disclosed transgenic animals, and their presence in plaques and neuritic tissue can be detected. The amount of these markers present in plaques or neuritic tissue is expected to increase with the age of

untreated transgenic animals. Preferred plaque-related markers are apolipoprotein E, glycosylation end products, amyloid P component, advanced glycosylation end products (Smith *et al.*, *Proc. Natl. Acad. Sci. USA* 91:5710 (1994)), growth inhibitory factor, laminin, collagen type IV
 5 (Kalaria and Perry (1993); Ueda *et al.* (1993)), receptor for advanced glycosylation products (RAGE), and ubiquitin.

While the above markers can be used to detect specific components of plaques and neuritic tissue, the location and extent of plaques can also be determined by using well known histochemical stains, such as Congo Red
 10 and thioflavin S, as described above and in some examples below.

C. Cytoskeletal and Neuritic Markers.

Many changes in cytoskeletal markers associated with AD have also been detected in transgenic PDAPP mice. These markers can be used in AD screening assays to determine the effect of compounds on AD. Many of the
 15 changes in cytoskeletal markers occur either in the neurofibrillary tangles or dystrophic neurites associated with plaques (Kosik *et al.* (1992); Lovestone and Anderton (1992); Brandan and Inestrosa (1993); Trojanowski *et al.* (1993); Masliah *et al.* (1993)).

The following are preferred cytoskeletal and neuritic markers that
 20 exhibit changes in and/or an association with AD. These markers can be detected, and changes can be determined, to measure the effect of compounds on the disclosed transgenic animals. Spectrin exhibits increased breakdown in AD. Tau and neurofilaments display an increase in hyperphosphorylation in AD, and levels of ubiquitin increase in AD. Tau, ubiquitin, MAP-2,
 25 neurofilaments, heparin sulfate, and chondroitin sulphate are localized to plaques and neuritic tissue in AD and in general change from the normal localization. GAP43 levels are decreased in the hippocampus and abnormally

phosphorylated tau and neurofilaments are present in PDAPP transgenic mice.

D. Inflammatory Markers.

Alzheimer's disease is also known to stimulate an

- 5 immunoinflammatory response, with a corresponding increase in inflammatory markers (Frederickson and Brunden (1994); McGeer *et al.* (1991); Wood *et al.* (1993)). The following are preferred inflammatory markers that exhibit changes in and/or an association with AD. Detection of changes in these markers are useful in AD screening assays. Acute phase
- 10 proteins and glial markers, such as α 1-antitrypsin, C-reactive protein, α 2-macroglobulin (Tooyama *et al.*, *Molecular & Chemical Neuropathology* 18:153-60 (1993)), glial fibrillary acidic protein (GFAP), Mac-1, F4/80, and cytokines, such as IL-1 α and β , TNF α , IL-8, MIP-1 α (Kim *et al.*, *J. Neuroimmunology* 56:127-134 (1995)), MCP-1 (Kim *et al.*, *J. Neurological*
- 15 *Sciences* 128:28-35 (1995); Kim *et al.*, *J. Neuroimmunology* 56:127-134 (1995); Wang *et al.*, *Stroke* 26:661-665 (1995)), and IL-6, all increase in AD and are expected to increase in the disclosed transgenic animals.
- Complement markers, such as C3d, C1q, C5, C4d, C4bp, and C5a-C9, are localized in plaques and neuritic tissue. Major histocompatibility complex
- 20 (MHC) glycoproteins, such as HLA-DR and HLA-A, D,C increase in AD. Microglial markers, such as CR3 receptor, MHC I, MHC II, CD 31, CD11a, CD11b, CD11c, CD68, CD45RO, CD45RD, CD18, CD59, CR4, CD45, CD64, and CD44 (Akiyama *et al.*, *Brain Research* 632:249-259 (1993)) increase in AD. Additional inflammatory markers useful in AD
- 25 screening assays include α 2 macroglobulin receptor, Fibroblast growth factor (Tooyama *et al.*, *Neuroscience Letters* 121:155-158 (1991)), ICAM-1 (Akiyama *et al.*, *Acta Neuropathologica* 85:628-634 (1993)), Lactotransferrin (Kawamata *et al.*, *American Journal of Pathology* 142:1574-85 (1993)), C1q,

- C3d, C4d, C5b-9, Fc gamma RI, Fc gamma RII, CD8 (McGeer *et al.*, *Can J Neurol Sci* 16:516-527 (1989)), LCA (CD45) (McGeer *et al.* (1989); Akiyama *et al.*, *Journal of Neuroimmunology* 50:195-201 (1994)), CD18 (beta-2 integrin) (Akiyama and McGeer, *Journal of Neuroimmunology* 30:81-93 (1990)), CD59 (McGeer *et al.*, *Brain Research* 544:315-319 (1991)), Vitronectin (McGeer *et al.*, *Canadian Journal of Neurological Sciences* 18:376-379 (1991); Akiyama *et al.*, *Journal of Neuroimmunology* 32:19-28 (1991)), Vitronectin receptor, Beta-3 integrin (Akiyama *et al.* (1991)), Apo J, clusterin (McGeer *et al.*, *Brain Research* 579:337-341 (1992)), type 2 plasminogen activator inhibitor (Akiyama *et al.*, *Neuroscience Letters* 164:233-235 (1993)), CD44 (Akiyama *et al.*, *Brain Research* 632:249-259 (1993)), Midkine (Yasuhara *et al.*, *Biochemical & Biophysical Research Communications* 192:246-251 (1993)), Macrophage colony stimulating factor receptor (Akiyama *et al.*, *Brain Research* 639:171-174 (1994)), MRP14, 27E10, and interferon-alpha (Akiyama *et al.*, *Journal of Neuroimmunology* 50:195-201 (1994)). Additional markers which are associated with inflammation or oxidative stress include 4-hydroxynonenal-protein conjugates (Uchida *et al.*, *Biochem. Biophys. Res. Comm.* 212:1068-1073 (1995); Uchida and Stadtman, *Methods in Enzymology* 233:371-380 (1994); Yoritaka *et al.*, *Proc. Natl. Acad. Sci. USA* 93:2696-2701 (1996)), I κ B, NF κ B (Kaltschmidt *et al.*, *Molecular Aspects of Medicine* 14:171-190 (1993)), cPLA₂ (Stephenson *et al.*, *Neurobiology Dis.* 3:51-63 (1996)), COX-2 (Chen *et al.*, *Neuroreport* 6:245-248 (1995)), Matrix metalloproteinases (Backstrom *et al.*, *J. Neurochemistry* 58:983-992 (1992); Bignami *et al.*, *Acta Neuropathologica* 87:308-312 (1994); Deb and Gottschall, *J. Neurochemistry* 66:1641-1647 (1995); Peress *et al.*, *J. Neuropathology & Experimental Neurology* 54:16-22 (1995)), Membrane lipid peroxidation, Protein oxidation (Hensley *et al.*, *J. Neurochemistry* 65:2146-2156 (1995); Smith *et al.*, *Proc.*

Natl. Acad. Sci. USA 88:10540-10543 (1991)), and diminished ATPase activity (Mark *et al.*, *J. Neuroscience* 15:6239 (1995)). These markers can be detected, and changes can be determined, to measure the effect of compounds on the disclosed transgenic animals.

5 **E. Neuronal and Neurotransmitter-related Markers.**

Changes in neuronal and neurotransmitter biochemistry have been associated with AD and in the disclosed PDAPP animals. In AD there is a profound reduction in cortical and hippocampal cholinergic innervation. This is evidenced by the dramatic loss of the synthetic enzyme choline
 10 acetyltransferase and decreased acetylcholinesterase, synaptosomal choline uptake (as measured by hemicholinium binding) and synthesis and release of acetylcholine (Rylett *et al.* (1983); Sims *et al.* (1980); Coyle *et al.*, *Science* 219:1184-1190 (1983); Davies and Maloney, *Lancet* 2:1403 (1976); Perry *et al.*, *Lancet* 1:189 (1977); Sims *et al.*, *J. Neurochem.* 40: 503-509 (1983)) all
 15 of which are useful markers. These markers can be used in AD screening assays to determine the effect of compounds on AD. There is also a loss of basal forebrain neurons and the galanin system becomes hypertrophic in AD.

In addition to changes in the markers described above in AD, there is also atrophy and loss of basal forebrain cholinergic neurons that project to
 20 the cortex and hippocampus (Whitehouse *et al.*, *Science* 215:1237-1239 (1982)), as well as alterations of entorhinal cortex neurons (Van Hoesan *et al.*, *Hippocampus* 1:1-8 (1991). Based upon these observations measurement of these enzyme activities, neuronal size, and neuronal count numbers are expected to decrease in the disclosed transgenic animals and are therefore
 25 useful targets for detection in AD screening assays. Basal forebrain neurons are dependent on nerve growth factor (NGF). Brain-derived neurotrophic factor (BDNF) may also decrease in the hippocampus in the disclosed

transgenic animals and is therefore a useful target for detection in AD screening assays.

It has also been shown that APP and A β release are affected by stimulation of muscarinic receptors both *in vitro* in tissue culture as well as in
 5 brain slices. Similar findings have also been obtained with application of other agonists linked to phosphoinositol turnover (Nitsch *et al.* (1992); Hung
et al., *J. Biol. Chem.* 268:22959-22962 (1993); Nitsch *et al.*, *Proceedings of the Eighth Meeting of the International Study Group on the Pharmacology of Memory Disorders Associated with Aging* 497-503 (1995); Masliah and Terry
 10 (1993); Greenamyre and Maragos (1993); McDonald and Nemeroff (1991); Mohr *et al.* (1994); Perry, *British Medical Bulletin* 42:63-69 (1986); Masliah
et al., *Brain Research* 574:312-316 (1992); Schwagerl *et al.*, *Journal of Neurochemistry* 64:443-446 (1995)). Based upon these observations, it is possible that neurotransmitter agonists will reduce the production of A β in
 15 the disclosed transgenic animals. Based on this reasoning, screening assays that measure the effect of compounds on neurotransmitter receptors can possibly be used to identifying compounds useful in treating AD.

In addition to the well-documented changes in the cholinergic system, dysfunction in other receptor systems such as the serotonergic, adrenergic,
 20 adenosine, and nicotine receptor systems, has also been documented. Markers characteristic of these changes, as well as other neuronal markers that exhibit both metabolic and structural changes in AD are listed below. Changes in the level and/or localization of these markers can be measured using similar techniques as those described for measuring and detecting the
 25 earlier markers.

The following are preferred cytoskeletal and neuritic markers that exhibit changes in and/or an association with AD. Levels of cathepsin (cat) D,B and Neuronal Thread Protein, and phosphorylation of elongation factor-

2, increase in AD. Cat D,B, protein kinase C, and NADPH are localized in plaque and neuritic tissue in AD. Activity and/or levels of nicotine receptors, 5-HT₂ receptor, NMDA receptor, α 2-adrenergic receptor, synaptophysin, p65, glutamine synthetase, glucose transporter, PPI kinase, drebrin, GAP43, cytochrome oxidase, heme oxygenase, calbindin, adenosine A1 receptors, mono amine metabolites, choline acetyltransferase, acetylcholinesterase, and symptosomal choline uptake are all reduced in AD.

Additional markers that are associated with AD or after treatment of cells with A β include (1) cPLA₂ (Stephenson *et al.*, *Neurobiology of Diseases* 3:51-63 (1996)), which is upregulated in AD, (2) Heme oxygenase-1 (Premkumar *et al.*, *J. Neurochemistry* 65:1399-1402 (1995); Schipper *et al.*, *Annals of Neurology* 37:758-768 (1995); Smith *et al.*, *American Journal of Pathology* 145:42-47 (1994); Smith *et al.*, *Molecular & Chemical Neuropathology* 24:227-230 (1995)), c-jun (Anderson *et al.*, *Experimental Neurology* 125:286-295 (1994); Anderson *et al.*, *J. Neurochemistry* 65:1487-1498 (1995)), c-fos (Anderson *et al.* (1994); Zhang *et al.*, *Neuroscience* 46:9-21 (1992)), HSP27 (Renkawek *et al.*, *Acta Neuropathologica* 87:511-519 (1994); Renkawek *et al.*, *Neuroreport* 5:14-16 (1993)), HSP70 (Cisse *et al.*, *Acta Neuropathologica* 85:233-240 (1993)), and MAP5 (Geddes *et al.*, *J. Neuroscience Research* 30:183-191 (1991); Takahashi *et al.*, *Acta Neuropathologica* 81:626-631 (1991)), which are induced in AD and in cortical cells after A β treatment, and (3) junB, junD, fosB, fra1 (Estus *et al.*, *J. Cell Biology* 127:1717-1727 (1994)), cyclin D1 (Freeman *et al.*, *Neuron* 12:343-355 (1994); Kranenburg *et al.*, *EMBO Journal* 15:46-54 (1996)), p53 (Chopp, *Current Opinion in Neurology & Neurosurgery* 6:6-10 (1993); Sakhi *et al.*, *Proc. Natl. Acad. Sci. USA* 91:7525-7529 (1994); Wood and Youle, *J. Neuroscience* 15:5851-5857 (1995)), NGFI-A (Vaccarino *et al.*,

Molecular Brain Research 12:233-241 (1992)), and NGFI-B, which are induced in cortical cells after A β treatment.

F. Measuring the Amounts and Localization of AD Markers.

Quantitative measurement can be accomplished using many standard
 5 assays. For example, transcript levels can be measured using RT-PCR and
 hybridization methods including RNase protection, Northern analysis, and R-
 dot analysis. Protein marker levels can be assayed by ELISA, Western
 analysis, and by comparison of immunohistochemically stained tissue
 sections. Immunohistochemical staining can also be used to assay
 10 localization of protein markers to particular tissues and cell types. The
 localization and the histopathological association of AD markers can be
 determined by histochemical detection methods such as antibody staining,
 laser scanning confocal imaging, and immunoelectron micrography.
 Examples of such techniques are described in Masliah *et al.* (1993) and in
 15 Example 6 below.

In the case of receptors and enzymatic markers, activity of the
 receptors or enzymes can be measured. For example, the activity of
 neurotransmitter metabolizing enzymes such as choline acetyltransferase and
 acetylcholine esterase can be measured using standard radiometric enzyme
 20 activity assays.

The activity of certain neurotransmitter receptors can be determined
 by measuring phosphoinositol (PI) turnover. This involves measuring the
 accumulation of inositol after stimulation of the receptor with an agonist.
 Useful agonists include carbachol for cholinergic receptors and
 25 norepinephrine for glutaminergic receptors. The number of receptors present
 in brain tissue can be assessed by quantitatively measuring ligand binding to
 the receptors.

The levels and turnover of receptor ligands and neurotransmitters can
 be determined by quantitative assays taken at various time points. Dopamine

turnover can be measured using DOPAC and HVA. MOPEG sulfate can be used to measure norepinephrine turnover and 5-HIAA can be used to measure serotonin turnover. For example, norepinephrine levels have been shown to be reduced 20% in the hippocampus of 12 to 13 month old PDAPP

- 5 transgenic mice relative to controls. Generally, the above assays can be performed as described in the literature, for example, in Rylett *et al.* (1983); Sims *et al.* (1980); Coyle *et al.*, *Science* 219:1184-1190 (1983); Davies and Maloney, *Lancet* 2:1403 (1976); Perry *et al.*, *Lancet* 1:189 (1977); Sims *et al.*, *J. Neurochem.* 40: 503-509 (1983). These markers are also described by
- 10 Bymaster *et al.*, *J. Pharm. Exp. Ther.* 269:282-289 (1994).

G. Screening Assays Using Cultured Cells.

- Screening assays for determining the therapeutic potential of compounds can also be performed using cells derived from animals transgenic for the disclosed APP constructs and cell cultures stably
- 15 transfected with the disclosed constructs. For example, such assays can be performed on cultured cells in the following manner. Cell cultures can be transfected generally in the manner described in International Patent Application No. 94/10569 and Citron *et al.* (1995). Derived transgenic cells or transfected cell cultures can then be plated in Corning 96-well plates at
- 20 1.5 to 2.5 x 10⁴ cells per well in Dulbecco's minimal essential media plus 10% fetal bovine serum.

- Following overnight incubation at 37°C in an incubator equilibrated with 10% carbon dioxide, media are removed and replaced with media containing a compound to be tested for a two hour pretreatment period and
- 25 cells were incubated as above. Stocks containing the compound to be tested are first prepared in 100% dimethylsulfoxide such that at the final concentration of compound used in the treatment, the concentration of dimethylsulfoxide does not exceed 0.5%, preferably about 0.1%.

At the end of the pretreatment period, the media are again removed and replaced with fresh media containing the compound to be tested as above and cells are incubated for an additional 2 to 16 hours. After treatment, plates are centrifuged in a Beckman GPR at 1200 rpm for five minutes at
 5 room temperature to pellet cellular debris from the conditioned media. From each well, 100 μ L of conditioned media or appropriate dilutions thereof are transferred into an ELISA plate precoated with antibody 266 (an antibody directed against amino acids 13 to 28 of A β) as described in International Patent Application No. 94/10569 and stored at 4°C overnight. An ELISA
 10 assay employing labelled antibody 6C6 (against amino acids 1 to 16 of A β) can be run to measure the amount of A β produced. Different capture and detection antibodies can also be used.

Cytotoxic effects of the compounds are measured by a modification of the method of Hansen *et al.*, *J. Immun. Method.* 119:203-210 (1989). To
 15 the cells remaining in the tissue culture plate, 25 μ L of a 3,(4,5-dimethylthiazol-2-yl)2,5-diphenyltetrazolium bromide (MTT) stock solution (5 mg/mL) is added to a final concentration of 1 mg/mL. Cells are incubated at 37°C for one hour, and cellular activity is stopped by the addition of an equal volume of MTT lysis buffer (20% w/v sodium
 20 dodecylsulfate in 50% dimethylformamide, pH 4.7). Complete extraction is achieved by overnight shaking at room temperature. The difference in the OD_{562nm} and the OD_{650nm} is measured in a Molecular Device's UV_{max} microplate reader, or equivalent, as an indicator of the cellular viability.

The results of the A β ELISA are fit to a standard curve and expressed
 25 as ng/mL A β . In order to normalize for cytotoxicity, these results are divided by the MTT results and expressed as a percentage of the results from a control assay run without the compound.

All publications cited herein are hereby incorporated by reference.

Example 1: Expression of pMTAPP-1 in NIH3T3 and PC12 Cells.

The clone pMTAPP-1 is an example of an APP770 expression construct as shown in Figure 1a where the promoter used is the metallothionine promoter. Stable cell lines were derived by transfecting

5 NIH3T3 and PC12 cell lines (ATCC #CCL92 and CRL1721). Five hundred thousand NIH3T3 or PC12 cells were plated into 100 mm dishes and transfected with a mixture of 5 mg of the *SalI* fragment and 1 mg of pSV2neo DNA (Southern and Berg (1982)) precipitated in the presence of 50 mg lipofectin (Gibco, BRL) in a final volume of 100 μ l. Polylysine-coated

10 plates were used for PC12 cells, which normally do not adhere well to tissue culture dishes. The cells were fed with selection medium containing 10% fetal bovine serum in DMEM or RPMI and supplemented with G418. Five hundred mg/ml (biological weight) and 250 mg/ml of G418 were used to select colonies form NIH3T3 and PC12 cells, respectively. Fifteen days

15 after transfection, colonies of cells resistant to G418 were isolated by cloning rings and expanded in T flasks. The presence of APP cDNA in the cells was detected by PCR using the procedure of Mullis and Faloona, *Methods Enzymol.* 155:335-350 (1987), the teachings of which are incorporated herein.

20 Expression of APP in 25 colonies from each cell line was analyzed by immunostaining (Majocha *et al.* (1988)). Cells were grown to subconfluence and fixed in a solution containing 4% paraformaldehyde, 0.12 M NaCl, and 20 mm Na_3PO_4 , pH 7.0. They were incubated overnight with a primary monoclonal antibody against a synthetic A β sequence (Masters *et al.* (1985);

25 Glenner and Wong) provided by Dr. Ronald Majocha, Massachusetts General Hospital, Boston, MA, followed by a generalized anti-mouse antibody conjugated to biotin (Jackson ImmunoResearch Labs, PA). Immunostaining was then performed by adding avidin-horseradish peroxidase (HRP) (Vector

Labs, Burlingame, CA) and diaminobenzidine as the chromogen (Majocha *et al.* (1985)). The results indicated that the pMTAPP-1 vector was expressing APP in both NIH3T3 and PC12 cells.

5 **Example 2: Expression of pEAPP-1 in PC12 Cells.**

pEAPP-1 is an example of an APP770 expression construct as shown in Figure 1a where the promoter used is the 25 kb human APP gene promoter. DNA from this construct was transfected into PC12 cells as described above. Certain clones of pEAPP-1 transfected cells exhibited a

10 differentiation phenotype morphologically similar to that exhibited by PC12 cells treated with nerve growth factor (NGF). PC12 cells normally are fairly round and flat cells. Those transfected with pEAPP-1 have cytoplasmic extensions resembling neurites. PC12 cells treated with NGF extend very long neuritic extensions. Thirteen PC12 cell clones transfected

15 with pEAPP-1 were selected and propagated. Eight of these cell clones exhibited the spontaneous differentiation phenotype with clones 1-8, 1-1, and 1-4 exhibiting the strongest phenotypes. Staining of pEAPP-1 transfected PC12 cells with antibody against the A β as described in Example 1 indicated that those cells exhibiting the differentiation were also expressing APP.

20 Because PC12 cells transfected with the pMTAPP-1 clone did not exhibit this phenotype even though the APP770 cDNA is expressed, these results suggest that expression of APP770 from the human promoter has novel properties regarding the physiology of the cell.

25 **Example 3: Expression of pMTA4 in PC12 Cells.**

pMTA4 is an example of the type of construct shown in Figure 4a where the promoter used is the metallothionine promoter. The protein encoded by this construct differs slightly from that depicted in Figure 4a. An APP770 cDNA clone was digested with *Asp*718 which cleaves after position

57 (number system of Kang *et al.* (1987)). The resulting 5' extension was filled in using the Klenow enzyme (Sambrook *et al.* (1989)). The same DNA preparation was also cleaved with *EcoRI* which also cuts after position 2020 and the resulting 5' extension was filled in using the Klenow enzyme

5 (Sambrook *et al.* (1989)). Self-ligation of this molecule results in an expression clone in which the truncated protein thus encoded contains the leader sequence, followed by a shortened version of the A β starting with the sequence Phe-Arg-Val-Gly-Ser-of the A β followed by the 56 terminal amino acids of APP. DNA from this construct was transfected into PC12 cells as

10 described above.

Example 4: Generation of Transgenic Mice expressing APP under the control of the MT-1 promoter.

Transgenic mice were made by microinjecting pMTAPP-1 vector

15 DNA into pronuclear embryos. pMTAPP-1 is an example of the type of construct shown in Figure 1a in which the APP770 coding sequence is operably linked to the metallothionine promoter. The procedures for microinjection into mouse embryos are described in *Manipulating the Mouse Embryo* by Hogan *et al.* (1986). Only a brief description of the procedures

20 is described below.

Mice were obtained from Taconic Laboratories (German Town, New York). Swiss Webster female mice were used for embryo retrieval and implantation. B6D2F₁ males were used for mating and vasectomized Swiss webster studs were used to simulate pseudopregnancy.

25 **A. Embryo Recovery.**

Female mice, 4 to 8 weeks of age, were induced to superovulate with 5 IU of pregnant mare's serum gonadotropin (PMSG; Sigma) followed 48 hours later by 5 IU of human chorionic gonadotropin (hCG; Sigma). Females were placed with males immediately after hCG injection. Embryos

were recovered from excised oviducts of mated females 21 hours after hCG in Dulbecco's phosphate buffered saline with 0.5% bovine serum albumin (BSA; Sigma). Surrounding cumulus cells were removed with hyaluronidase (1 mg/ml). Pronuclear embryos were then washed and placed in Earle's
 5 balanced salt solution containing 0.4% BSA (EBSS) in a 37.5°C incubator with a humidified atmosphere at 7% CO₂, 5% O₂, and 88% N₂ until the time of injection.

B. Microinjection.

Elutip-D™ purified *SalI* DNA was dissolved in 5 mM Tris (pH 7.4)
 10 and 0.1 mM EDTA at 3 µg/ml concentration for microinjection. Microneedles and holding pipettes were pulled from Fisher coagulation tubes (Fisher) on a DKI model 720 pipette puller. Holding pipettes were then broken at approximately 70 µm (O.D.) and fire polished to an I.D. of about 30 µm on a Narishige microforge (model MF-83). Pipettes were mounted on
 15 Narishige micromanipulators which were attached to a Nikon Diaphot microscope. The air-filled injection pipette was filled with DNA solution through the tip after breaking the tip against the holding pipette. Embryos, in groups of 30 to 40, were placed in 100 µl drops of EBBS under paraffin oil for micromanipulation. An embryo was oriented and held with the
 20 holding pipette. The injection pipette was then inserted into the male pronucleus (usually the larger one). If the pipette did not break through the membrane immediately the stage was tapped to assist in penetration. The nucleus was then injected and the injection was monitored by swelling of the nucleus. Following injection, the group of embryos was placed in EBSS
 25 until transfer to recipient females.

C. Transfer.

Randomly cycling adult female mice were paired with vasectomized Swiss Webster males. Recipient females were mated at the same time as donor females. At the time of transfer, the females were anesthetized with

avertin. The oviducts were exposed by a single midline dorsal incision. An incision was then made through the body wall directly over the oviduct. The ovarian bursa was then torn with watch makers forceps. Embryos to be transferred were placed in DPBS and in the tip of a transfer pipet (about 10 to 12 embryos). The pipet tip was inserted into the infundibulum and embryos were transferred. After the transfer, the incision was closed by two sutures.

D. Analysis Of Mice For Transgene Integration.

At three weeks of age or older, tail samples about 1 cm long were excised for DNA analysis. The tail samples were digested by incubating with shaking overnight at 55°C in the presence of 0.7 ml 5 mM Tris, pH 8.0, 100 mM EDTA, 0.5% SDS and 350 µg of proteinase K. The digested material was extracted once with an equal volume of phenol and once with an equal volume of phenol:chloroform (1:1 mixture). The supernatants were mixed with 70 µl 3 M sodium acetate (pH 6.0) and the DNA was precipitated by adding equal volume of 100% ethanol. The DNA was spun down in a microfuge, washed once with 70% ethanol, dried and dissolved in 100 µl TE buffer (10 mM Tris pH 8.0 and 1 mM EDTA).

Ten to twenty microliters of DNA from each sample was digested with *Bam*HI, electrophoresed on 1% agarose gels, blotted onto nitrocellulose paper, and hybridized with ³²P-labeled APP cDNA fragment. Transgenic animals were identified by autoradiography of the hybridized nitrocellulose filters. The DNAs were also analyzed by PCR carried out by synthetic primers to generate an 800 bp fragment of APP DNA.

A total of 671 pronuclear embryos were microinjected out of which 73 live and 6 dead pups were born. DNA analysis identified 9 transgenic mice (5 females and 4 males) which were bred to generate F₁ and F₂ transgenics. These animals can be analyzed for expression of mRNA and protein of APP in different tissues and for analysis of behavioral and

pathological abnormalities as described above. Transgenic mice with this construct express transgenic RNA.

5 **Example 5: Construction of APP construct containing a combination cDNA/genomic coding sequence.**

A cDNA/genomic APP construct containing introns 6, 7 and 8 was prepared by combining APP cDNA encoding exons 1-6 and 9-18 with genomic APP sequences encoding introns 6, 7 and 8, and exons 7 and 8 (see ~~Figure 6~~ ^{Figures 6a and 6b}). In order to create a splicing cassette small enough for convenient insertion in a pUC vector, two deletions in intronic sequences were made. A deletion was made in intron 6 from position 143 of intron 6 to the *Bam*HI site located upstream of the beginning of exon 7 (1658 bp before the beginning of exon 7). Another deletion was made in intron 8 from the first *Bam*HI site in intron 8 to a site at 263 bp before the beginning of exon 9.

10 To avoid confusion, these truncated forms of APP introns 6 and 8 are referred to herein as intron Δ 6 and intron Δ 8. *Bam*HI sites were engineered at the sites of these deletions, so that they are marked by the presence of *Bam*HI sites. In this construct, referred to as PDAPP, exons 7 and 8 and intron 7 are intact genomic sequences, except that the unique *Xho*I site in

15 intron 7 was destroyed.

DNA fragments containing the truncated introns were generated as follows: a *Bam*HI site was engineered 143 bp into intron 6 nucleotide by PCR mutagenesis ("Mutagenesis by PCR" in *PCR Technology: Current Innovations* (Griffith and Griffith, eds., CRC Press, 1994) pages 69-83) and

25 another *Bam*HI site was engineered by PCR mutagenesis 263 bp prior to the beginning of exon 9. These sites were engineered into separate APP genomic DNA clones containing the junctions of exon 6 and intron 6, and intron 8 and exon 9, respectively, resulting in modified APP genomic DNA clones.

The entire cassette was assembled in the APP cDNA clone as follows (Figure 11). The 889 bp *Bam*HI to *Xcm*I fragment of APP cDNA containing exons 1 through 5 and part of exon 6 (including nucleotides 1 to 843 of SEQ ID NO:5) was cloned into a vector containing *Bam*HI and *Xho*I sites

- 5 downstream from the insertion site to make APP770x-oligo-x. APP770x-oligo-x was then cut with *Xcm*I and *Bam*HI. Then two fragments were obtained from the modified APP genomic DNA clone containing the junction of exon 6 and intron 6 described above by cutting with *Xcm*I and *Bam*HI. The resulting 34 bp fragment from the *Xcm*I in exon 6 to the *Xcm*I in intron
- 10 6, and 131 bp fragment from the *Xcm*I in intron 6 to the artificially created *Bam*HI site at position 143 bp of intron 6 were ligated into APP770x-oligo-x in a three-way ligation step to make APP770x-E6oligo-x. The orientation of the fragments was confirmed by sequencing. APP770x-E6oligo-x was then cut with *Bam*HI and *Xho*I. Then the 313 bp *Bam*HI and *Xho*I fragment from
- 15 the modified APP genomic DNA clone containing the junction of intron 8 and exon 9 was ligated into APP770x-E6oligo-x to make APP770xE6E9x.

- APP770xE6E9x was then cut with *Bam*HI and the 6.8 kb *Bam*HI fragment of APP genomic DNA encoding the KPI and OX-2 domains (exons 7 and 8) was inserted at this site. This fragment starts at the *Bam*HI site
- 20 1658 bp upstream of the start of exon 7 and extends to the first *Bam*HI site in intron 8. This *Bam*HI fragment was obtained from a lambda phage genomic clone encoding this portion of the APP gene, that was obtained from a Human Placental genomic library in the Lambda FIXII vector obtained from Stratagene. This *Bam*HI fragment originally contained an *Xho*I site
 - 25 which was destroyed by cutting, filling in, and religation. The locations of the deletions are diagramed in Figure 10. This clone, containing exons 1-8 and part of 9, and introns 6, 7 and 8, was termed the "APP splicing cassette." The APP splicing cassette was cut out with *Nru*I and *Xho*I and

used to replace the *NruI* to *XhoI* cDNA fragment of APP cDNA bearing the Hardy mutation. This mutant form of APP cDNA was produced by converting the G at nucleotide position 2145 to T by site directed mutagenesis. This changes the encoded amino acid from Val to Phe. The
 5 resulting construct is a combination cDNA/genomic APP "minigene."

Sequencing of the 6.8 kb *BamHI* fragment containing APP exons 7 and 8 derived from the APP genomic clone used to generate this construct showed that intron 7 is 2.6 kb long, and that the first *BamHI* site in intron 8, the upstream site of the deletion in intron 8 engineered into the APP
 10 minigene construct, is 2329 bp downstream from the end of exon 8. This does not coincide with the restriction map of the APP gene published by Yoshikai *et al.* (1990) and Yoshikai *et al.* (1991). Comparison of their map to our sequence indicates that Yoshikai *et al.* switched the order of two
 15 *EcoRI* fragments in their restriction mapping. The 1.60 kb *EcoRI* fragment containing exon 8 is actually upstream of the 1.48 kb *EcoRI* fragment and the 1.48 kb *EcoRI* fragment Yoshikai *et al.* mapped in intron 7 is actually in intron 8. We have confirmed this location for the *EcoRI* fragment containing exon 8 by sizing of PCR generated fragments from human DNA.

This APP minigene was operatively linked to the PDGF-B promoter
 20 to provide expression of the APP cDNA/genomic construct in mammalian cells. The PDGF β -chain 5' flanking sequence was inserted upstream of the *NruI* site at the beginning of the APP minigene. This fragment includes 1.3 kb upstream of the transcription initiation site, where the PDGF-B promoter resides, and approximately 70 bp of 5' untranslated region, ending at the
 25 *AurII* site (Higgins *et al.* (1994)). The late SV40 polyadenylation signal, carried on a 240 bp *BamHI* to *BclI* fragment, was added downstream of the APP minigene. This construct, combining the PDGF-B promoter, the APP

splicing cassette, the Hardy mutation, and the SV40 polyadenylation signal is referred to as PDAPP (Figure 9).

Example 6: Transgenic mice containing the PDAPP construct.

5 Transgenic mice were generated using the PDAPP construct described in Example 5. Transgenic mice were generated by microinjection using standard techniques as described above. PDAPP DNA was microinjected into the embryos at the two-cell stage. Plasmid sequences (pUC) were removed by *SacI* and *NotI* digestion before microinjection. Seven founder
10 mice were generated and line 109 was used for extensive analysis. Only heterozygous animals were used. Southern analysis of 104 animals from four generations showed that approximately 40 copies of the transgene were inserted at a single site and transmitted in a stable manner. Human APP messenger RNA was produced in several tissues of the transgenic mouse, but
15 at especially high levels in brain. RNase protection assays revealed at least 20-fold more APP expression in the brains of line 109 animals than in the mouse lines expressing neuron-specific enolase (NSE)-promoter-driven APP transgenes previously described by Quon *et al.* (1991), Mucke *et al.*, *Brain Res.* 666:151-167 (1994), McConlogue *et al.*, *Neurobiol. Aging* 15:S12
20 (1994), and Higgins *et al.*, *Ann Neurol.* 35:598-607 (1994).

A. Expression of APP Transcripts and Protein.

RNA was isolated from brain tissue as described by Chomaczynski and Sacchi, *Analyt. Biochem.* 162:156-159 (1987), and subjected to RT-PCR as described by Wang *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 86:9717-9721
25 (1989), using human-specific APP primers
(5'-CCGATGATGACGAGGACGAT-3', SEQ ID NO:7;
5'-TGAACACGTGACGAGGCCGA-3', SEQ ID NO:8) using 40 cycles of 1 minute at 94°C, 40 seconds at 60°C, and 50 seconds at 72°C. RT-PCR

analysis demonstrated the presence of transcripts encoding the 695, 751 and 770 isoforms of human APP in transgenic animal brains but not in brains from non-transgenic littermates. The identities of the human APP RT-PCR bands from the transgenic mouse RNA were verified by subcloning and sequencing.

The relative levels and alternative splicing of APP transcripts in brains of PDAPP transgenic mice, NSE-APP transgenic mice, non-transgenic mice, and humans with and without AD were compared in RNase protection assays (Rockenstein *et al.*, *J. Biol. Chem.* 270:28257-28267 (1995)).

PDAPP mice expressed approximately 5-fold higher total APP mRNA levels than non-transgenic controls, and at least 20-fold higher human APP mRNA levels than most NSE-APP transgenic mice. While NSE-driven human APP expression does not affect the levels of murine APP mRNA, PDAPP transgenic mice showed a significant 30% decrease in murine APP transcripts. While the relative abundances of murine APP770:751:695 mRNAs in non-transgenic mouse brains were roughly 1:1:35, the corresponding human APP mRNA levels in PDAPP transgenic mouse brains were 5:5:1.

Analysis of holo-APP was performed by brain homogenization in 10 volumes of PBS containing 0.5 mM EDTA, 10 $\mu\text{g ml}^{-1}$ leupeptin and 1 mM PMSF. Samples were spun at 12,000g for 10 min and the pellets resuspended in RIPA (150 mM NaCl, 50 mM Tris, pH 8.0, 20 mM EDTA, 1.0% deoxycholate, 1.0% Triton X-100, 0.1% SDS, 1 mM PMSF and 10 $\mu\text{g ml}^{-1}$ leupeptin). Samples (each containing 30 μg total protein) were analyzed by SDS-PAGE, transferred to Immobilon membranes and reacted with either the holo-APP antibody, anti-6 (anti Bx 6), described by Oltersdorf *et al.*, *J. Biol. Chem.* 285:4492-4497 (1990), or 8E5 monoclonal antibody. 8E5 was prepared against a bacterial fusion protein encompassing human APP residues

444-692 (Oltersdorf *et al.* (1990)) and is human-specific, showing essentially no crossreactivity against mouse APP. Immunoblot analysis of total APP expression (human and mouse) in transgenic mouse line 109 and control littermate brain tissue using C-terminal APP antibody anti-6 showed much
 5 higher levels of expression in the transgenic mice. Immunoblot analysis of brain homogenates using either the holo-APP polyclonal antibody anti-6 or the human-specific APP monoclonal antibody 8E5 revealed human APP over-expression in the transgenic mouse at levels at least 3-fold higher in hippocampus than either endogenous mouse APP levels or those in AD brain.

10 For immunoblot analysis of A β , a 9-month-old mouse brain was homogenized in 5 ml 6 M guanidine HCl, 50 mM Tris, pH 7.5. The homogenate was centrifuged at 100,000g for 15 min and the supernatant was dialyzed against H₂O overnight adjusted to PBS with 1 mM PMSF and 25 μ g ml⁻¹ leupeptin. This material was immunoprecipitated with antibody 266
 15 resin, and immunoblotted with the human-specific A β antibody, 6C6, as described by Seubert *et al.*, *Nature* 359:325-327 (1992). Using this human-specific A β antibody (6C6), a 4 kD β amyloid-immunoreactive peptide was isolated from the brains of the transgenic animals, which corresponds to the relative molecular mass of A β . Brain levels of A β were at least 10-fold
 20 higher in line 109 animals than in the previously described human APP transgenic mice. Embryonic day 16 cortical cell cultures from transgenic animals constitutively secreted human A β , including a substantial fraction of A β 1-42 (5 ng ml⁻¹ total A β ; 0.7 ng ml⁻¹ A β 1-42), as detected in media by human-specific A β enzyme-linked immunosorbent assays, as described by
 25 Seubert *et al.* (1992) and McConlogue *et al.* (1994), and as described in Example 8. Thus, line-109 animals greatly overexpressed human APP mRNA, holo-APP and A β in their brains.

B. Histopathology of PDAPP Transgenic Mice.

Brains from 180 transgenic and 160 age-matched non-transgenic age-matched controls (4 to 20 months old) representing five generations of the line 109 pedigree were extensively examined histopathologically. Some mouse brains were removed and placed in alcohol fixative (Arai *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 87:2249-2253 (1990)) for 48 hours before paraffin embedding. Other mice were perfused with saline followed by 4% paraformaldehyde in 0.1 M sodium phosphate. For paraffin embedded brains, 6 μ m coronal or parasagittal sections from transgenic and non-transgenic mice were placed adjacent to each other on poly-L-lysine coated slides. The sections were deparaffinized, rehydrated and treated with 0.03% H_2O_2 for 30 min before overnight incubation at 4°C with a 1:1,000 dilution of the A β antibody, R1280 (Tamaoka *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 89:1345-1349 (1992)). For absorption studies, synthetic human A β 1-40 peptide (Games *et al.*, *Neurobiol. Aging* 13:569-576 (1992)) in 10% aqueous dimethylsulphoxide was added to a final concentration of 7.0 μ M to the diluted antibody and incubated for 2 hours at 37°C. The diluent was applied to the sections and processed under the same conditions as the standard antibody solution. Peroxidase rabbit IgG kit (Vector Labs) was then used as recommended, with 3,3'-diaminobenzidine (DAB) as the chromogen. Similarly fixed human AD brain was processed simultaneously under identical conditions.

Before 4 months of age, no obvious A β deposition was detected. However, by approximately 4 months of age, the transgenic animals began to exhibit deposits of human A β in the hippocampus, corpus callosum and cerebral cortex. These A β plaques increased with age, and by eight months many deposits of 30 to 200 μ m were seen. As the animals aged beyond 9 months, the density of the plaques increased until the A β -staining pattern resembled that of AD. Vascular amyloid, another feature of AD pathology,

developed in older mice. Robust pathology was also seen in another transgenic line generated from the PDAPP vector (line 35).

$A\beta$ deposits of varying morphology were clearly evident as a result of using a variety of $A\beta$ antibodies, including well characterized human-specific $A\beta$ antibodies and antibodies specific for the free amino and carboxy termini of $A\beta$ 1-42. Antibody 9204, described by Saido *et al.*, *J. Biol. Chem.* 289:15253-15257 (1994), is specific to $A\beta$ 1-5 and was used at a concentration of $7.0 \mu\text{g ml}^{-1}$. Antibody 277-2, specific for $A\beta$ 1-42, was prepared by immunizing New Zealand white rabbits with the peptide cysteine-aminoheptanoic acid- $A\beta$ 33-42 conjugated to cationized BSA ('Super Carriers'; Pierce) using a standard immunization protocol ($500 \mu\text{g}$ per injection). Specific antibodies were affinity-purified from serum against the immunogen immobilized on agarose beads. Before incubation with antibody 277-2, sections were treated for 1 to 2 min with 80% formic acid. For detection, the antibody was reacted using the peroxidase rabbit IgG kit (Vector Labs). The product was then visualized using DAB as the chromogen. Some sections were then incubated overnight at 4°C with a 1:500 dilution of polyclonal anti-GFAP (Sigma). The GFAP antibody was reacted using the alkaline phosphatase anti-rabbit IgG kit and alkaline phosphatase substrate kit 1 (Vector Labs; used according to the manufacturer's recommendations). Additional sections were incubated overnight with the F480 antibody (Serotec) used at a 1:40 dilution to visualize microglial cells. The mouse peroxidase kit (Vector Labs) was then used according to the manufacturer's recommendations. Some sections were stained with thioflavin S using standard procedures (Dickson *et al.*, *Acta Neuropath.* 79:486-493 (1990)) and viewed with ultraviolet light through an FITC filter of maximum wavelength 440 nm.

Serial sections demonstrated many plaques were positively stained with both the 9204 and 277-2 antibodies. The forms of the $A\beta$ deposition

ranged from diffuse irregular types to compacted plaques with cores.
 Roughly spherical, and wispy, irregular deposits, were labelled with antibody
 9204 specific for the free amino terminus of A β . Astrocytic gliosis
 associated with A β deposition was evident after double immunolabelling with
 5 antibodies to glial fibrillary acidic protein (GFAP) and human A β . A
 compacted A β core and 'halo' was evident in several plaques. Non-
 transgenic littermates showed none of these neuropathological changes.
 Immunostaining was fully absorbable with the relevant synthetic peptide, and
 was apparent using a variety of processing conditions, including fixation with
 10 paraformaldehyde and Trojanowski methods. Many plaques were stained
 with thioflavin S, and some were also stained using the Bielschowsky silver
 method and were birefringent with Congo Red, indicating the true amyloid
 nature of these deposits.

The majority of plaques were intimately surrounded by GFAP-positive
 15 reactive astrocytes, similar to the gliosis found in AD plaques. The
 neocortices of the transgenic mice contained diffusely activated microglial
 cells, as defined by their amoeboid appearance, shortened processes, and
 staining with Mac-1 antibody. Staining by antibodies recognizing
 phosphorylated neurofilaments and phosphorylated tau indicated that aberrant
 20 phosphorylation occurred in PDAPP brain that was similar to AD. These
 phosphorylations are seen in AD and are thought to preclude formation of
 neurofibrillary tangles. Although paired helical filaments (PHF) have not yet
 been detected in PDAPP mice, the detection of abnormally phosphorylated
 neurofilaments and tau are thought to be associated with, and the initial step
 25 in, the formation of PHF in AD.

Clear evidence for neuritic pathology was apparent using both
 conventional and confocal immunomicroscopy. Forty μ m thick vibratome
 sections were incubated overnight at 4°C with R1280 (1:1,000) in
 combination with polyclonal anti-synaptophysin (1:150; Dako) or 8E5 (7.0

μg^{-1}). Some sections were incubated with anti-synaptophysin or monoclonal anti-MAP 2 (1:20, Boehringer-Mannheim), and then reacted with a goat anti-rabbit biotinylated antibody (1:100) followed by a mixture of FITC-conjugated horse anti-mouse IgG (1:75) and avidin D Texas red (1:100) (Vector Labs). The double-immunolabelled sections were viewed on a Zeiss Axiovert 35 microscope with attached laser confocal scanning system MRC 600 (Bio-Rad). The Texas red channel collected images of the R1280 or synaptophysin labelling, and the FITC channel collected synaptophysin, 8E5, or MAP 2 labelling. Optical z-sections $0.5 \mu\text{m}$ in thickness were collected from each region, similar to the image processing and storage described by Masliah *et al.*, *J. Neuropath. Exp Neurol.* 52:619-632 (1993).

Many $\text{A}\beta$ plaques were closely associated with distorted neurites that could be detected with human APP-specific antibodies and with anti-synaptophysin antibodies, suggesting that these neurites were derived in part from axonal sprouts, as observed in the AD brain. The plaques compressed and distorted the surrounding neuropil, also as in the AD brain. Synaptic and dendritic density were also reduced in the molecular layer of the hippocampal dentate gyrus of the transgenic mice. This was evident by reduced immunostaining for the presynaptic marker synaptophysin and the dendritic marker MAP-2 in AD brain (Masliah *et al.*, *Am. J. Path.* 138:235-246 (1991)).

Confirmation of the presence of extracellular $\text{A}\beta$ was obtained using immunoelectron microscopy. For immunoelectron microscopy, mice were perfused with saline followed by 2.0% paraformaldehyde and 1.0% glutaraldehyde in cacodylate buffer. Forty μm thick vibratome sections were incubated with the R1280 antibody, and reacted using a peroxidase rabbit IgG kit (Vector Labs). Immunolabelled sections with $\text{A}\beta$ deposits were then fixed in 1.0% ammonium tetroxide and embedded in epon/araldite before viewing

ultrathin sections with a Jeol CX100 electron microscope (Masliah *et al.*,
Acta Neuropath. 81:428-433 (1991)).

**Table 3. Ultrastructural Similarities and Differences
Between AD and PDAPP Transgenic Plaques.**

	Alzheimer's Disease	PDAPP
Amyloid fibrils size electron density pinocytic vesicles	9-11 nm moderate abundant	9-11 nm high occasional
Dystrophic neurites TYPE I dense laminar bodies synaptic vesicles and contacts neurofilament accumulation TYPE II paired helical filaments	abundant yes yes yes	abundant yes yes none?
Cells associated with amyloid formation microglia neurons neurosecretory granules rough endoplasmic reticulum coated pits	abundant occasional abundant abundant yes	occasional abundant abundant abundant yes

Tables 3 and 4 present a summary of the above results, showing
cytological and pathological similarities between AD and PDAPP mice. For
every feature examined, with the exception of paired helical filaments, the
PDAPP mice exhibited pathology characteristic of AD. These findings show
5 that production of human APP in transgenic (TG) mice is sufficient to cause
not only amyloid deposition, but also many of the complex subcellular
degenerative changes associated with AD.

Table 4. Pathology in Alzheimer's Disease and the PDAPP Mouse.

		Alzheimer's Disease	PDAPP
	A β Deposition into Plaques		
5	Diffuse	+	+
	Neuritic	+	+
	Vascular	+	+
	Brain Region Specificity	+	+
	Neuritic Dystrophy	+	+
	Synaptic Loss	+	+
10	Inflammatory Response		
	Astrocytosis	+	+
	Microgliosis	+	+
	Cytoskeletal Alterations		
15	Phosphorylated Neurofilaments	+	+
	Phosphorylated Tau	+	+
	PHF/Tangles	+	-(?)

The most notable feature of these transgenic mice is their Alzheimer-like neuropathology, which includes extracellular A β deposition, dystrophic neuritic components, gliosis, and loss of synaptic density with regional specificity resembling that of AD. Plaque density increases with age in these transgenic mice, as it does in humans (Selkoe, *Rev. Neurosci.* 17:489-517 (1994)), implying a progressive A β deposition that exceeds its clearance, as also proposed for AD (Maggio *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* 89:5462-5466 (1992)). The PDAPP transgenic mice provide strong new evidence for the primacy of APP expression and A β deposition in AD neuropathology. Such mice also provide a sufficiently robust AD model in which to test whether compounds that lower A β production and/or reduce its neurotoxicity *in vitro* can produce beneficial effects in an animal model prior to advancing such drugs into human trials.

Example 7: Construction APP transgenes expressing APP from the PDGF-B promoter.

PDAPP transgenic mice contain a splicing cassette that permits expression of all three major human APP isoforms, where expression is driven by the PDGF-B promoter, and which includes a mutation in amino acid 717, the site of familial AD mutations. It is expected that these features, and others described above, can be used independently to produce transgenic mice useful as models of Alzheimer's disease. Some specific examples of such constructs are described below.

A. Construction of PDAPP-wt.

A wild type version of the cDNA/genomic clone PDAPP was constructed in which the mutation to amino acid 717 was replaced with the wild type. This was accomplished by replacing the 1448 bp *XhoI* to *SpeI* fragment of PDAPP, which includes the part of the APP cDNA sequence that encodes the Hardy mutation in which Val717 is replaced by Phe, with the 1448 bp *XhoI* to *SpeI* fragment of a wild type APP clone. This fragment corresponds to the region from position 1135 to 2588 of SEQ ID NO:5. None of the intron sequences of PDAPP are replaced or removed by this substitution. This construct is referred to as PDAPP-wt. A schematic of PDAPP-wt and its construction is shown in Figure 12.

B. Construction of PDAPP-SwHa.

Another version of the cDNA/genomic clone PDAPP was constructed in which the Swedish mutant at amino acids 670 and 671 was introduced. Plasmid pNSE751.delta3'spl.sw contains cDNA of the human APP751 which includes the Swedish mutation of Lys to Asn and Met to Leu at amino acids 670 and 671, respectively. A 563 bp *EcoRI* to *SpeI* fragment from this plasmid was replaced with the corresponding 563 bp *EcoRI* to *SpeI* fragment of PDAPP, which includes an identical part of the APP cDNA sequence with the exception of Phe717 of the Hardy mutation. This fragment corresponds

to the region from position 2020 to 2588 of SEQ ID NO:5. This results in pNSE.delta3'spl.sw/ha, which contains both the Swedish mutation at amino acids 670 and 671, and the Hardy mutation at amino acid 717.

The 1448 bp *Xho*I to *Spe*I fragment of PDAPP was then replaced with
 5 the 1448 bp *Xho*I to *Spe*I fragment of pNSE752.delta3'spl.sw/ha, which contains both the Swedish mutation and the Hardy mutation, to form PDAPP-Sw/Ha. A schematic of PDAPP-Sw/Ha and its construction is shown in Figure 13.

C. Construction of PDAPP695_{v.F}.

10 A construct encoding only APP695, but retaining the Hardy mutation, PDGF-B promoter, and vector sequences of PDAPP, can be made. This can be accomplished by ligating the 6.6 kb *Xho*I to *Nru*I fragment from PDAPP, which contains the C-terminal part of the APP sequences, and the polyadenylation, pUC, and PDGF-B promoter sequences, to the 1.2 kb *Xho*I
 15 to *Bcl*I fragment of pCK695, which contains a hybrid splice signal and the remaining N-terminal portion of the APP sequences (on a 911 bp *Xho*I to *Nru*I fragment of APP695 cDNA). The hybrid splice signal is the same as was described earlier and is also present in vector pohCK751, which is described by Dugan *et al.*, *J Biological Chem.* 270:10982-10989 (1995).
 20 pCK695 is identical to pohCK751 except that the herpes simplex virus replication and packaging sequences of pohCK751 were removed, and the plasmid encodes APP695 instead of APP751.

In this vector the PDGF-B promoter drives the expression of APP695 containing the mutation of Val717 to Phe. The hybrid splice signal is
 25 included to potentially enhance expression. Additional vectors derived from this may be constructed which lack any splice signals, or into which other splice signals have been added to obtain this same function.

D. Construction of PDAPP751_{V-F}.

A construct encoding only APP751, but retaining the Hardy mutation, PDGF-B promoter, and vector sequences of PDAPP, can be made. This can be accomplished by ligating the 6.65 kb *XhoI* to *KpnI* fragment of PDAPP, including part of the APP sequences, the polyadenylation signals, pUC and PDGF-B promoter sequences to the 1.0 kb *KpnI* to *XhoI* fragment containing the remainder of the human APP751 cDNA sequences (nucleotides 57 to 1084 of SEQ ID NO:3) to make the intermediate plasmid PDAPP δ sp751_{V-F}. The 1.0 kb *KpnI* to *XhoI* fragment encoding a portion of human APP751 can be obtained from the plasmid poCK751, which is identical to pohCK751 except that the herpes simplex viral sequences were removed.

To introduce splicing sequences, the first intron from PDAPP, which is intron Δ 6, is then inserted into PDAPP δ sp751_{V-F} to make PDAPP751_{V-F}. To accomplish this, the 2,758 bp *Asp718* to *ScaI* fragment of PDAPP containing exons 2 through 6, intron Δ 6, and part of exon 7, is ligated to the 6,736 bp fragment obtained by complete digestion of PDAPP δ sp751_{V-F} with *Asp718* and partial digestion with *ScaI*. This 6,736 bp fragment provides the remaining additional APP sequences (part of exon 1, the rest of exon 7, and exons 9 through 18), polyadenylation signals, pUC and PDGF-B promoter sequences. The resulting construct is referred to as PDAPP751_{V-F}.

In this vector the PDGF-B promoter drives the expression of APP751 containing the mutation of Val717 to Phe. One splice signal (derived from intron 6) is included to potentially enhance expression. Additional vectors derived from this may be constructed which lack any splice signals, or into which other splice signals have been added to obtain this same function.

E. Construction of PDAPP770_{V-F}.

A construct encoding only APP770, but retaining the Hardy mutation, PDGF-B promoter, and vector sequences of PDAPP, can be made. This can

be accomplished by replacing the *KpnI* to *XhoI* fragment of PDAPP751_{V.F} containing APP exons 2-7 and a part of exon 9, with the *KpnI* to *XhoI* fragment of APP770 cDNA, which contains exons 2-8 and a part of exon 9. This fragment corresponds to nucleotides 57 to 1140 of SEQ ID NO:5. The
 5 resulting construct is referred to as PDAPP770_{V.F}.

In this vector the PDGF-B promoter drives the expression of APP770 containing the mutation of Val717 to Phe. PDAPP770_{V.F} contains the same intron sequences present in PDAPP751_{V.F}. Additional vectors derived from this may be constructed into which a splice signals have been added to obtain
 10 enhanced expression.

Example 8: Expression Levels of APP Expression Products in Brain Tissue of PDAPP Mice.

The PDAPP mouse line described in Example 6 was examined for the
 15 levels of several derivatives of the APP in hippocampal, cortical, and cerebellar brain regions of mice of various ages. Levels of APP cleaved at the beta-secretase site (APP β) and APP containing at least 12 amino acids of A β (FLAPP+APP α ; a mixture of APP α and full length APP (FLAPP)) were found to be nearly constant within a given brain region at all ages evaluated.
 20 The hippocampus expressed the highest level of all APP forms. In contrast, guanidine extractable levels of A β showed remarkable age-dependent increases in a manner that mirrored the amyloid plaque deposition observed immunohistochemically. Specifically, A β levels in hippocampus increased 17-fold by 8 months of age and 106-fold by 1 year of age, compared to that
 25 found in 4 month old animals. At 1 year of age A β constitutes approximately 1% of the total protein in hippocampus. The cerebral cortex also showed large increases in A β with age. In contrast, the mean level of A β in cerebellum across all age groups was comparatively low and unchanging.

Further analysis of the A β in these brains using an ELISA specific for A β ₁₋₄₂ showed that this longer version made up 27% of the 19 pmoles/g of the A β present in the brains of young animals; this percentage increased to 97% of the 690 pmoles/g in 12 month old animals. The selective deposition
 5 of A β ₁₋₄₂ and the spacial distribution of the A β deposits are further evidence that the pathological processes ongoing in the PDAPP transgenic mice parallel the human Alzheimer's diseased condition.

Levels of A β -containing proteins were measured through the use of ELISAs configured with antibodies specific to A β , A β ₁₋₄₂, APP cleaved at the
 10 β -secretase site (Seubert *et al.* (1993)), and APP containing the first 12 amino acids of A β (FLAPP+APP α ; a mixture of full length APP and α -secretase cleaved APP (Esch *et al.*)). Striking similarities in both the regional variation and depositing form of A β are noted between the mouse model and the human AD condition. The results also show that, because of
 15 the magnitude and temporal predictability of A β deposition, the PDAPP mouse is a practical model in which to test agents that either inhibit the processing of APP to A β or retard A β amyloidosis.

A. Materials and Methods.

1. Brain Tissue Preparation.

20 The heterozygote transgenic (Line 109, Games *et al.*; Rockenstein *et al.*) and non-transgenic animals were anesthetized with Nembutol (1:5 solution in 0.9% saline) and perfused intracardially with ice cold 0.9% saline. The brain was removed and one hemisphere was prepared for immunohistochemical analysis, while four brain regions (cerebellum,
 25 hippocampus, thalamus, and cortex) were dissected from the other hemisphere and used for A β and APP measures.

To prepare tissue for ELISAs, each brain region was homogenized in 10 volumes of ice cold guanidine buffer (5.0 M guanidine-HCl, 50 mM Tris-

Cl, pH 8.0) using a motorized pestle (Kontes). The homogenates were gently mixed on a Nutator for three to four hours at room temperature, then either assayed directly or stored at -20°C prior to quantitation of A β and APP. Preliminary experiments showed the analytes were stable to this storage condition.

2. A β Measurements.

The brain homogenates were further diluted 1:10 with ice-cold casein buffer (0.25% casein, phosphate buffered saline (PBS), 0.05% sodium azide, 20 μ g/ml aprotinin, 5 mM EDTA pH 8.0, 10 μ g/ml leupeptin), reducing the final concentration of guanidine to 0.5 M, before centrifugation (16,000 x g for 20 minutes at 4°C). The A β standards (1-40 or 1-42 amino acids) were prepared such that the final composition included 0.5 M guanidine in the presence of 0.1% bovine serum albumin (BSA).

The "total" A β sandwich ELISA consists of two monoclonal antibodies (mAb) to A β . The capture antibody, 266, is specific to amino acids 13-28 of A β (Seubert *et al.* (1992)); while the antibody 3D6, which is specific to amino acids 1-5 of A β , was biotinylated and served as the reporter antibody. The 3D6 biotinylation procedure employs the manufacturer's (Pierce) protocol for NHS-biotin labeling of immunoglobulins except 100 mM sodium bicarbonate, pH 8.5 buffer was used. The 3D6 antibody does not recognize secreted APP or full-length APP but detects only A β species with amino terminal aspartic acid. The assay has a lower limit of sensitivity of approximately 50 pg/ml (11.4 pM) and showed no cross-reactivity to the endogenous murine A β peptide at concentrations up to 1 ng/ml.

The configuration of the A β ₁₋₄₂-specific sandwich ELISA employs the mAb 21F12, which was generated against amino acids 33-42 of A β . The antibody shows less than 0.4% cross-reactivity with A β ₁₋₄₀ in either ELISA or competitive radioimmunoassay (RIA). Biotinylated 3D6 is also the reporter

antibody in this assay which has a lower limit of sensitivity of approximately 125 pg/ml (28.4 pM).

The 266 and 21F12 mAbs were coated at 10 μ g/ml into 96-well immunoassay plates (Costar) overnight at room temperature. The plates were then aspirated and blocked with 0.25% human serum albumin in PBS buffer for at least 1 hour at room temperature, then stored desiccated at 4°C until use. The plates were rehydrated with wash buffer prior to use. The samples and standards were added to the plates and incubated at room temperature for 1 hour. The plates were washed at least 3 times with wash buffer (Tris buffered saline, 0.05% Tween 20) between each step of the assay.

The biotinylated 3D6, diluted to 0.5 μ g/ml in casein assay buffer (0.25% casein, PBS, 0.05% Tween 20, pH 7.4), was incubated in the well for 1 hour at room temperature. Avidin-HRP (Vector, Burlingame, CA), diluted 1:4000 in casein assay buffer, was added to the wells and incubated for 1 hour at room temperature. The colorimetric substrate (100 μ l), Slow TMB-ELISA (Pierce), was added and allowed to react for 15 minutes, after which the enzymatic reaction is stopped with 25 μ l of 2 N H₂SO₄. Reaction product was quantified using a Molecular Devices Vmax measuring the difference in absorbance at 450 nm and 650 nm.

20 3. APP ELISAs.

Two different APP assays were utilized. The first recognizes APP α and full length forms of APP (FLAPP+APP α), while the second recognizes APP β (APP ending at the methionine preceding the A β domain (Seubert *et al.* (1993)). The capture antibody for both the FLAPP+APP α and APP β assays is 8E5, a monoclonal antibody raised to a bacterially expressed fusion protein corresponding to human APP amino acids 444-592 (Games *et al.*). The reporter mAb (2H3) for the FLAPP+APP α assay was generated against amino acids 1-12 of A β . The lower limit of sensitivity for the 8E5/2H3 assay is approximately 11 ng/ml (150 pM). For the APP β assay, the

polyclonal antibody 192 was used as the reporter. This antibody has the same specificity as antibody 92 (Seubert *et al.* (1993)), that is, it is specific to the carboxy-terminus of the β -secretase cleavage site of APP. The lower limit of sensitivity for the β -secretase 8E5/192 assay is approximately 43
 5 ng/ml (600 pM).

For both APP assays, the 8E5 mAb was coated onto 96-well Costar plates as described above for 266. Purified recombinant secreted APP α (the APP751 form) and APP596 were the reference standards used for the FLAPP+APP α and APP β assays, respectively. APP was purified as
 10 described previously (Esch *et al.*) and APP concentrations were determined by amino acid analysis. The 5 M guanidine brain homogenate samples were diluted 1:10 in specimen diluent for a final buffer composition of 0.5 M NaCl, 0.1% NP-40, 0.5 M guanidine. The APP standards for the respective
 15 assays were diluted into buffer of the same final composition as for the samples. The APP standards and samples were added to the plate and incubated for 1.5 hours at room temperature. The plates were thoroughly washed between each step of the assay with wash buffer. Reporter antibodies 2H3 and 192 were biotinylated following the same procedure as for 3D6 and were incubated with samples for 1 hour at room temperature. Streptavidin-
 20 alkaline phosphatase (Boehringer Mannheim), diluted 1:1000 in specimen diluent, was incubated in the wells for 1 hour at room temperature. The fluorescent substrate 4-methyl-umbelliperyl-phosphate, was added, and the plates read on a Cytofluor™ 2350 (Millipore) at 365 nm excitation and 450 nm emission.

25 4. Monoclonal Antibody Production.

The immunogens for 3D6, 21F12, and 2H3 were separately conjugated to sheep anti-mouse immunoglobulin (Jackson ImmunoResearch Labs) using maleimidohexanoyl-N-hydroxysuccinimide (Pierce). A/J mice (Jackson Laboratories) were given intraperitoneal injections (IP) of 100 μ g of

the appropriate immunogen emulsified in Freund's complete adjuvant (Sigma) and two subsequent IP injections of 100 μ g immunogen were given on a biweekly basis in Freund's incomplete adjuvant (Sigma). Two to three weeks after the third boost, the highest titer mouse of a given immunogen was

5 injected intravenously and intraperitoneally with 50-100 μ g of immunogen in PBS. Three days post injection, the spleen was removed, splenocytes were isolated and fused with SP2/0-Ag14 mouse myeloma cells. The hybridoma supernatants were screened for high affinity monoclonal antibodies by RIA as previously described (Seubert *et al.* (1992)). Purified monoclonal antibodies

10 were prepared from ascites.

5. Immunohistochemistry.

The tissue from one brain hemisphere of each mouse was drop-fixed in 4% paraformaldehyde and post-fixed for three days. The tissue was mounted coronally and 40 μ m sections were collected using a vibratome.

15 The sections were stored in anti-freeze at -20°C prior to staining. Every sixth section, from the posterior part of the cortex through the hippocampus, was immunostained with biotinylated 3D6 at 4°C, overnight. The sections were then incubated with horseradish peroxidase avidin-biotin complex (Vector) and developed using 3,3'-diaminobenzidine (DAB) as the

20 chromogen.

B. Results.

1. A β and APP Assays.

The FLAPP+APP α assay recognizes secreted APP including the first 12 amino acids of A β . Since the reporter antibody (2H3₁₋₁₂) is not specific to

25 the alpha clip site occurring between A β amino acids 16 and 17 (Esch *et al.*), this assay also recognizes full length APP. Preliminary experiments using immobilized APP antibodies to the cytoplasmic tail of full length APP to deplete the mixture suggest that approximately 30 to 40% of the FLAPP+APP α is full length. The APP β assay recognizes only the APP

clipped immediately amino-terminal to the A β region due to the specificity of the polyclonal reporter antibody 192 (Seubert *et al.* (1993)).

The specific nature of the A β immunoreactivity was further characterized as follows. Guanidine homogenates of brain (excluding cerebellum and brain stem) were subjected to size exclusion chromatography (Superose 12) and the resulting fractions analyzed using the total A β assay. Comparisons were made of 2, 4, and 12 month transgenic mouse brain homogenates and a non-transgenic mouse brain homogenate to which A β_{1-40} had been spiked at a level roughly equal to that found in the 12 month old transgenic mice. The elution profiles of the transgenic brain homogenate were similar in that the peak fractions of A β immunoreactivity occurred in the same position, a single broad symmetric peak which was coincident with the immunoreactive peak of spiked A β_{1-40} . Attempts were then made to immunodeplete the A β immunoreactivity using resin bound antibodies against A β (mAb 266 against A β_{13-28}), the secreted forms of APP (mAb 8E5 against APP₄₄₄₋₅₉₂ of the APP695 form), the carboxy-terminus of APP (mAb 13G8 against APP₆₇₆₋₆₉₅ of the APP695 form), or heparin agarose. Only the 266 resin captured A β immunoreactivity, demonstrating that full length APP or carboxy-terminal fragments of APP are not contributing to the A β measurement. The A β_{1-42} ELISA employs a capture antibody that recognizes A β_{1-42} but not A β_{1-40} . The A β_{1-42} assay, like the total A β assay, is not affected by the full length or carboxy-terminal forms of APP containing the A β region in the homogenates as shown by similar immunodepletion studies.

2. Total A β and APP Measures.

Table 5 shows the levels of total A β , FLAPP+APP α , and APP β in the hippocampus, cortex, cerebellum, and thalamus of transgenic mice as a function of age. Each data point represents the mean value for each age group. The relative levels of FLAPP+APP α and APP β in all four brain regions remain relatively constant over time. The hippocampus expresses the

highest levels of FLAPP+APP α and APP β followed by the thalamus, cortex, and cerebellum, respectively. In the hippocampus, the levels of FLAPP+APP α are approximately 3.5 to 4.5-fold higher than APP β at all ages. The mean value of all ages for FLAPP+APP α and APP β assays in the hippocampus were 674 (± 465) pmoles/gram and 175 (± 11) pmoles/gram, respectively. From this it can be estimated that the pool of brain APP consists of approximately 50% APP α , 30% full length APP, and 20% APP β .

TABLE 5. PDAPP Transgene Cohort Animal Data
Total A β & APP Measures in pmoles/gram of Brain Tissue.

AGE IN MONTHS	A β & APP FORM	CEREBELLUM	HIPPOCAMPUS	CORTEX	THALAMUS
2	A β	4.03 \pm 1.08 (n=8)	35.41 \pm 6.38 (n=8)	14.25 \pm 2.27 (n=8)	6.41 \pm 1.59 (n=8)
2	FLAPP+APP α	ND	ND	ND	ND
2	APP β	ND	ND	ND	ND
4	A β	4.10 \pm 0.61 (n=14)	38.08 \pm 6.51 (n=14)	15.95 \pm 2.60 (n=14)	7.60 \pm 1.52 (n=14)
4	FLAPP+APP α	395 \pm 120 (n=14)	703 \pm 106 (n=14)	446 \pm 70 (n=14)	6.37 \pm 166 (n=14)
4	APP β	78 \pm 38 (n=14)	198 \pm 30 (n=14)	126 \pm 23 (n=14)	70 \pm 17 (n=14)
6	A β	4.55 \pm 1.38 (n=10)	87.48 \pm 30.33 (n=10)	30.19 \pm 8.33 (n=10)	8.34 \pm 2.40 (n=10)
6	FLAPP+APP α	403 \pm 77 (n=10)	694 \pm 107 (n=10)	506 \pm 97 (n=10)	670 \pm 156 (n=10)
6	APP β	51 \pm 87 (n=10)	194 \pm 35 (n=10)	129 \pm 25 (n=10)	56 \pm 33 (n=10)
6.5	A β	5.42 \pm 1.08 (n=10)	133.63 \pm 57.10 (n=10)	33.27 \pm 12.19 (n=10)	8.83 \pm 1.19 (n=10)

6.5	FLAPP+ APPα	346 \pm 74 (n=10)	580 \pm 115 (n=10)	436 \pm 63 (n=10)	553 \pm 123 (n=10)
6.5	APPβ	27 \pm 77 (n=10)	169 \pm 41 (n=10)	108 \pm 16 (n=10)	58 \pm 22 (n=10)
7	Aβ	4.44 \pm 0.56 (n=10)	200.77 \pm 94.68 (n=10)	60.55 \pm 27.13 (n=10)	8.94 \pm 1.19 (n=10)
7	FLAPP+ APPα	378 \pm 70 (n=10)	656 \pm 73 (n=10)	469 \pm 62 (n=10)	604 \pm 107 (n=10)
7	APPβ	56 \pm 52 (n=10)	176 \pm 27 (n=10)	101 \pm 20 (n=10)	56 \pm 28 (n=10)
7.5	Aβ	5.14 \pm 1.39 (n=10)	461.35 \pm 345.95 (n=10)	81.839 \pm 53.00 (n=10)	10.84 \pm 5.22 (n=10)
7.5	FLAPP+ APPα	362 \pm 54 (n=10)	554 \pm 77 (n=10)	409 \pm 44 (n=10)	503 \pm 80 (n=10)
7.5	APPβ	20 \pm 58 (n=10)	168 \pm 27 (n=10)	118 \pm 21 (n=10)	57 \pm 22 (n=10)
8	Aβ	4.42 \pm 0.73 (n=13)	635.52 \pm 302.45 (n=13)	128.68 \pm 62.80 (n=13)	10.87 \pm 3.39 (n=13)
8	FLAPP+ APPα	386 \pm 52 (n=13)	660 \pm 102 (n=13)	494 \pm 87 (n=13)	672 \pm 150 (n=13)
8	APPβ	64 \pm 77 (n=13)	174 \pm 27 (n=13)	102 \pm 26 (n=13)	57 \pm 30 (n=13)
8.5	Aβ	5.54 \pm 1.11 (n=10)	633.11 \pm 363.14 (n=10)	118.39 \pm 59.91 (n=10)	13.96 \pm 7.34 (n=10)
8.5	FLAPP+ APPα	439 \pm 79 (n=10)	764 \pm 114 (n=10)	558 \pm 80 (n=10)	750 \pm 132 (n=10)
8.5	APPβ	28 \pm 59 (n=10)	185 \pm 34 (n=10)	108 \pm 42 (n=10)	47 \pm 28 (n=10)
9	Aβ	5.52 \pm 1.11 (n=10)	1512.39 \pm 624.286 (n=10)	254.83 \pm 105.927 (n=10)	19.46 \pm 8.99 (n=10)
9	FLAPP+ APPα	500 \pm 112 (n=10)	763 \pm 125 (n=10)	549 \pm 78 (n=10)	815 \pm 167 (n=10)
9	APPβ	4 \pm 83 (n=10)	169 \pm 25 (n=10)	121 \pm 32 (n=10)	49 \pm 26 (n=10)

10	Aβ	4.04 \pm 1.02 (n=11)	2182.21 \pm 1194.49 (n=11)	343.49 \pm 165.531 (n=11)	15.46 \pm 13.38 (n=11)
10	FLAPP+ APPα	452 \pm 130 (n=11)	678 \pm 93 (n=11)	491 \pm 102 (n=11)	693 \pm 166 (n=11)
10	APPβ	52 \pm 32 (n=11)	159 \pm 22 (n=11)	87 \pm 15 (n=11)	46 \pm 10 (n=11)
12	Aβ	3.26 \pm 0.35 (n=9)	4356.23 \pm 1666.44 (n=9)	691.17 \pm 360.93 (n=9)	18.08 \pm 13.50 (n=9)
12	FLAPP+ APPα	385 \pm 166 (n=10)	638 \pm 272 (n=10)	444 \pm 171 (n=10)	708 \pm 278 (n=10)
12	APPβ	41 \pm 29 (n=10)	134 \pm 47 (n=10)	76 \pm 31 (n=10)	35 \pm 19 (n=10)

ND = not determined

In contrast to APP levels, A β levels increased dramatically with age in the hippocampus and cortex. However, no such increase was noted in the cerebellum of the PDAPP transgenic mice, and only a moderate increase was seen in thalamus (Table 5). The increase of A β is greater in the

5 hippocampus relative to the cortex, which also correlates with the 3D6 immunohistochemical results (see discussion below). Compared to the cortex levels of 4 month old mice, A β levels increase 10-fold by 8 months of age and 41-fold at 12 months old (660 \pm 380 pmoles A β /gram tissue at age 12 months). The corresponding increases in A β observed in hippocampus are

10 even more impressive, as the 8 month value is 15 times that at 4 months old and increases to 106-fold at 12 months old (4,040 \pm 1750 pmoles A β /g tissue at 12 months). At 12 months of age, A β constitutes approximately 1% of protein in hippocampus of the PDAPP mice.

To see if the dramatic rise in brain A β concentration is due to

15 amyloid deposition, we next visualized A β deposits immunohistochemically, using the opposite hemisphere of the same mice used for A β measurements. Notably, a parallel increase in A β plaque burden and A β level exists. These

findings strongly argue that the rise in brain A β concentration determined by ELISA is due to the age-dependent amyloidosis.

3. A β_{1-42} Measures in Transgenic Mouse Brain.

Concentrations of A β_{1-42} in the cortex of transgenic mice were evaluated at different ages. As shown in Table 6, the percentage of A β which is A β_{1-42} in the cortex of transgenic mice, also increases with age. The ELISA data suggest that A β_{1-42} is preferentially depositing in the transgenic mice, and that the deposits detected by mAb 3D6 immunostaining are primarily A β_{1-42} .

Table 6. A β_{1-42} Levels in the Cortex of Transgenic Brain.

Age (months)	A β_{1-42} (pmoles/g)
4	4.71
8	75.65
10	247.43
12	614.53

4. A β Immunostaining in PDAPP Transgenic Brain.

Transgenic animals with A β values representing the mean A β value of the age group were used for 3D6 immunostaining. A progression of A β deposition is seen in the 4, 8, 10, and 12 months old animals. At four months of age, transgenic brains contained small, rare punctate deposits, 20 μ m in diameter, that were only infrequently observed in the hippocampus and frontal and cingulate cortex. By eight months of age, these regions contained a number of thioflavin-positive A β aggregates that formed plaques as large as 150 μ m in diameter. At ten months of age, many large A β deposits were found throughout the frontal and cingulate cortex, and the molecular layers of the hippocampus. The outer molecular layer of the dentate gyrus receiving perforant pathway afferents from the entorhinal cortex was clearly delineated by A β deposition. This general pattern was more pronounced by heavier A β

deposition at one year of age. The anatomical localization of A β deposition is remarkably similar to that seen in Alzheimer's disease.

C. Discussion.

A β amyloidosis is an established diagnostic criteria of Alzheimer's disease (Mirra *et al.*, *Neurology* 41:479-486 (1991)) and is consistently seen in higher cortical areas as well as the hippocampal formation of the brain in affected subjects. It is believed that A β amyloidosis is a relatively early event in the pathogenesis of AD that subsequently leads to neuronal dysfunction and dementia through a complex cascade of events (Mann *et al.*, *Neurodegeneration* 1:201-215 (1992); Morris *et al.*, *Neurology* 46:707-719 (1996)). Various pathways of APP processing have been described (reviewed in Schenk *et al.*, *J. Med. Chem.* 38:4141-4154 (1995)) including the major α -secretase pathway where cleavage of APP occurs with A β (Esch *et al.*) and the amyloidogenic or β -secretase pathway where cleavage of APP occurs at the N-terminus of A β (Seubert *et al.* (1993)). Further cleavage of APP leads to the constitutive production of A β forms including those ending at position 40 (A β ₁₋₄₀) or 42 (A β ₁₋₄₂). ELISAs that detect specific APP products arising from these individual pathways in the PDAPP mouse brain allow determination of whether differential processing of APP contributes to the regional or temporal specificity of amyloid formation and deposition.

A β amyloid deposition seen in the PDAPP mouse brain is highly age and region specific. Amyloid deposition begins at around 7 months of age, and by 12 months of age, amyloid deposition is very profound throughout the hippocampus and in the rostral region of the cortex. The age dependent increases in amyloid deposition correlate well with the dramatic rise in A β levels in these brain regions as measured by ELISA assay. An increase in A β is measurable by 7 months of age and by 10 months the hippocampus as 2180 pmoles/g of A β , a concentration equivalent to that of my cytoskeletal

proteins and comparable to the levels found in the cortex of human AD brain (Gravina *et al.*, *J. Biol. Chem.* 270:7013-7016 (1995)). A β levels in the cerebellum, an unaffected brain region, still are at 4 pmoles/g -- essentially unchanged relative to the levels at 4 months of age -- again correlating with amyloid deposition measured by histological analyses. These results indicate that in aged PDAPP mice, brain A β levels reflects amyloid burden and therefore direct immunoassay measurement of brain A β levels can be used to test for compounds that reduce amyloid plaque burden.

In the PDAPP mouse, individuals suffering Down's Syndrome, and individuals with certain forms of FAD, overproduction of A β is almost certainly an accelerating factor not only in A β deposition but in subsequent neuropathology (Citron *et al.*, Mann *et al.*, Miller *et al.*, *Archives of Biochem. Biophys.* 301:41-52 (1993)). A comparison of the A β measurements seen in the PDAPP mouse with those reported for AD brain tissue reveals several striking similarities. For example, in the PDAPP mouse, the relative levels of A β peptide in hippocampus from young (2 months of age) versus old (10 months of age) mice is nearly a hundred fold. Similar findings were noted by Gravina *et al.* in comparing control brain tissue relative to that of AD. The rise in brain A β levels in the PDAPP mouse is rather pronounced between the ages of six to nine months of age. Again, this timecourse parallels, in an accelerated manner, that seen in Down's Syndrome brain tissue, where amyloid deposition begins at approximately 30 years of age and increases substantially until approximately age 60 (Mann).

In summary, the above results show that a reproducible increase in measurable A β occurs in the brain tissue of the PDAPP mice and that this increase correlates with the severity of amyloid deposition. These findings

indicate that these mice can be used to identify agents or compounds that pharmacologically reduce A β peptide production or affect its deposition.

Example 9: Behavioral Differences in PDAPP Transgenic Mice.

5 Alzheimer's disease is characterized by cognitive deficits including memory loss, and impairment of memory functions. To determine if the disclosed transgenic mice exhibit similar deficits, transgenic (TG) and non-transgenic (nTG) mice were evaluated for task performance in three types of maze apparatus used to test working and reference memory; the Y maze, the
10 radial arm maze (RAM), and the water maze. The transgenic mice tested represent the fifth generation derived from the PDAPP mice described in Example 6. The Y maze and the radial arm maze are used to assess spontaneous alternation which is a function of working memory. For the Y maze task, the mouse is placed in the stem of a Y maze twice, each time
15 allowing a choice entry into one of the arms. Entering both arms is a successful alternation, requiring memory of the previously entered arm, while entering the same arm on both trials is a failure. Chance performance is 50% alternation, that is, 50% of the mice alternate.

For the radial arm maze task, the mouse is placed at the center of a
20 maze with multiple arms radiating from the center. In the testing described below, a radial eight-arm maze was used. Alternation performance is measured by allowing only eight entries, with the number of different arms entered being the measure of performance. The number of different-arm entries can be compared to the number of different-arm entries expected by
25 chance, which is 5.25 (Spetch and Wilkie, "A program that stimulates random choices in radial arm mazes and similar choice situations" *Behavior Research Methods & Instrumentation* 12:377-378 (1980)). Performance above chance, that is, above 5.25, requires memory of the previously entered arms.

The water maze used for the tests described below consists of a pool of water in which a submerged platform is placed. This hidden platform (HP) can be found by swimming mice either by chance (first trial) or through memory of positional clues visible from the tank (subsequent trials). Subject mice were trained in the hidden platform task according to standard procedures. Briefly, mice were first pretrained in a small pool (47 cm diameter, 20 cm platform), which teaches them how to navigate in water, that the platform is the goal, that there is no other escape, and that to find it they must resist their natural inclination to stay along the sides of the pool. They were then trained to find a single platform position in the hidden platform task using a larger pool and smaller platform (71 cm pool, 9 cm platform).

During the HP task, visual cues were located inside the pool (intramaze cues; black pieces of cardboard - circle, plus, or horizontal lines - located in three quadrants at the top of the wall, which was 38 cm high above water level), and various room cues were visible outside the pool (extramaze cues).

The mice assigned to the characterization cohort study were tested on the behavioral tasks described above over 3 days during the week or two before euthanasia. Their transgenic status was not known to the tester. Non-transgenic littermates were used for comparison. Each morning the subject mice were run in the Y maze and RAM as described above. They were then tested for general strength on the inclined plane (INP) test. For this, mice were placed in a 10-cm-wide runway lined with ridged plastic and elevated with the head up at 35°. The angle was then increased until the mouse slid off, and the angle was recorded. This was repeated three times each day. The average scores for the three days were calculated for each mouse for the Y maze (0=repeat, 1=alternate), RAM (number of different arms and time to finish, 10 minute limit), and INP (average of all nine trials). General

activity was also rated on the first day of testing. Each mouse was observed in the cage, and picked up and held. A mouse that remained calmly in the hand was scored 1, with progressively greater activity and reaction to handling scored up to 4.

5 Following the above tests each day, mice were tested in the water maze as described above. Briefly, mice were pretrained in a small pool to climb on a large submerged platform as their only means of escape from the water. They were then given six blocks of four trials each to learn the location of a small platform in a large pool. For analysis, all four trials
10 within each block were averaged. The exception was the first hidden platform block, for which only the last three trials were averaged. The first trial was analyzed separately, because it is the only one for which platform location could not be known, and thus did not relate to spatial learning. It is thus used as a control for non-spatial factors, such as motivation and
15 swimming speed. The performance effects between blocks were analyzed as a repeated measure for the hidden platform task. Standard analysis of variance (ANOVA) calculations were used to assess the significance of the results.

20 Results in the RAM show that TG performed significantly worse than nTG across all ages (Group effect: $p=0.00006$). The time to finish was also significantly different between TG and nTG mice (Group effect: $p=0.005$). The correlation between the time to finish and the number of arms chosen was small ($R = -0.15$, $p=0.245$ in each group). This suggests that the consistent impairment in the RAM is not accounted for by the increased time
25 to complete the task taken by TG mice. Results in the Y maze were also significantly different for TG and nTG mice (Group effect: $p=0.011$). Validation studies performed on non-transgenic mice indicate that the Y maze is a less sensitive measure than the RAM.

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ability unrelated to cognitive ability), and the amount of time need to find the platform (a measure of the combination of both the distance travelled and the swimming speed). This was done in older and younger mice than reported above, using three trials per block and no pretraining. The time needed to
5 find the platform was significantly different in TG and nTG mice (Group effect: $p < 0.0005$), with the TG mice taking longer. However, the swimming speed was not significantly different between TG and nTG mice (Group effect: $p = 0.879$). Thus, the difference in time needed to find the platform is likely to be due to a cognitive difference between TG and nTG
10 mice. This is confirmed by measures of the distance travelled to find the platform. The TG mice travelled significantly further than the nTG mice before reaching the platform (Group effect: $p < 0.0005$). These results indicate that the differences seen between TG and nTG mice in the time to reach the platform in the water maze tasks are due to differences in cognitive
15 ability.

To test whether nTG mice retain a better memory of the platform location than TG mice, a probe trial was given immediately following hidden platform training in which the platform was removed. Video tracking was used to determine the number of crossings of the former platform location
20 made by the mice relative to crossings of non-platform locations. There was a significant difference seen between the relative crossings of TG and nTG mice (Group effect: $p = 0.006$). This is evidence that the nTG mice remember the former location of the platform better than TG mice.

It was also possible that the difference observed between TG and nTG
25 mice in the time needed to reach the platform could have been influenced by differences in perception of the cues or motivational differences. To test this, TG and nTG mice were subjected to visible platform tasks in the water maze. For these tasks, a platform was placed in the pool so that it was visible above the water. Three different platforms were tested, a dark

platform 25 mm above the surface (most visible), a gray platform 25 mm above the surface, and a dark platform 5 mm above the surface (both less visible). The results show no difference in the time to find the most visible platform between TG and nTG mice (Group effect: $p=0.403$). There was not any greater decrease in performance in TG mice when less visible platforms were used, suggesting that their vision was as good as nTG mice. These results indicate that perceptual and motivational differences do not influence the time to reach the platform in the water maze tasks described above.

- Performance differences between TG and nTG mice were shown for RAM, Y maze, and water maze cognitive tasks in mice aged 4 to 8 months (2 to 12 months for the water maze). All of these differences indicate, and are consistent with, cognitive deficits in the transgenic mice as a group. The various tasks combined to test working memory and reference memory, both of which are implicated in cognitive impairment observed in Alzheimer's victims.

Example 10: Detection and Measurement of Alzheimer's Disease Markers.

A. Detection and Measurement of GFAP.

- Glial fibrillary acidic protein (GFAP), a marker which increases in AD brain tissue, was measured in the following manner. Tissue extracts were prepared from hippocampi of control and PDAPP transgenic mice, as described in Example 6, aged 14 months. Tissue was sonicated in 10 volumes (v/w) of 10 mM Tris, pH 7.5, 1 mM EDTA, 1 mM EGTA, 0.1 mM PMSF, 10 $\mu\text{g/ml}$ leupeptin, 5 $\mu\text{g/ml}$ calpain inhibitor 1. Protein determinations were made on the extract and SDS-PAGE sample buffer added before boiling the samples for 5 minutes. SDS-PAGE was performed using 12.5 μg of protein of each sample loaded onto 10% Tris-glycine gels

(Novex). The proteins were transferred to Pro-Blot PVDF membranes by standard methods. GFAP immunoreactive proteins were detected using an anti-GFAP antibody from Sigma (G9269) used at a dilution of 1:2,000. An increase in immunoreactivity in general was observed, and a smaller anti-GFAP reactive species was also found to increase substantially, in the transgenic animals. In non-transgenic animals, this approximately 40 kD fragment gave a mean densitometer signal of 142.47, while in the transgenic animals, it gave a mean densitometer signal of 591.51. This difference was significant, with a P value of 0.0286.

B. Detection of Gliosis.

Gliosis is one of the changes that is associated with the neuropathology of Alzheimer's disease. The isoquinoline carboxamide PK 11195 has been shown to be a preferential marker of the peripheral benzodiazepine sites associated with gliosis. These sites have been shown to be enhanced in several diseases and animal models associated with neuronal damage and activated microglia including stroke (Stephenson *et al.*, J. Neuroscience 15:5263-5274 (1991)) and Alzheimer's disease (Diorio *et al.*, Neurobiology of Aging 12:255-258 (1991)). In particular Diorio and colleagues have shown an approximate 200% increase in [³H] PK 11195 binding in some brain regions of AD patients, such as the temporal cortex compared with age-matched controls. In this example, the brains from the PDAPP mouse, described in Example 6, were examined for qualitative and quantitative changes in the binding of [³H] PK 11195 in order to correlate with the previously described AD disease pathology. Two different approaches were utilized; radioreceptor binding to homogenates of different brain regions and receptor autoradiography.

1. Methods.

For the homogenate binding studies, PDAPP mice were euthanised by cervical dislocation and the brains rapidly dissected on ice. Homogenates (10 mg/ml wet weight) of cerebral cortex, hippocampus and cerebellum were prepared in 50 mM Tris HCl, pH 7.4. 0.3, 1.0 and 3.0 nM [3 H] PK 11195 was incubated with these brain regions for 60 minutes at 23°C followed by rapid filtration over Whatman GF/B filters using a Brandell cell harvester. Non-specific binding was determined using 1 μ M unlabelled PK 11195. Quantitation was performed by liquid scintillation spectrometry.

In the autoradiographic studies, PDAPP mice were euthanised using carbon dioxide, the brains removed and snap frozen in methyl butane/dry ice. The brains were sectioned in the coronal plane through the hippocampus. Twenty micron thick sections were mounted on glass slides and stored at -20°C. Sections were incubated at 1 hour at 23°C in 170 mM Tris-HCl, pH 7.4 containing 1 nM [3 H] PK 11195. Non-specific binding was determined using 1 μ M unlabelled PK 11195. Incubations were terminated by rinsing sections twice for 5 minutes in ice-cold incubation buffer followed by a brief wash in ice-cold distilled water. Following rapid drying, sections were exposed to tritium Hyperfilm (Amersham International) for up to 5 weeks.

2. Results.

Four heterozygous transgenic mice 3-4 months of age were evaluated in the homogenate binding studies and compared with litter-mate controls. No significant differences were observed between any of the brain regions of the transgenic animals and their respective controls. [3 H] PK 1119 autoradiography was performed to compare binding in a 12-month old heterozygotic transgenic mouse with an aged-matched non-transgenic control. Preliminary results from an autoradiogram exposed for five weeks indicated that several plaque-like structures were labeled in the retrosplenial cortex of the transgenic mouse, a region that invariably contains A β deposits. The

pattern of labeling corresponded to microglial cell or astrocytic clumps associated with plaques, rather than the more widespread pattern of astrocytosis or microgliosis in the hippocampal and cortical parenchyma. The non-transgenic mouse did not show this labeling pattern.

- 5 No changes were observed in the 3-4 month animals but some evidence for an increase in [^3H] PK 11195 binding was seen in the 12-month animal.

C. Detection and Measurement of Cholinergic Nerve Terminals.

- 10 A population of cholinergic neurones projecting to the forebrain have been shown to be selectively decreased in the postmortem brains of patients diagnosed with Alzheimer's disease. Hemicholinium-3 is a potent inhibitor of high affinity choline uptake and has been shown to be a good marker of cholinergic nerve terminals (Pascual *et al.*, *J Neurochem* 54:792-800 (1990)).
- 15 The total number of high affinity choline uptake sites in PDAPP transgenic animals, which are described in Example 6, has been measured using both crude whole-brain preparations and homogenates from selective brain regions using the selective ligand [^3H]-Hemicholinium-3 ([^3H]HCh-3).

- 20 [^3H]-Hemicholinium-3 binding was determined using a modification of the methods described in Pascual *et al.* Mice were euthanised by asphyxiation with carbon dioxide and the brains rapidly removed and dissected on ice. The cortex, cerebellum, striatum and hippocampus were homogenized in 5 ml of 10 mM phosphate buffer without NaCl. Samples were spun at 17,000 x g for 10 minutes, and the pellets washed twice in 5 ml 10 mM PO_4 buffer. The final pellet was resuspended in 5 ml 1X phosphate
- 25 buffered saline (PBS) to produce a protein concentration of 0.5 mg/ml. Brain regions were assayed in triplicate for high affinity choline uptake sites by the addition of [^3H]HCh-3 (3 nM final concentration). Following a 20 minute incubation, assays were terminated by rapid filtration through Whatman GF/B filters using a Brandell cell harvester and washing with PBS.

Filters were transferred into scintillation vials, and specific binding estimated by liquid scintillation spectrometry.

D. Detection and Measurement of Sodium-Potassium ATPase.

Ouabain has been shown to bind specifically to high affinity sites in mammalian brain and that these sites correspond to a neuronal form of sodium-potassium ATPase (Na/K-ATPase; Hauger *et al.*, *J Neurochem* 44:1709-1715 (1985)). These sites have been shown to decrease in animal models of neurodegenerative diseases. Alzheimer's disease is characterized by massive neurodegeneration (DeLacoste and White, *Neurobiology of Aging* 4(1):1-16 (1993)).

In order to estimate the extent of neurodegeneration in the PDAPP mouse the binding of ouabain was determined in mouse brain homogenates. Methods were adapted from those described by Hauger *et al.* Brain tissue was homogenized in 100 mM Tris HCl, pH 7.4 containing 200 mM NaCl and 10 mM MgCl₂ and resuspended in assay buffer to produce a final concentration of 100 μ g protein per assay. Specific binding was determined with 1 to 200 nM [³H] ouabain in a solution of 5 mM ATP, 100 mM Tris HCl, pH 7.4, 10 mM MgCl₂, and 200 mM NaCl, incubated for 30 minutes at 37°C. Non specific binding was determined in the presence of 100 mM ouabain and the absence of ATP. Assays were terminated by rapid filtration over Whatman GF/B filters. Tubes were washed with ice cold 50 mM Tris HCl, pH 7.4, 15 mM KCl, 5 mM MgCl₂. Filters were transferred into scintillation vials, and specific binding estimated by liquid scintillation spectrometry.

Measurements of Na/K-ATPase and Mg-ATPase activity in brain tissue of PDAPP transgenic mice of various ages and in non-transgenic control brain tissue. These results show some significant differences in activity between transgenic and non-transgenic samples in older mice. Mouse brain homogenates from 4, 8, and 12 month old PDAPP transgenic

(TG) mice, and from non-transgenic (nTG) mice, were prepared and assayed generally as described above. The activity of Mg-ATPase was also determined. The results are shown in Tables 7 and 8.

**Table 7. Na/K-ATPase Activity
in PDAPP Transgenic Mouse Brain.**

Age	Tissue	Na/K-ATPase	
		rate (pmole Pi/mg protein/min)	% of nTG
5	4 TG hippocampus	2.57 ± 0.62	88 ± 14
	4 nTG hippocampus	2.53 ± 0.51	-
	4 TG cortex	0.57 ± 0.13	72 ± 9
	4 nTG cortex	0.77 ± 0.17	-
	4 TG cerebellum	1.39 ± 0.17	85 ± 14
	4 nTG cerebellum	2.14 ± 0.53	-
15	8 TG hippocampus	4.63 ± 1.72	153 ± 53
	8 nTG hippocampus	2.87 ± 0.41	-
	8 TG cortex	1.16 ± 0.08	121 ± 23
	8 nTG cortex	1.15 ± 0.20	-
	8 TG cerebellum	2.74 ± 0.81	183 ± 53
	8 nTG cerebellum	1.47 ± 0.13	-
20	12 TG hippocampus	1.66 ± 0.36	58 ± 9
	12 nTG hippocampus	3.11 ± 0.94	-
	12 TG cortex	1.45 ± 0.40	109 ± 17
	12 nTG cortex	1.60 ± 0.46	-
	12 TG cerebellum	1.43 ± 0.32	74 ± 7
	12 nTG cerebellum	2.04 ± 0.64	-

**Table 8. Mg-ATPase Activity
in PDAPP Transgenic Mouse Brain.**

Age	Tissue	Mg-ATPase	
		rate (pmole Pi/mg protein/min)	% of nTG
5	4 TG hippocampus	2.83±0.38	110±14
	4 nTG hippocampus	2.44±0.54	-
	4 TG cortex	1.74±0.14	92±5
	4 nTG cortex	1.87±0.18	-
10	4 TG cerebellum	2.58±0.44	100±12
	4 nTG cerebellum	2.59±0.35	-
	8 TG hippocampus	3.28±0.69	99±22
	8 nTG hippocampus	3.32±0.39	-
	8 TG cortex	1.72±0.11	73±6
15	8 nTG cortex	2.40±0.31	-
	8 TG cerebellum	3.19±0.49	113±15
	8 nTG cerebellum	2.77±0.23	-
	12 TG hippocampus	1.48±0.21	65±7
	12 nTG hippocampus	2.33±0.46	-
20	12 TG cortex	1.60±0.30	76±7
	12 nTG cortex	2.06±0.40	-
	12 TG cerebellum	1.61±0.26	78±8
	12 nTG cerebellum	1.93±0.99	-

The difference in Na/K-ATPase activity between transgenic and non-transgenic tissue is significant ($p < 0.05$) in the case of 12 month old cerebellum, and is highly significant ($p < 0.01$) in the case of 12 month old hippocampus. The difference in Mg-ATPase activity between transgenic and non-transgenic tissue is significant ($p < 0.05$) in the case of 8 and 12 month

old cortex, and is highly significant ($p < 0.01$) in the case of 12 month old hippocampus.

E. *In situ* Hybridization with Probes to Neurotrophic Factors.

The use of *in situ* hybridization to detect and localize mRNAs for specific gene products is well documented in the literature (Lewis *et al.*, *Molecular Imaging in Neuroscience: A Practical Approach* (New York, Oxford University Press. 1st ed., 1-21, 1993), Lu and Gillett, *Cell Vision* 1(2):169-176 (1994), Sirinathsinghji and Dunnett, *Molecular Imaging in Neuroscience: A Practical Approach* (New York, Oxford University Press. 1st., ed. 43-67, 1993), Lawrence and Singer, *Nuc. Acids Res.* 13:1777-1799 (1985), Zeller and Rogers, *Current Protocols in Molecular Biology* (New York, John Wiley and Sons. 14.3.1-14.5.5, 1995)). For illustrative purposes, the specific example described below utilizes ^{35}S -radiolabeled oligodeoxyribonucleotide probes to detect BDNF mRNA in cryostat sectioned mouse brain from the PDAPP transgenic mouse described in Example 6 and non-transgenic control mice. However, ^{33}P -labeled radioactive DNA probes, as well as *in vitro* transcribed complementary RNA probes, could be used as well. Non radioactive probe labeling methods may also be used (Knoll, *Current Protocols in Molecular Biology* (New York, John Wiley and Sons. 14.7.1-14.7.14, 1995)). Additionally, the choice of tissue pre-treatment for hybridization with probe (for example, paraffin embedded sections) and post-hybridization washes depend on the method used, examples of which are described in the references cited above. Known and appropriate precautions against RNase contamination should be employed and are also discussed in the above references.

1. Tissue Preparation.

Freshly dissected whole brains, or sub-regions of interest, from transgenic or control mice at various developmental stages, or post-natal

ages, are snap frozen in isopentane pre-equilibrated to -70°C . If desired, the animals may be perfused with PBS to eliminate circulating cells from brain prior to dissection. The brains are removed following 15 to 20 seconds immersion in isopentane, wrapped in aluminum foil, labeled appropriately, and stored at -80°C for sectioning. It should be noted that although the signal from *in situ* hybridization to cryostat sectioned tissues is more sensitive than to paraffin embedded sections, it is dependent upon the time from dissection to hybridization. Frozen tissue is preferably analyzed by hybridization with probe within six weeks. RNA integrity in tissues declines beyond this time. Thus, if longer time periods between dissection and analysis are anticipated, the tissue should be fixed (see, for example, Lu and Gillett) before long term storage at -80°C .

Prior to sectioning, Probe-On-Plus glass slides (Fisher Scientific, Pittsburgh, PA) can be made RNAase free by overnight soaking in absolute ethanol, air dried briefly in a dust free environment, and baked at 180°C for a minimum of 4 hours. After cooling to room temperature, the slides are coated with 0.01% poly-lysine (prepared in DEPC treated H_2O) for approximately 5 seconds, and air dried in a dust free area. The coated slides can be stored for up to one month before use in a slide box with silica gel or drierite pellets.

For sectioning, the frozen brain stored at -80°C is transferred to a cryostat at -20°C , mounted onto a sectioning block, embedded in OCT.[®], and allowed to equilibrate. The tissue is then cut into 7 to 14 μm thick sections using a sterilized microtome knife (treated with 70% EtOH in DEPC H_2O), and thaw mounted onto poly-lysine coated slides. The slides are kept at -20°C until the sectioning is complete. The sections are fixed and dehydrated by immersing the slides sequentially in the solutions noted below.

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2. Probe Design and Preparation.

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For radiolabeling, the probes are gel purified on denaturing acrylamide gels and reconstituted in H₂O using standard protocols (Sambrook *et al.*). The probes (30 to 35 ng, 2 pmoles) are labeled by 3' homopolymeric tailing using terminal deoxynucleotidyl transferase (Promega, Madison WI) and ³⁵S-dATP (1000 Ci/mmol, Amersham Inc.) according to the enzyme manufacturer's recommendation. The radiolabeled probes are purified by column chromatography on size exclusion mini-spin columns (Biospin-6, Biorad Inc., Hercules, CA). The specific activity of the probes is quantitated by scintillation counting. Typical specific activities of the probes ranged from 1 X 10⁹ to 5 X 10⁹ cpm/μg.

In preparation for hybridization, the desired number of slides are removed from storage under alcohol, and allowed to air dry thoroughly in the slide rack (approximately 1 hour). Meanwhile, the probe is heat denatured in a boiling H₂O bath for 2 to 5 minutes, quick chilled in an ice/H₂O bath, and diluted in hybridization buffer (10% dextran sulfate, 50% deionized formamide, 4X SSC, 5X Dehardts, 100 µg/ml sheared salmon sperm DNA, 100 µg/ml polyadenylic acid) to a final concentration of 5 X 10³ to 10 X 10³ cpm/µl. DTT is added to a 10 mM final concentration.

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paper towels), saturated in 4X SSC, 50% formamide, and humidified by pre-incubating them with closed lids in a 42°C incubator for 1 to 3 hours before the slides are placed inside them. Although cover slips can be placed on the sections after the hybridization buffer is applied, this is not necessary

5 provided the hybridization chambers are adequately humidified during the procedure. If pre-hybridization is used to obtain a lower background, the sections may be incubated at 42°C under 50 μ l hybridization buffer (minus probe) per section for 1 to 2 hours. After this time, an equal volume of hybridization buffer containing probe at twice the concentration described

10 above is applied to each section, and hybridization is carried out as described above.

For washes, the slides are transferred from the hybridization chamber to a slide holder. The slides can be placed in the slide holder every 4th or 5th slot so as to allow adequate flow of wash solution over the surface of

15 each section. This placement can significantly lowers background on the sections. A moderate flow rate of wash solution over the surface of the sections promotes removal of unhybridized probe, and consequently reduces background. This is best accomplished during the 55°C wash steps by suspending the slides in the slide holder, above a magnetic stir bar. The stir

20 bar is preferably placed approximately one inch under the slide holder. This can be done by hanging the slide holder(s) from pipets straddling the wash chamber. A large beaker, or a 4 to 6 inch deep Pyrex baking dish makes a suitable wash chamber. The wash chamber is placed on a hot-plate stirrer, the temperature setting of which is precalibrated to maintain the wash

25 solution at 55°C during the procedure. The changes of wash solution are made using pre-equilibrated solution. Washes and post wash dehydration of the sections are carried out as follows:

1. twice in 1X SSC at room temperature for 5 minutes each time;
2. three times in 1X SSC at 55°C for 30 minutes each time;

3. once in 1X SSC at room temperature for 1 min.
4. once in 0.1X SSC at room temperature for 15 seconds;
5. once in ultra pure H₂O at room temperature for 15 seconds;
6. once in 50% EtOH for 15 seconds;
- 5 7. once in 70% EtOH for 15 seconds;
8. once in 95% EtOH for 15 seconds;

The sections are air dried thoroughly at room temperature for 2 hours, followed by 30 minutes at 55°C. The dried sections on the slides are placed in autoradiographic cassette and exposed to X-ray film (Hyperfilm, β -max, Amersham Inc., Arlington Heights, IL) at 4°C for 2 to 3 days to estimate the exposure time required under emulsion. The X-ray film is developed according to the manufacturers recommendation. The sections are coated with emulsion (Amersham LM-1, #RPN40) by dipping in emulsion at 42°C under appropriate safelight conditions. The emulsion coated slides are air dried on a cooled surface for approximately 30 minutes, and transferred to a plastic slide box containing a drying agent (drierite pellets). The seams of the box can be sealed with black tape, and the box wrapped in several layers of aluminum foil to ensure a light-tight enclosure. Following 2 to 4 hours at room temperature to finish the drying, the boxes are transferred to 4°C for autoradiographic exposure for 2 to 6 weeks. Prior to developing the emulsion coated slides, the box is removed from the refrigerator and allowed to equilibrate to room temperature for approximately 1 hour. The slides are then developed according to the manufacturers instructions, air dried for 1 to 2 hours, and if desired, counterstained with the appropriate counterstain.

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Modifications and variations of the making and testing of transgenic animal models for testing of Alzheimer's disease will be obvious to those skilled in the art from the foregoing detailed description. Such modifications and variations are intended to come within the scope of the following claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Dora K. Games, Dale B. Schenk, Lisa C. McConlogue,
Peter A. Seubert, and Russell E. Rydel
- (ii) TITLE OF INVENTION: Method For Identifying Alzheimer's Disease
Therapeutics Using Transgenic Animal Models
- (iii) NUMBER OF SEQUENCES: 10
- (iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: Patrea L. Pabst
(B) STREET: 2800 One Atlantic Center
1201 West Peachtree Street
(C) CITY: Atlanta
(D) STATE: GA
(E) COUNTRY: USA
(F) ZIP: 30309-3450
- (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:
- (vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: 08/480,653
(B) FILING DATE: June 7, 1995
(C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Pabst, Patrea L.
(B) REGISTRATION NUMBER: 31,284
(C) REFERENCE/DOCKET NUMBER: ANS101CIP
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: (404)-873-8794
(B) TELEFAX: (404)-873-8795

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2085 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1-2085
(D) OTHER INFORMATION: /function= "coding region for APP695."
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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Ala Leu Glu Val Pro Thr Asp Gly Asn Ala Gly Leu Leu Ala Glu Pro
      20             25             30

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GCC	ACC	ACC	ACC	ACC	ACC	ACC	ACA	GAG	TCT	GTG	GAA	GAG	GTG	GTT	CGA	864
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GTT	CCT	ACA	ACA	GCA	GCC	AGT	ACC	CCT	GAT	GCC	GTT	GAC	AAG	TAT	CTC	912
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	290					295					300					
GAG	ACA	CCT	GGG	GAT	GAG	AAT	GAA	CAT	GCC	CAT	TTC	CAG	AAA	GCC	AAA	960
Glu	Thr	Pro	Gly	Asp	Glu	Asn	Glu	His	Ala	His	Phe	Gln	Lys	Ala	Lys	
305					310				315						320	
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Glu	Arg	Leu	Glu	Ala	Lys	His	Arg	Glu	Arg	Met	Ser	Gln	Val	Met	Arg	
				325				330						335		
GAA	TGG	GAA	GAG	GCA	GAA	CGT	CAA	GCA	AAG	AAC	TTG	CCT	AAA	GCT	GAT	1056
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Asn	Met	Leu	Lys	Lys	Tyr	Val	Arg	Ala	Glu	Gln	Lys	Asp	Arg	Gln	His	
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Arg	Met	Asn	Gln	Ser	Leu	Ser	Leu	Leu	Tyr	Asn	Val	Pro	Ala	Val	Ala	
465					470					475					480	
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TAT	TCA	GAT	GAC	GTC	TTG	GCC	AAC	ATG	ATT	AGT	GAA	CCA	AGG	ATC	AGT	1536
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 Pro Trp His Ser Phe Gly Ala Asp Ser Val Pro Ala Asn Thr Glu Asn
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 CAT CAT CAA AAA TTG GTG TTC TTT GCA GAA GAT GTG GGT TCA AAC AAA 1872
 His His Gln Lys Leu Val Phe Phe Ala Glu Asp Val Gly Ser Asn Lys
 610 615 620
 GGT GCA ATC ATT GGA CTC ATG GTG GGC GGT GTT GTC ATA GCG ACA GTG 1920
 Gly Ala Ile Ile Gly Leu Met Val Gly Gly Val Val Ile Ala Thr Val
 625 630 635 640
 ATC GTC ATC ACC TTG GTG ATG CTG AAG AAG AAA CAG TAC ACA TCC ATT 1968
 Ile Val Ile Thr Leu Val Met Leu Lys Lys Lys Gln Tyr Thr Ser Ile
 645 650 655
 CAT CAT GGT GTG GTG GAG GTT GAC GCC GCT GTC ACC CCA GAG GAG CGC 2016
 His His Gly Val Val Glu Val Asp Ala Ala Val Thr Pro Glu Glu Arg
 660 665 670
 CAC CTG TCC AAG ATG CAG CAG AAC GGC TAC GAA AAT CCA ACC TAC AAG 2064
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 Phe Phe Glu Gln Met Gln Asn
 690 695

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 695 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Leu Pro Gly Leu Ala Leu Leu Leu Leu Ala Ala Trp Thr Ala Arg
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 Gln Ile Ala Met Phe Cys Gly Arg Leu Asn Met His Met Asn Val Gln
 35 40 45
 Asn Gly Lys Trp Asp Ser Asp Pro Ser Gly Thr Lys Thr Cys Ile Asp
 50 55 60
 Thr Lys Glu Gly Ile Leu Gln Tyr Cys Gln Glu Val Tyr Pro Glu Leu
 65 70 75 80
 Gln Ile Thr Asn Val Val Glu Ala Asn Gln Pro Val Thr Ile Gln Asn
 85 90 95
 Trp Cys Lys Arg Gly Arg Lys Gln Cys Lys Thr His Pro His Phe Val
 100 105 110
 Ile Pro Tyr Arg Cys Leu Val Gly Glu Phe Val Ser Asp Ala Leu Leu
 115 120 125
 Val Pro Asp Lys Cys Lys Phe Leu His Gln Glu Arg Met Asp Val Cys
 130 135 140
 Glu Thr His Leu His Trp His Thr Val Ala Lys Glu Thr Cys Ser Glu
 145 150 155 160
 Lys Ser Thr Asn Leu His Asp Tyr Gly Met Leu Leu Pro Cys Gly Ile
 165 170 175
 Asp Lys Phe Arg Gly Val Glu Phe Val Cys Cys Pro Leu Ala Glu Glu
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 210 215 220
 Val Val Glu Val Ala Glu Glu Glu Glu Val Ala Glu Val Glu Glu Glu
 225 230 235 240
 Glu Ala Asp Asp Asp Glu Asp Asp Glu Asp Gly Asp Glu Val Glu Glu
 245 250 255
 Glu Ala Glu Glu Pro Tyr Glu Glu Ala Thr Glu Arg Thr Thr Ser Ile
 260 265 270
 Ala Thr Thr Thr Thr Thr Thr Thr Glu Ser Val Glu Glu Val Val Arg
 275 280 285
 Val Pro Thr Thr Ala Ala Ser Thr Pro Asp Ala Val Asp Lys Tyr Leu
 290 295 300
 Glu Thr Pro Gly Asp Glu Asn Glu His Ala His Phe Gln Lys Ala Lys
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Phe Phe Glu Gln Met Gln Asn
690 695

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2253 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS

(B) LOCATION: 1-2253

(D) OTHER INFORMATION: /function= "coding region for APP751."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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ACC AAG GAA GGC ATC CTG CAG TAT TGC CAA GAA GTC TAC CCT GAA CTG 240
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CAG ATC ACC AAT GTG GTA GAA GCC AAC CAA CCA GTG ACC ATC CAG AAC 288
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85 90 95

TGG	TGC	AAG	CGG	GGC	CGC	AAG	CAG	TGC	AAG	ACC	CAT	CCC	CAC	TTT	GTG	336
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ATT	CCC	TAC	CGC	TGC	TTA	GTT	GGT	GAG	TTT	GTA	AGT	GAT	GCC	CTT	CTC	384
Ile	Pro	Tyr	Arg	Cys	Leu	Val	Gly	Glu	Phe	Val	Ser	Asp	Ala	Leu	Leu	
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GTT CCT GAC AAG TGC AAA TTC TTA CAC CAG GAG AGG ATG GAT GTT TGC 432
Val Pro Asp Lys Cys Lys Phe Leu His Gln Glu Arg Met Asp Val Cys
130 135 140

324868.1

324868.1

(2) INFORMATION FOR SEQ ID NO:4:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 751 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: protein
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

324868.1

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Lys	Ser	Thr	Asn	Leu	His	Asp	Tyr	Gly	Met	Leu	Leu	Pro	Cys	Gly	Ile
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		355					360					365			
His	Ala	His	Phe	Gln	Lys	Ala	Lys	Glu	Arg	Leu	Glu	Ala	Lys	His	Arg
	370					375					380				
Glu	Arg	Met	Ser	Gln	Val	Met	Arg	Glu	Trp	Glu	Glu	Ala	Glu	Arg	Gln
385					390					395					400
Ala	Lys	Asn	Leu	Pro	Lys	Ala	Asp	Lys	Lys	Ala	Val	Ile	Gln	His	Phe
				405					410					415	

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```
(2) INFORMATION FOR SEQ ID NO:5:
      (i) SEQUENCE CHARACTERISTICS:
            (A) LENGTH: 2310 base pairs
            (B) TYPE: nucleic acid
            (C) STRANDEDNESS: double
            (D) TOPOLOGY: linear
      (ii) MOLECULE TYPE: cDNA
      (iii) HYPOTHETICAL: NO
      (iv) ANTI-SENSE: NO
      (ix) FEATURE:
            (A) NAME/KEY: CDS
            (B) LOCATION: 1-2310
            (D) OTHER INFORMATION: /function= "coding region for APP770."
      (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
```

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Lys	Ser	Thr	Asn	Leu	His	Asp	Tyr	Gly	Met	Leu	Leu	Pro	Cys	Gly	Ile	
			165					170						175		
GAC	AAG	TTC	CGA	GGG	GTA	GAG	TTT	GTG	TGT	TGC	CCA	CTG	GCT	GAA	GAA	576
Asp	Lys	Phe	Arg	Gly	Val	Glu	Phe	Val	Cys	Cys	Pro	Leu	Ala	Glu	Glu	
			180					185					190			
AGT	GAC	AAT	GTG	GAT	TCT	GCT	GAT	GCG	GAG	GAG	GAT	GAC	TCG	GAT	GTC	624
Ser	Asp	Asn	Val	Asp	Ser	Ala	Asp	Ala	Glu	Glu	Asp	Asp	Ser	Asp	Val	
		195					200					205				
TGG	TGG	GGC	GGA	GCA	GAC	ACA	GAC	TAT	GCA	GAT	GGG	AGT	GAA	GAC	AAA	672
Trp	Trp	Gly	Gly	Ala	Asp	Thr	Asp	Tyr	Ala	Asp	Gly	Ser	Glu	Asp	Lys	
	210					215					220					
GTA	GTA	GAA	GTA	GCA	GAG	GAG	GAA	GAA	GTG	GCT	GAG	GTG	GAA	GAA	GAA	720
Val	Val	Glu	Val	Ala	Glu	Glu	Glu	Glu	Val	Ala	Glu	Val	Glu	Glu	Glu	
225				230					235						240	
GAA	GCC	GAT	GAT	GAC	GAG	GAC	GAT	GAG	GAT	GGT	GAT	GAG	GTA	GAG	GAA	768
Glu	Ala	Asp	Asp	Asp	Glu	Asp	Asp	Glu	Asp	Gly	Asp	Glu	Val	Glu	Glu	
				245					250					255		
GAG	GCT	GAG	GAA	CCC	TAC	GAA	GAA	GCC	ACA	GAG	AGA	ACC	ACC	AGC	ATT	816
Glu	Ala	Glu	Glu	Pro	Tyr	Glu	Glu	Ala	Thr	Glu	Arg	Thr	Thr	Ser	Ile	
			260					265					270			
GCC	ACC	ACC	ACC	ACC	ACC	ACC	ACA	GAG	TCT	GTG	GAA	GAG	GTG	GTT	CGA	864
Ala	Thr	Thr	Thr	Thr	Thr	Thr	Thr	Glu	Ser	Val	Glu	Glu	Val	Val	Arg	
		275					280					285				
GAG	GTG	TGC	TCT	GAA	CAA	GCC	GAG	ACG	GGG	CCG	TGC	CGA	GCA	ATG	ATC	912
Glu	Val	Cys	Ser	Glu	Gln	Ala	Glu	Thr	Gly	Pro	Cys	Arg	Ala	Met	Ile	
	290					295					300					
TCC	CGC	TGG	TAC	TTT	GAT	GTG	ACT	GAA	GGG	AAG	TGT	GCC	CCA	TTC	TTT	960
Ser	Arg	Trp	Tyr	Phe	Asp	Val	Thr	Glu	Gly	Lys	Cys	Ala	Pro	Phe	Phe	
305					310					315					320	
TAC	GGC	GGA	TGT	GGC	GGC	AAC	CGG	AAC	AAC	TTT	GAC	ACA	GAA	GAG	TAC	1008
Tyr	Gly	Gly	Cys	Gly	Gly	Asn	Arg	Asn	Asn	Phe	Asp	Thr	Glu	Glu	Tyr	
				325					330					335		
TGC	ATG	GCC	GTG	TGT	GGC	AGC	GCC	ATG	TCC	CAA	AGT	TTA	CTC	AAG	ACT	1056
Cys	Met	Ala	Val	Cys	Gly	Ser	Ala	Met	Ser	Gln	Ser	Leu	Leu	Lys	Thr	
			340					345					350			
ACC	CAG	GAA	CCT	CTT	GCC	CGA	GAT	CCT	GTT	AAA	CTT	CCT	ACA	ACA	GCA	1104
Thr	Gln	Glu	Pro	Leu	Ala	Arg	Asp	Pro	Val	Lys	Leu	Pro	Thr	Thr	Ala	
		355					360					365				
GCC	AGT	ACC	CCT	GAT	GCC	GTT	GAC	AAG	TAT	CTC	GAG	ACA	CCT	GGG	GAT	1152
Ala	Ser	Thr	Pro	Asp	Ala	Val	Asp	Lys	Tyr	Leu	Glu	Thr	Pro	Gly	Asp	
	370					375					380					
GAG	AAT	GAA	CAT	GCC	CAT	TTC	CAG	AAA	GCC	AAA	GAG	AGG	CTT	GAG	GCC	1200
Glu	Asn	Glu	His	Ala	His	Phe	Gln	Lys	Ala	Lys	Glu	Arg	Leu	Glu	Ala	
385					390					395					400	

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(2) INFORMATION FOR SEQ ID NO:6:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 770 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: protein
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

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Val Met Leu Lys Lys Lys Gln Tyr Thr Ser Ile His His Gly Val Val
725 730 735

Glu Val Asp Ala Ala Val Thr Pro Glu Glu Arg His Leu Ser Lys Met
740 745 750

Gln Gln Asn Gly Tyr Glu Asn Pro Thr Tyr Lys Phe Phe Glu Gln Met
755 760 765

Gln Asn
770

- (2) INFORMATION FOR SEQ ID NO:7:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: DNA
 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CCGATGATGA CGAGGACGAT

20

- (2) INFORMATION FOR SEQ ID NO:8:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: DNA
 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TGAACACGTG ACGAGGCCGA

20

- (2) INFORMATION FOR SEQ ID NO:9:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 5 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: peptide
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Phe Arg Val Gly Ser
5

- (2) INFORMATION FOR SEQ ID NO:10:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 8 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: peptide
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Asp Ala Glu Phe Arg Gly Gly Cys
5